

**UNIVERSIDADE ESTADUAL DE PONTA GROSSA
PRÓ-REITORIA DE PESQUISA E PÓS-GRADUAÇÃO
PROGRAMA DE PÓS-GRADUAÇÃO EM ODONTOLOGIA - DOUTORADO
ÁREA DE CONCENTRAÇÃO: CLÍNICA INTEGRADA**

MIGUEL ANGEL MUÑOZ PÉREZ

**EFEITO DE DIFERENTES PROTOCOLOS DE APLICAÇÃO DE ADESIVOS
UNIVERSAIS NAS PROPRIEDADES E LONGEVIDADE DA UNIÃO À DENTINA**

PONTA GROSSA

2014

**UNIVERSIDADE ESTADUAL DE PONTA GROSSA
PRÓ-REITORIA DE PESQUISA E PÓS-GRADUAÇÃO
PROGRAMA DE PÓS-GRADUAÇÃO EM ODONTOLOGIA - DOUTORADO
ÁREA DE CONCENTRAÇÃO: CLÍNICA INTEGRADA**

MIGUEL ANGEL MUÑOZ PÉREZ

**EFEITO DE DIFERENTES PROTOCOLOS DE APLICAÇÃO DE ADESIVOS
UNIVERSAIS NAS PROPRIEDADES E LONGEVIDADE DA UNIÃO À DENTINA**

Tese apresentada para obtenção do título doutor em Odontologia na Universidade Estadual de Ponta Grossa, no Curso de Doutorado em Odontologia - Área de Concentração: Clínica Integrada. Linha de Pesquisa: Propriedades Físico-químicas e Biológicas dos Materiais Odontológicos

Orientadora: Prof^a. Dr^a. Nara Hellen Campanha Bombarda

Co-orientador: Prof. Dr. Alessandro Dourado Loguercio

PONTA GROSSA

2014

Ficha Catalográfica
Elaborada pelo Setor de Tratamento da Informação BICEN/UEPG

Muñoz Pérez, Miguel Angel
M967e Efeito de diferentes protocolos de aplicação de adesivos universais nas propriedades e longevidade da união à dentina/ Miguel Angel Muñoz Pérez. Ponta Grossa, 2014.
113f.

Tese (Doutorado em Odontologia - Área de Concentração: Clínica Integrada), Universidade Estadual de Ponta Grossa.
Orientadora: Prof^a Dr^a Nara Hellen Campanha.

1.Dentina. 2.Sistemas adesivos. 3.Adesivos convencionais. 4.Adesivos autocondicionantes. 5.Resistência de união. I.Campanha, Nara Hellen. II. Universidade Estadual de Ponta Grossa. Doutorado em Odontologia. III. T.

CDD: 617.67

MIGUEL ANGEL MUÑOZ PÉREZ

**EFEITO DE DIFERENTES PROTOCOLOS DE APLICAÇÃO DE ADESIVOS
UNIVERSAIS NAS PROPRIEDADES E LONGEVIDADE DA UNIÃO À
DENTINA**

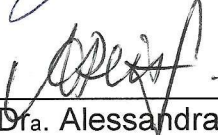
Tese apresentada para a obtenção do título de Doutor em Odontologia na
Universidade Estadual de Ponta Grossa, no curso de Doutorado em
Odontologia

Área de concentração: Clínica Integrada.


Ponta Grossa, 7 de Fevereiro de 2014.



Prof.a. Dra. Nara Hellen Campanha Bombarda
Universidade Estadual de Ponta Grossa – UEPG



Prof.a. Dra. Alessandra Reis
Universidade Estadual de Ponta Grossa – UEPG



Prof.a. Dra. César Augusto Galvão Arrais
Universidade Estadual de Ponta Grossa – UEPG



Prof.a. Dra. Monica Yamauti
Universidade Federal de Ceará – UFC



Prof. Dr. Carlos Francci
Universidade de São Paulo – USP

DADOS CURRICULARES

Miguel Angel Muñoz Pérez

NASCIMENTO 17.04.1977	Arica – Chile
FILIAÇÃO	Amanda Doris Pérez Delgado Miguel Angel Muñoz Ibarra
1997 – 2003	Curso de Graduação em Odontologia da Universidad de Valparaíso. Valparaíso – Chile.
2008 – 2010	Curso de Pós-graduação em Odontologia, Universidade Estadual de Ponta Grossa – UEPG, nível de Mestrado em Odontologia - Área de Concentração em Clínica Integrada.
2010 – 2014	Curso de Pós-graduação em Odontologia, Universidade Estadual de Ponta Grossa – UEPG, nível de Doutorado em Odontologia - Área de Concentração em Clínica Integrada.

DEDICATÓRIA

A meus pais Amanda Pérez e Miguel Muñoz,

pela paciência na realização de meus sonhos,

por serem meu exemplo de vida baseado

em princípios, esforço e respeito pelos outros. Com

vocês aprendi a enfrentar os problemas e os

desafios com coragem, dignidade e sabedoria.

A minha irmã Sandra, pelo carinho, paciência e

companheirismo nas horas difíceis e a cumplicidade

nas horas felizes

A minha noiva Issis , por ser parte fundamental da minha vida,

energia no meu dia a dia,

inspiração inesgotável.

Otimismo em um futuro feliz

Te amo!!

A Deus

AGRADECIMENTOS ESPECIAIS

A minha orientadora Prof^a. Dr^a. Nara Hellen Campanha Bombarda por me acolher como orientado, pelo apoio e compreensão no caminho que escolhi e a grandeza ao me permitir percorrê-lo. Muito obrigado pela confiança depositada e pelo carinho oferecido junto a sua maravilhosa família.

Aos **Prof^s. Dr^s Alessandro Loguercio e Alessandra Reis**. Mestres queridos, obrigado pelos conhecimentos, paciência, confiança, oportunidades e incentivo que sempre me ofereceram. Obrigado por me permitir compartilhar com sua família momentos incríveis que levarei como grandes presentes. Obrigado pela amizade, risadas e os felizes momentos que compartilhamos. Obrigado por cuidar sempre da Issis, o amor da minha vida. Minha casa será sempre a sua.

Ao Prof. Dr. João Carlos Gomes, pela oportunidade de participar no curso de Mestrado e Doutorado, além de seu apoio incondicional em tudo o que eu necessitei. Obrigado por me ensinar sobre liderança e por acreditar em atividades como o CIAO, que nos permitiram viver momentos inspiradores.

A Prof^a. Dr^a. Osnara Maria Mongruel Gomes, por ser um exemplo de pessoa, compreensiva, justa e positiva, realmente nossa Mãe no Brasil. Muito obrigado pela ajuda em todo momento, pela paciência e exemplo de competência e profissionalismo.

AGRADECIMENTOS

Gostaria de tornar pública a minha gratidão ao **Brasil e toda a sociedade brasileira** por ter me acolhido de braços abertos.

À **Universidade Estadual de Ponta Grossa – UEPG**, na pessoa do Reitor Carlos Luciano Sant’Ana Vargas pela oportunidade a mim outorgada.

Ao **Programa de Pós-Graduação Stricto Sensu em Odontologia**, na pessoa de sua coordenadora **Prof^a. Dr^a. Osnara Maria Mongruel Gomes**, pelo seu comprometimento com a qualificação de seus discentes o que tem permitido meu crescimento profissional e pessoal. Espero que meu esforço conte como parte da minha dívida, eternizando aqui a minha passagem por esta instituição.

A **CAPES e Fundação Araucária** pelo financiamento de uma bolsa de estudo e pesquisa.

À **Universidade de São Paulo – USP** e o **Departamento de Biomateriais e Biologia Oral** por me permitir trabalhar em seus laboratórios.

À **University of Minnesota**, por me permitir realizar um estágio de excelência.

À **Faculdade de Odontologia da Universidad de Valparaíso**, na pessoa de seu Diretor Prof. Dr. Gaston Zamora, pela confiança e mim depositada.

A **Prof^a. Dr^a. Rosa Helena Miranda Grande**, pela dedicação brindada e constantes conselhos em num momento intenso e difícil de meu doutorado.

Ao **Prof. Dr. Jorge Perdigão**, por ter oportunizado, na *Univesity of Minnesota*, que eu e a Iسس tivéssemos uma grande experiência profissional e pessoal plena de ensinamentos que levaremos eternamente, como parte de nossas melhores lembranças.

Ao **Prof. Jaime Sarmiento**, Chefe de Operatória Dental da Universidade de Valparaíso, pela confiança em mim.

A **meu amigo Luis Alfonso**, por ser um grande parceiro neste caminho e ter se transformado num irmão.

A **meu amigo Eugenio**, “el che”, uma grande pessoa, amigo incansável, sincero como ninguém. Obrigado por me receber em São Paulo e me ajudar a suportar esse momento.

A **Viviane Hass**, obrigado pelo apoio, amizade em momentos fundamentais, obrigado por querer tão bem à Issis e cuidar sempre de nós.

A **Anna Luiza**, pela amizade, tempo dedicado e as intenções de sempre ajudar.

A meus amigos, **Che Patrício, Davila, Montenegro, Camilo, Millan, Thays, Zé, Fabi Siqueira, Adelaida, Joha, Sibelli, Felipe, Fabi Coppla, Juliana, Rosana, Marcinha, Bruna, Paola, Ana K, Reila, Ligia e a Ritinha**, por todos os momentos vividos.

A **Alex, Yileng, Eloisa, Michelle, Giovanna, Baggio, Daniel, Stella, Yasmine, Carlos, Sindianara, Dalton**. Lembrarei sempre dos bons momentos.

A **meus amigos Flavia e Sergio**, muito obrigado por sua amizade, trabalho e disposição para oferecer ajuda sempre que eu precisei.

A **Andrea e Milko**, por sua amizade e inesquecível apoio

Aos **meus amigos brasileiros**, por me receberem e por terem sempre disposição para ajudar.

Ao **Prof. Paulo Vitor**, pela paciência em todo momento, por ter sempre disposição para explicar, ensinar e procurar soluções. Um grande exemplo de docência e pessoa.

A meus amigos **Milton, Nilson, Simone e Barbara** pelo apoio técnico nos laboratórios, mas fundamentalmente pelo positivismo em todo momento.

A todos **os professores do Programa de pós-graduação em Odontologia da UEPG** que transmitiram seus conhecimentos durante o curso de **Doutorado**.

Aos **funcionários do Programa de pós-graduação em Odontologia da UEPG**, especialmente à Morgana e Luiz, pela competência e amizade.

A meu amigo **Leo**, pelo positivismo em todo momento, um grande exemplo de pessoa.

MUÑOZ MA. Efeito de diferentes protocolos de aplicação de adesivos universais nas propriedades e longevidade da união à dentina. [Tese – Doutorado em Clínica Integrada]. Ponta Grossa: Universidade Estadual de Ponta Grossa; 2013.

RESUMO

O objetivo do presente trabalho foi avaliar *in vitro* as propriedades e longevidade da união à dentina dos novos sistemas adesivos universais aplicados segundo as estratégias adesivas convencional (CON) e autocondicionante (AC). Os objetivos específicos para cada um dos três estudos foram: avaliar a resistência de união à microtração (RU), a nanoinfiltração (NI) e o grau de conversão (GC) no tempo imediato - 24 h - (Experimento 1) e seis meses (Experimento 2) dos sistemas adesivos universais para as estratégias CON e AC; e o efeito de uma camada hidrofóbica adicional (Experimento 3, no tempo imediato) sobre essas mesmas propriedades. Cento e quarenta terceiros molares foram planejados na face oclusal até se obter dentina e alocados aleatoriamente em 28 grupos (n=5). Nos Experimentos 1 e 2, 40 dentes foram divididos em oito grupos de acordo com o adesivo (Peak Universal [Pk], Scotchbond Universal [Sc], Allbond Universal [Al]) e estratégia adesiva (CON e AC) tendo como controle, o sistema adesivo convencional Adper Single Bond 2 (SB) e o autocondicionante Clearfil SE Bond (CSE). Para o Experimento 3, 60 dentes foram divididos em 12 grupos de acordo com o adesivo (Sc, Al, G-Bond Plus [Gbp]), estratégia adesiva (CON e AC) e com a presença ou não de camada hidrofóbica (Heliobond, HE). Após aplicação das variáveis adesivas e do procedimento restaurador, os dentes foram armazenados em água destilada (37°C/24 h) e cortados para obter palitos de dentina-resina (0,8 mm² de secção transversal) sendo submetidos aos teste de RU e NI nos tempos imediato (Experimentos 1 e 3) e também seis meses (Experimento 2); e análise de GC no tempo imediato. Os dados do Experimento 1 foram submetidos à análise de variância de um fator (Estratégia Adesiva) e os dos Experimentos 2 e 3, à de dois fatores (Estratégia adesiva versus Tempos de análise; e Estratégia adesiva versus Camada hidrofóbica), respectivamente. As análises foram complementadas pelo teste de Tukey ($\alpha=0,05$). Os resultados de RU obtidos no Experimento 1 demonstraram que apenas PkCon e PkAc foram similares aos respectivos grupos controle ($p>0,05$), que os outros materiais universais resultaram em médias de RU inferiores aos seus controles CON e AC ($p<0,05$) e que AlAc apresentou as médias mais baixas de RU ($p<0,05$). Para NI, ScCon, ScAc, AlAc, e AlCon apresentaram baixos níveis de NI similares aos grupos controle ($p<0,05$). Para GC, unicamente ScAc demonstrou baixo GC, quando comparado com os outros materiais ($p<0,05$). Segundo os resultados do Experimento 2, os sistemas Pk e AlCon apresentaram diminuição da RU ($p<0,05$) e apenas Pk teve degradação significativa (NI) após seis meses ($p<0,05$). De acordo com o Experimento 3, Al e Gbp obtiveram maiores resultados de RU na estratégia CON ($p<0,05$); o uso de HE aumentou a RU unicamente na estratégia AC ($p<0,05$); baixa NI foi observada somente em Sc e Al na estratégia CON+HE e em Gbp em AC+HE; Sc e Gbp apresentaram alto GC quando usados em estratégia CON, que foi aumentado com a aplicação de HE ($p<0,05$). O GC de Al foi similar em todas as condições. A partir dos três experimentos que compõem o presente trabalho, pôde-se concluir que: 1) os adesivos universais apresentaram um comportamento

dependente do material em ambas estratégias (CON e AC) e foram inferiores, pelo menos em uma das propriedades avaliadas (RU, NI ou GC), quando comparados aos adesivos controle (CSE e SB); 2) Em geral, após seis meses os adesivos AC mantêm mais estáveis os valores de RU e NI, o que parece estar relacionado principalmente à sua capacidade de união química ao substrato dental; 3) A utilização de uma camada hidrofóbica na estratégia AC pôde aumentar a RU e o GC de adesivos universais e a redução da NI foi mais dependente da composição do adesivo do que da estratégia adesiva.

Palavras-chave: dentina; sistemas adesivos; adesivos convencionais; adesivos autocondicionantes; resistência de união; grau de conversão; nanoinfiltração.

MUÑOZ MA. Effect of different universal adhesive application protocols on the properties and longevity of bond to dentin [Thesis – Doctorate in Integrated Clinic]. Ponta Grossa: State University of Ponta Grossa; 2013.

ABSTRACT

The aim of the present study was to make an *in vitro* evaluation of the properties and stability of the bond to dentin of new universal adhesive systems applied according to the etch-and-rinse (ER) and self-etching (SE) adhesive strategies. The specific objectives for each of the three studies were: To evaluate the microtensile bond strength (RU), nanoleakage (NI) and degree of conversion (DC) in the time intervals of 24 h (Experiment 1) and six months (Experiment 2) of the universal adhesive systems applied in accordance with the manufacturers' recommendations for strategies ER and SE; and the effect of an additional hydrophobic layer (Experiment 3, in the immediate time interval) on these same properties. The occlusal surfaces of 140 third molars were flattened until dentin was obtained, and then the teeth were randomly allocated to 28 groups (n=5). In Experiments 1 and 2, 40 teeth were divided into eight groups according to the adhesive used (Peak Universal [Pk], Scotchbond Universal [Sc], Allbond Universal [Al]) and adhesive strategy (ER and SE), with the ER adhesive system Adper Single Bond 2 (SB) and self-etching Clearfil SE Bond (CSE) used as controls. For Experiment 3, 60 teeth were divided into 12 groups according to the adhesive (Sc, Al, G-Bond Plus [Gbp]), adhesive strategy (ER and SE) and with or without hydrophobic layer (Heliobond, HE). After application of the variables *adhesives* and *restorative procedure*, the teeth were stored in distilled water (37°C/24 h) and cut to obtain dentin-resin sticks (0.8 mm² cross-section) which were submitted to the RU and NI tests in the following time intervals: immediate (Experiments 1 and 3) and six months (Experiment 2); and analysis of DC in the immediate time interval. The data of Experiment 1 were submitted to one-way analysis of variance (Adhesive Strategy) and those of Experiments 2 and 3, to two-way analysis of variance (Adhesive strategy vs. Time intervals; and Adhesive strategy vs. Hydrophobic layer) respectively. The analyses were complemented by the Tukey Test ($\alpha=0.05$). The RU results obtained in Experiment 1 demonstrated that only PkEr and PkSe were similar to the respective control groups ($p>0.05$); that the other universal materials resulted in lower RU means than those of their controls ER and SE ($p<0.05$) and that AlSe presented the lowest mean RU values ($p<0.05$). For NI, the Groups ScEr, ScSe, AlSe, and AlEr presented low levels of NI similar to those of the control groups ($p<0.05$). For DC, only Group ScSe demonstrated low DC when compared with the other materials ($p<0.05$). According to the results of Experiment 2, the systems Pk and AlEr presented a reduction in RU ($p<0.05$) and only Pk underwent significant degradation (NI) after six months ($p<0.05$). According to Experiment 3, Al and Gbp obtained higher RU results with the ER strategy ($p<0.05$); the use of HE increased the RU only in strategy SE ($p<0.05$); a low level of NI was observed only in Groups Sc and Al with the CON+HE strategy and in Group Gbp with SE+HE; Groups Sc and Gbp presented a high DC when used with the ER strategy, which was increased with the application of HE ($p<0.05$). The DC if Al was similar under all the conditions. From the three experiments of which this study was composed, it could be concluded that: 1) the universal adhesives presented a

material-dependent behavior in both strategies (ER and SE) and were inferior, in at least one of the properties evaluated (RU, NI or DC), when compared with the control adhesives (CSE and SB); 2) In general, after six months the adhesives SE maintained the most stable RU and NI values, which appears to be related mainly to their capacity to form a chemical bond to dental substrate; 3) The use of a hydrophobic layer in strategy SE was able to increase the RU and DC of universal adhesives and the reduction in NI was more dependent on the composition of the adhesive than on the adhesive strategy.

Key Words: dentin; adhesive systems; etch-and-rinse; self-etch; bond strength; degree of conversion; nanoleakage.

LISTA DE ILUSTRAÇÕES

Material e método

- Figura 1** Preparo da superfície de dentina. (A) dente hígido; (B) superfície oclusal planificada e em dentina; desgaste de todas as bordas periféricas de esmalte..... 30
- Figura 2** Estratégia adesiva convencional. (A) condicionamento ácido; (B) lavagem e secagem; (C) Aplicação do adesivo; (D) secagem para evaporação do solvente; (E) fotoativação. Todos os procedimentos realizados segundo as indicações do fabricante, os quais estão discriminados no quadro 2..... 31
- Figura 3** Estratégia adesiva autocondicionante (dispensa a aplicação e a lavagem do ácido fosfórico). (A) Aplicação do adesivo; (B) secagem para evaporação do solvente; (C) fotoativação. Todos os procedimentos realizados segundo as indicações do fabricante..... 31
- Figura 4** Estratégia adesiva convencional com aplicação de camada hidrofóbica. (A) condicionamento ácido; (B) lavagem e secagem; (C) Aplicação do adesivo; (D) secagem para evaporação do solvente; (E) aplicação de uma camada de resina hidrofóbica; (F) fotoativação. Vide Quadro 3..... 32
- Figura 5** Estratégia adesiva autocondicionante, a qual dispensa o condicionamento com ácido fosfórico, com aplicação de camada hidrofóbica; (A) aplicação do adesivo; (B) secagem para evaporação do solvente; (C) aplicação de camada de resina hidrofóbica; (D) fotoativação. Vide Quadro 3..... 33
- Figura 6** Restauração dos dentes após aplicação das variáveis adesivas. (A) Colocação da resina composta; (B) fotoativação do primeiro incremento; (C) Colocação do incremento final; (D) fotoativação do segundo incremento..... 36
- Figura 7** Sequência de corte para a obtenção dos palitos. (A) superfície oclusal das coroas de resina compostas pintadas com caneta marcadora permanente de cores diferentes para identificação dos palitos de cada grupo. (B) dente fixado com cera pegajosa ao dispositivo da máquina de corte montada com um disco diamantado; (C) duas sequências perpendiculares de corte, uma no eixo x e outra no eixo y, permitiram a obtenção dos palitos para o teste de resistência de união; (D) em eppendorfs, os palitos de cada dente foram armazenados em água destilada por 24 h..... 37

Figura 8	Sequência para o teste de microtração. (A) a seta azul indica a interface adesiva entre dente e restauração, numa vista lateral do palito. (B) mensuração da largura e da espessura do mesmo palito, com paquímetro digital, para o cálculo da área da interface adesiva. (C) cola de cianocrilato em gel (que permite controle da aplicação). (D) aplicação da cola nas extremidades do palito, protegendo a interface com a pinça; (E) posicionamento do palito nas garras de microtração deixando a interface adesiva no espaço central da garra; (F) adaptação das garras na máquina de ensaios universais após a polimerização da cola.....	38
Figura 9	Tipos de fratura. Em (A) coesiva de resina composta e (B) coesiva de dentina; (C) adesiva na camada híbrida; (D) adesiva na camada adesiva; E, mista.....	39
Figura 10	<i>Stub</i> com palitos lixados previamente à leitura no micro-Raman. Isto facilita a localização da camada híbrida.....	41
Figura 11	Espectros representativos obtidos do sistema adesivo Peak Bond Universal não polimerizado, e após sua fotoativação na camada híbrida in situ em sua versão ER e SE. As setas indicam os picos de referencia (1609 cm ⁻¹) e reativo (1639 cm ⁻¹).....	41
Figura 12	Preparação de amostras para NI: (A) palitos imersos em uma solução aquosa de nitrato de prata (pH=7,0); (B) armazenados por 24 h sob total escuridão; (C) após lavagem, imersos em solução reveladora por 8 h sob luz fluorescente; (D) palitos revelados, montados em <i>stubs</i> de alumínio para facilitar o polimento uniforme; (E) palitos dessecados em sílica por 24 h; (F) palitos metalizados para a análise de MEV.....	42
Figura 13	Análise de pH. Vista do eletrodo totalmente submerso na solução para a leitura do pH.....	43
Quadro 1	Materiais adesivos, composição e fabricante.....	29
Quadro 2	Sistemas adesivos e estratégias de aplicação de acordo com as instruções do fabricante para o experimento 1 e 2.....	34
Quadro 3	Materiais adesivos e estratégias de aplicação dos sistemas adesivos utilizados no experimento 3.....	35

Capítulo 1

- Figura 1** Representative backscatter SEM images of the resin-dentin adhesive interfaces of each experimental groups (Control etch-and-rinse group = Adper Single Bond 2 and control self-etch group = Clearfil SE Bond). The amount of nanoleakage was lower and practically occurred within the hybrid layer for Scotchbond Universal (C, G); All Bond Universal (D, H) and controls (*white arrows*) (A, E). For Peak Universal Adhesive System, the amount of nanoleakage was higher than the other materials; with most silver nitrate uptake occurring throughout the entire thickness of the HL (*white arrows*) and in the AL, forming the so-called “water trees”(white hands) (B, F)..... 50

Capítulo 2

- Figura 1** Representative backscatter SEM images of the resin-dentin adhesive interfaces of each experimental group for immediate and 6 month times..... 75

Capítulo 3

- Figura 1** Representative backscattered SEM images of the resin–dentín adhesive interfaces of each universal adhesive system (Scotchbond Universal Adhesive [SBU]; All Bond Universal [ABU] and G-Bond Plus [GBP]), on different bonding strategies (etch-and-rinse [ER] and self-etch [SE]) with or without an additional hydrophobic resin bonding layer [HE]. For SBU and ABU, lower NL was seen in the ER strategy when HE was used (B and F) compared with their controls (A and E). However, for the SE strategy, nanoleakage occurred similarly within the hybrid layer for both ABU and SBU using or not HE. (*white arrows*) (C, D, G and H). For GBP showed higher amount of nanoleakage in ER strategy on the hybrid layer (*white arrows*) (I), when compared with the SBU and ABU, and most silver nitrate uptake occurring throughout the entire thickness of the HL (*white arrows*) and in the AL, forming the so-called “water trees (*white hands*) (J), and the SE strategy without HE (K). Only in the SE strategy with HE, the nanoleakage was lower and similar to SBU and ABU (D, H and L)..... 83

LISTA DE TABELAS

Capítulo 1

Tabela 1	Adhesive materials (batch number), composition and application mode of the adhesive systems used.....	47
Tabela 2	Microtensile bond strength (μ TBS), nanoleakage (NL) and degree of conversion (DC) values (means \pm standard deviations) of the different experimental groups.....	49
Tabela 3	Number and percentage of specimens (%) according to fracture mode and the premature failures of all experimental group.....	49
Tabela 4	The pH values (means \pm standard deviations) of each adhesive system.....	50

Capítulo 2

Tabela 1	Adhesive materials (batch number), composition and application mode of the adhesive systems used.....	76
Tabela 2	Number specimens (%) according to fracture mode and the premature failures of all experimental groups.....	77
Tabela 3	Microtensile bond strength (μ TBS) and values (means \pm standard deviations) of the different experimental groups.....	78
Tabela 4	Nanoleakage (NL) values (means \pm standard deviations) of the different experimental groups.....	78

Capítulo 3

Tabela 1	Adhesive materials (batch number), composition and application mode of the adhesive systems used.....	84
Tabela 2	Number (%) of specimens according to fracture pattern mode for the experimental groups.....	85
Tabela 3	Microtensile bond strength (μ TBS) values (means \pm standard deviations) of the different experimental groups.....	85
Tabela 4	Nanoleakage (NL) values (means \pm standard deviations) of the different experimental groups.....	85
Tabela 5	Degree of conversion (DC) values (means \pm standard deviations) of the different experimental groups.....	86

LISTA DE SIGLAS E ABREVIATURAS

A	Fratura adesiva
AC ou Ac	Estratégia Adesiva Autocondicionante
A/M	Fratura adesiva/mista
AI	Allbond Universal
Bis-GMA	Bisfenol Glicidil dimetacrilato
C	Fraturas coesivas
COEP	Comitê de Ética em Pesquisa
CON ou Con	Estratégia Adesiva Convencional
Cp	Corpo de prova
CSE	Clearfil SE Bond
FP	Número de falhas prematuras
Gbp	G-Bond Plus
GC	Grau de conversão
HE	Heliobond
HEMA	2-Hidroxi-etil-metacrilato
I	Integral
LED	Diodo emissor de luz, pela frase em inglês <i>Light Emitting Diode</i>
MDP	10-Metacrilóiloxidecil dihidrogênio fosfato
MEV	Microscopia Eletrônica de Varredura
n°	Número
NI	Nanoinfiltração
np	Não polimerizado

p	Polimerizado
PAC	Copolímero de ácido polialquenoico
pH	Potencial hidrogeniônico
Pk	Peak Universal
RU	Resistência de União à microtração
SB	Adper Single Bond 2
Sc	Scotchbond Universal
SiC	Carbeto de silício
TEGDMA	Trietilenoglicoldimetacrilato

Capítulos

ABU	Allbond Universal
DC	Degree of conversion
Er	Etch-and-rinse
GBP	G-Bond Plus
NL	Nanoleakage
PF	Premature failures
R	Ratio of aliphatic and aromatic peak areas
SBU	Scotchbond Universal
Se	Self-etch
SEM	Scanning electron microscope
μ TBS	Microtensile bond strengths

LISTA DE SÍMBOLOS E UNIDADES

#	Número
%	Porcentagem
=	Igual
<	Menor que
>	Maior que
±	Mais ou menos
≈	Aproximadamente
μm	Micrometro
cm ⁻¹	Centímetro(s) recíproco(s) ou centímetro(s) inverso(s)
s	Segundo(s)
min	Minuto(s)
MPa	Mega Pascal
mm	Milímetro(s)
mm ²	Milímetro(s) quadrado(s)
mm/min	Milímetro(s) por minuto
mW	MiliWatts
mW/cm ²	MiliWatts por centímetro(s) quadrado
°C	Grau(s) Celsius
n	Número de corpos de prova
N	Newton
nm	Nanômetro
X	Número de vezes
rpm	Rotações por minuto

h	Hora(s)
α	Nível de significância
p	Valor de probabilidade

SUMÁRIO

1.	INTRODUÇÃO	23
2.	PROPOSIÇÃO	28
3.	MATERIAL E MÉTODOS	29
4.	ARTIGOS	44
4.1	CAPÍTULO 1.....	44
4.2	CAPÍTULO 2.....	53
4.3	CAPÍTULO 3.....	80
5.	DISCUSSÃO	89
6.	CONCLUSÕES	97
	REFERÊNCIAS	98
	ANEXO	110

1. INTRODUÇÃO

Vários fatores podem afetar a longevidade de restaurações, entretanto, a cárie adjacente às margens ainda é uma das maiores causas de sua substituição (Demarco et al.¹ 2012). Esta falha pode estar relacionada à falta de selamento da interface dente/restauração, o que faz com que haja falta de resistência desta interface aos agentes internos - enzimas proteolíticas - e externos - fluidos e microorganismos (Spencer et al.² 2010; Perdigão et al.³ 2013; Tjaderhane et al.⁴ 2013). Esta interface adesiva dente/restauração é produzida pela interação de um sistema adesivo sobre os tecidos dentários duros após a substituição de minerais removidos desses tecidos por monômeros resinosos, formando um polímero interligado micro-mecanicamente com os substratos dentais (Nakabayashi et al.⁵ 1982).

Os atuais sistemas adesivos presentes no mercado podem ser classificados segundo sua estratégia de aplicação clínica em: convencional (CON) que inclui condicionamento ácido prévio (*etch-and-rinse*, no idioma Inglês); ou autocondicionante (AC) (*self-etch*, em inglês). Ambas são subdivididas segundo os passos de aplicação em versões de três (unicamente em CON), dois e um passo, este último unicamente em AC (Van Meerbeek et al.⁶ 1998; De Munck et al.⁷ 2005). As versões simplificadas de ambas categorias foram possíveis através da inclusão de monômeros hidrofílicos e do aumento da quantidade de solventes, para torná-los compatíveis com a umidade inerente da dentina (Van Meerbeek et al.⁸ 2003; Yiu et al.⁹ 2005).

Na estratégia CON, um gel de ácido fosfórico (20-40%) é aplicado no substrato dentário para remoção da *smear layer* e a exposição das fibrilas de colágeno na dentina. No esmalte, este ácido promove o aumento da área de superfície e da energia superficial. A seguir é aplicado um *primer* que é a mistura de solventes (acetona, etanol e/ou água) e monômeros resinosos hidrofílicos (por exemplo, HEMA) que infiltram o substrato para torná-lo adequado para receber uma mistura de monômeros resinosos relativamente mais hidrofóbicos denominado de *bond*. Estas etapas executadas separadamente (ácido, *primer* e *bond*) exemplificam os sistemas adesivos CON de 3-passos. Da mistura do *primer* e do *bond* numa

mesma solução nasceram as versões simplificadas de 2-passos (Van Meerbeek et al.⁶ 1998; De Munck et al.⁷ 2005; Van Meerbeek et al.¹⁰ 1992).

A principal desvantagem dos sistemas CON, principalmente nas versões de 2-passos, é o risco de as fibrilas colágenas colapsarem durante o processo de secagem da dentina desmineralizada, o que leva a uma diminuição da resistência de união (Spencer, Swafford¹¹ 1999; Tay et al.¹² 1996). O colapso das fibrilas colágenas é evitado mantendo-se a dentina desmineralizada úmida, tarefa difícil de ser realizada clinicamente. Isto porque a umidade adequada depende do tipo de solvente utilizado no material (Reis et al.¹³ 2003) e da interpretação do clínico das instruções do fabricante. A infiltração incompleta das fibrilas de colágeno (Sano et al.¹⁴ 1994) e a necessidade de protegê-las contra os mecanismos de degradação presentes no meio oral (Burrow et al.¹⁵ 1996; Hashimoto et al.¹⁶ 2000) levaram ao desenvolvimento da segunda categoria de sistemas adesivos denominados AC.

Na estratégia adesiva AC (de 1- ou 2-passos) não se indica aplicar previamente o ácido fosfórico nos substratos dentários. Nesta estratégia, utiliza-se uma solução que mistura solventes com monômeros ácidos não laváveis acontecendo simultaneamente desmineralização e infiltração da dentina (De Munck et al.⁷ 2005; Miyazaki et al.¹⁷ 2002), assim os cristais de hidroxiapatita dissolvidos e a *smear layer* residual são incorporados na camada híbrida (De Munck et al.⁷ 2005; Van Meerbeek et al.¹⁸ 2003). O procedimento de união com os adesivos autocondicionantes pode ser realizado em dois ou um passos, dependendo se o *primer*/condicionador encontra-se separado do *bond* (2-passos) ou combinado com ele (1-passo). Exceto para os sistemas AC com pH muito baixo (Carvalho et al.¹⁹ 2005; Wang, Spencer²⁰ 2005), toda a extensão em profundidade da dentina desmineralizada é impregnada por monômeros de resina.

Assim, uma das vantagens desta técnica é que ela não é sensível à umidade, já que o adesivo é aplicado em substrato seco. Esta pode ser a razão pela qual os sistemas AC não estão associados com as características de sensibilidade da técnica de união à dentina condicionada úmida (Reis et al.¹³ 2003; Tay et al.²¹ 1998; Perdigão²² 2010). Por outro lado, uma desvantagem evidente do protocolo AC é a redução da eficácia da união ao esmalte (Kanemura et al.²³ 1999; Pashley, Tay²⁴ 2001), devido à menor capacidade de remoção dos minerais desse tecido.

Conseqüentemente, o aumento na área de superfície do esmalte obtido com adesivos AC é menor do que o atingido com o ácido fosfórico, sendo esta capacidade dependente do pH do adesivo AC (Pashley, Tay²⁴ 2001).

Assim, alguns estudos têm investigado a utilização de ácido fosfórico antes da aplicação de adesivos AC (Van Meerbeek et al.¹⁸ 2003; Rotta et al.²⁵ 2007; Frankenberger et al.²⁶ 2008), observando uma melhora no comportamento desses adesivos. No entanto, este procedimento tem-se mostrado inadequado na dentina (Van Landuyt et al.²⁷ 2006; Van Landuyt et al.²⁸ 2006; Hanabusa et al.²⁹ 2012) situação que poderia acontecer acidentalmente durante o condicionamento de esmalte por escoamento de ácido fosfórico de baixa viscosidade. O efeito do condicionamento intencional da dentina com ácido fosfórico prévio à aplicação dos adesivos AC tem sido estudado (Van Landuyt et al.²⁷ 2006; Hanabusa et al.²⁹ 2012; Erhardt et al.³⁰ 2008; Proenca et al.³¹ 2007; Taschner et al.³² 2012), com resultados controversos e material-dependentes.

Considerando as diferenças no julgamento profissional a respeito da seleção da estratégia adesiva e número de passos em diferentes situações clínicas, alguns fabricantes lançaram sistemas adesivos mais versáteis que incluem as opções convencional (de 2-passos) e autocondicionante (2- e 1-passo). Estes novos materiais são chamados de “Universais,” “Multimodo” ou “Multipropósito” (Hanabusa et al.²⁹ 2012; Perdigão et al.³³ 2012) mas ainda existe pouca informação acerca do comportamento desta nova classe de sistemas adesivos. Estes materiais, em geral, são adesivos simplificados na sua concepção e, portanto, presume-se com grande grau de hidrofobicidade, o que poderia ocasionar, em parte, os mesmos problemas de degradação da união dos adesivos anteriores.

Algumas pesquisas têm avaliado os mecanismos de envelhecimento envolvidos na degradação das interfaces de união (Liu et al.³⁴ 2011; Breschi et al.³⁵ 2008), porém este mecanismo não é claramente compreendido (Breschi et al.³⁵ 2008; Vaidyanathan, Vaidyanathan³⁶ 2009; Hashimoto³⁷ 2010). Acredita-se que a degradação se inicia com a extração da resina infiltrada na matriz dentinária através de nano-espaços cheios de água presentes na camada híbrida e pelo ataque enzimático das fibrilas colágenas expostas, levando à sua instabilidade (Spencer et al.² 2010; Sano³⁸ 2006).

Como relatado anteriormente, versões simplificadas foram possíveis através da inclusão de monômeros hidrofílicos, ácidos, e do aumento da quantidade de solventes (Van Meerbeek et al.⁸ 2003; Yiu et al.⁹ 2005). Porém, isto vai conduzir a uma maior quantidade de solvente residual retido na camada híbrida e camada de adesivo (Yiu et al.⁹ 2005), impedindo a formação de um polímero com reticulado elevado (Ye et al.³⁹ 2007; Loguercio et al.⁴⁰ 2009; Paul et al.⁴¹ 1999), diminuindo o grau de conversão (Cadenaro et al.⁴² 2005), reduzindo a resistência de união resina-dentina (Hass et al.⁴³ 2011; Reis et al.⁴⁴ 2010; Takahashi et al.⁴⁵ 2002; Cho, Dickens⁴⁶ 2004) e aumentando a permeabilidade da camada adesiva, após polimerização (Malacarne et al.⁴⁷ 2006; Ito et al.⁴⁸ 2005). Conseqüentemente, os polímeros serão mais susceptíveis à degradação ao longo do tempo (De Munck et al.⁷ 2005; Breschi et al.⁴⁹ 2008). Por outro lado, tem se demonstrado que a infiltração de monômeros dos sistemas adesivos CON é sempre menor do que a extensão da desmineralização da dentina (Wang, Spencer⁵⁰ 2003; Wang et al.⁵¹ 2006; Santini, Miletic⁵² 2008) e que isto também poderia acontecer com os sistemas adesivos AC (Carvalho et al.¹⁹ 2005; Wang, Spencer²⁰ 2005), resultando em zonas incompletamente infiltradas ao longo da parte inferior da camada híbrida que contêm fibrilas de colágenas expostas (Sano et al.⁵³ 1995) e nanoespaços com água denominados de nanoinfiltração (Sano et al.⁵³ 1995). Assim, o sinergismo entre a falta de infiltração dos monômeros e a instabilidade do polímero permitiria uma exposição contínua de fibrilas colágenas as quais ficariam susceptíveis à atividade de enzimas proteolíticas como as metaloproteinases (MMPs) (De Munck et al.⁵⁴ 2003; Nishitani et al.⁵⁵ 2006; Pashley et al.⁵⁶ 2004) e cisteína catepsina (Tersariol et al.⁵⁷ 2010; Nascimento et al.⁵⁸ 2011).

Reis e colaboradores (Reis et al.⁵⁹ 2013) indicaram diferentes alternativas clínicas que podem ser usadas para diminuir a degradação da interface de união à dentina, e assim, aumentar a longevidade das restaurações. Dentre as técnicas discutidas se inclui a aplicação de uma camada adicional de resina hidrofóbica sobre o adesivo simplificado (King et al.⁶⁰ 2005). Esta camada extra de resina tem como objetivo aumentar a espessura e uniformidade da interface adesiva, bem como reduzir o fluxo de fluido através dela (King et al.⁶⁰ 2005; de Andrade e Silva et al.⁶¹ 2009). Excelentes resultados *in vitro* e clínicos tem sido mostrados após a colocação de um revestimento de resina hidrofóbica sobre um adesivo AC de 1-passo (Reis et

al.⁶² 2008; Reis et al.⁶³ 2009), no entanto esta técnica não foi testada com os novos adesivos “universais”.

2. PROPOSIÇÃO

O objetivo deste estudo foi avaliar o efeito da aplicação das estratégias adesivas convencional ou autocondicionante, modificadas ou não pela aplicação de uma camada de resina hidrofóbica adicional, nas propriedades de união à dentina de diferentes sistemas adesivos universais.

2.1 Objetivos específicos

1. Avaliar resistência de união à microtração, nanoinfiltração e grau de conversão na camada híbrida de adesivos universais aplicados em dentina, de acordo com as estratégias adesivas convencional e autocondicionante, no tempo imediato (24 h).
2. Avaliar resistência de união à microtração e nanoinfiltração de adesivos universais aplicados em dentina de acordo as estratégias adesivas convencional e autocondicionante no tempo imediato (24 h) e após seis meses.
3. Avaliar a resistência de união à microtração, nanoinfiltração e grau de conversão na camada híbrida de adesivos universais aplicados com uma camada hidrofóbica em dentina para as estratégias adesivas convencional e autocondicionante, no tempo imediato (24 h).

3. MATERIAL E MÉTODOS

3.1. Discriminação dos experimentos constantes no presente trabalho

Os experimentos que compõem o presente trabalho são: Experimento 1. Avaliação das propriedades imediatas de união de adesivos universais à dentina (Capítulo 1: *Immediate bonding properties of universal adhesives to dentine*); Experimento 2: Avaliação da longevidade das propriedades de união de adesivos universais à dentina (Capítulo 2: *Longevity of bonding properties of universal adhesives to dentin*); e Experimento 3: Influência de uma camada de resina hidrofóbica na performance adesiva de adesivos universais à dentina (Capítulo 3: *Influence of a hydrophobic resin coat on the bonding efficacy of three universal adhesives*).

Foram utilizados sete tipos de materiais adesivos distribuídos nos três experimentos desenvolvidos (**Quadro 1**).

Quadro 1 – Materiais adesivos, composição e fabricante

Material adesivo	Componente: Composição	Fabricante
Adper Single Bond 2	1. Condicionador: ácido fosfórico 35% (Scotchbond Etchant) 2. Adesivo: Bis-GMA, HEMA, dimetacrilato, etanol, água, fotoiniciador, metacrilato copolimérico funcional de ácido poliacrílico e politacônico, partículas de sílica esféricas de 5 nm de diâmetro (10% peso)	3M ESPE, Saint Paul, Minnesota, EUA
Clearfil SE Bond	1. <i>Primer</i> : água, MDP, HEMA, camforquinona, dimetacrilato hidrofílico 2. <i>Bond</i> : MDP, Bis-GMA, HEMA, canforquinona, dimetacrilato hidrofílico, N,N-dietanol p-toluidina bond, sílica coloidal	Kuraray, Minami-ku, Okayama, Japão
Peak Universal	1. Condicionador: ácido fosfórico (Ultraetch) 2. <i>Primer</i> : álcool etílico, ácido metacrílico, 2-hidroxietil metacrilato (Peak SE Primer) 3. Adesivo: álcool etílico, 2-hidroxietil metacrilato (Peak LC Bond)	Ultradent Products Inc., South Jordan, Utah, EUA
Scotchbond Universal *	1. Condicionador: ácido fosfórico 34%, água, sílica amorfa sintética, polietileno glicol, óxido de alumínio (Scotchbond Universal Etchant) 2. Adesivo: monômero de fosfato MDP, dimetacrilato, HEMA, copolímero de ácido polialquenoico metacrilato modificado, carga de partículas, etanol, água, iniciadores, silano	3M ESPE, Saint Paul, Minnesota, EUA
All-Bond Universal	1. Condicionador: ácido fosfórico 32%, cloreto de benzalcônio (Uni-Etch) 2. Adesivo: MDP, Bis-GMA, HEMA, etanol, água, iniciadores	Bisco Inc., Schaumburg, Illinois, EUA
G-Bond Plus**	Acetona, dimetacrilato, 4-metacrietil trimetilico anidrido, monômero éster de ácido fosfórico, dióxido de silício, fotoiniciador, água destilada.	GC Corporation Tokyo, Japão
Heliobond	Bis-GMA, TEGDMA	Ivoclar Vivadent, Schaan, Liechtenstein

* distribuído no Brasil como Single Bond Universal, **distribuído também como Gaenial Bond; Bis-GMA: bisfenol glicidil metacrilato; HEMA: metacrilato de 2-hidroxietila; MDP: 10-metacrilóiloxidecil dihidrogênio fosfato; TEGDMA: trietileno glicol dimetacrilato; * distribuído no Brasil como Single Bond Universal, **distribuído também como Gaenial Bond

3.1.1 Obtenção e preparação dos dentes

Cento e quarenta molares humanos livres de cárie foram obtidos do Banco de Dentes da Universidade Estadual de Ponta Grossa sob o protocolo de pesquisa número 17878/11, aprovado pelo Comitê de Ética (Anexo A).

Os dentes foram desinfetados em cloramina 0,5% e armazenados em água destilada trocada semanalmente até sua utilização. Com o auxílio de uma politriz (Arotec E, Cotia, São Paulo, Brasil) e usando lixa de carbetto de silício de granulação #180, sob irrigação constante, foi retirado o esmalte oclusal e planificada a superfície dos molares até exposição da dentina. As bordas periféricas de esmalte foram totalmente desgastadas com pontas diamantadas nº 4137 (KG Sorensen, Barueri, São Paulo, Brasil) em alta rotação e irrigação constante deixando unicamente tecido dentinário (**Figura 1**). A seguir, as superfícies de dentina foram polidas manualmente sob irrigação por 60 s para a padronização da *smear layer* utilizando lixas de carbetto de silício de granulação #600 (Extec, Enfield, Connecticut, EUA).

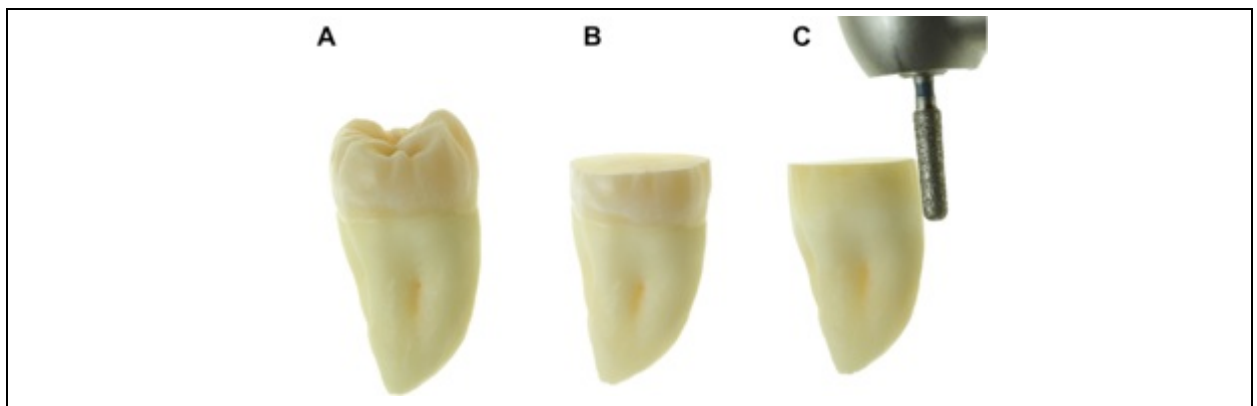


Figura 1 - Preparo da superfície de dentina. (A) dente hígido; (B) superfície oclusal planificada e em dentina; desgaste de todas as bordas periféricas de esmalte.

3.1.2 Desenho experimental e procedimentos adesivos e restaurador

Para o Experimento 1, bem como para o Experimento 2, 40 dentes preparados como descrito anteriormente foram distribuídos aleatoriamente em oito grupos experimentais (n=5), de acordo com o adesivo e as estratégias adesivas convencional (CON) ou autocondicionante (AC). A sequência técnica das estratégias adesivas está demonstrada nas **Figuras 2 e 3**, respectivamente. Como grupos controle foram selecionados os sistemas adesivos Adper Single Bond 2 (SB),

segundo a estratégia CON de 2 passos; e Clearfil SE Bond (CSE), segundo a estratégia autocondicionante (AC), também de 2-passos. Os grupos experimentais do estudo foram constituídos por três adesivos universais aplicados com estratégias CON ou AC: Peak Universal (Peak SE Primer e Peak LC Bond), aplicado como 2-passos CON (PkCon) e 2-passos AC (PkAc); Scotchbond Universal, aplicado como 2-passos CON (ScCon) e 1-passo AC (ScAc); e All-Bond Universal aplicado como 2-passos CON (AlCon) e 1-passo AC (AlAc) (**Quadro 2**).

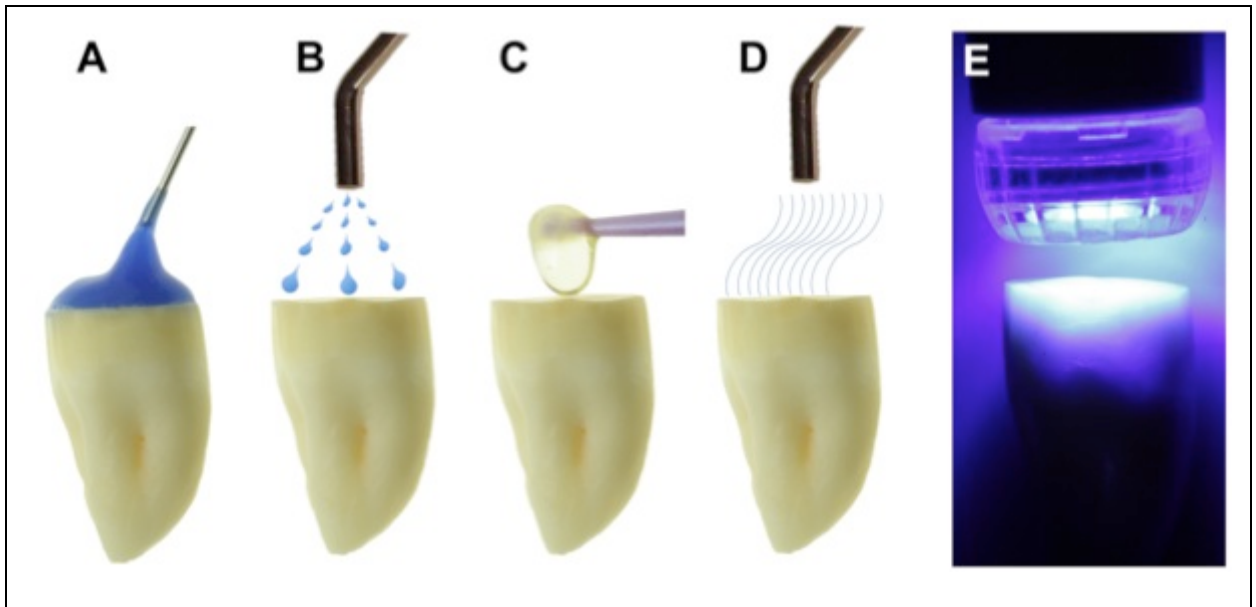


Figura 2 – Estratégia adesiva convencional. (A) condicionamento ácido; (B) lavagem e secagem; (C) Aplicação do adesivo; (D) secagem para evaporação do solvente; (E) fotoativação. Todos os procedimentos realizados segundo as indicações do fabricante, os quais estão discriminados no quadro 2.

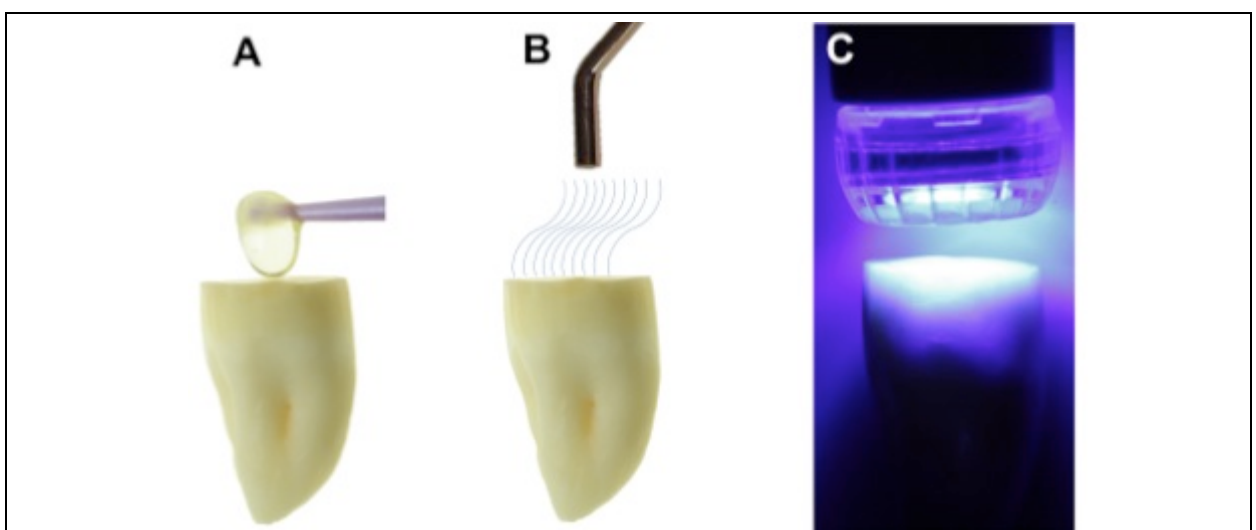


Figura 3 - Estratégia adesiva autocondicionante (dispensa a aplicação e a lavagem do ácido fosfórico). (A) Aplicação do adesivo; (B) secagem para evaporação do solvente; (C) fotoativação. Todos os procedimentos realizados segundo as indicações dos fabricantes, os quais estão discriminados no quadro 2.

Para o Experimento 3, 60 terceiros molares foram divididos em 12 grupos (n=5), de acordo com o adesivo e a estratégia adesiva (**Figuras 2 e 3**), da mesma maneira como descrito para os Experimentos 1 e 2, porém variando-se alguns dos materiais selecionados. Além disso, este Experimento avaliou o efeito da adição (**Figuras 4 e 5**) ou não (**Figuras 2 e 3**) de uma camada final de adesivo hidrofóbico nas duas estratégias de união. Os grupos experimentais formados pelas combinações descritas anteriormente foram: Scotchbond Universal convencional (ScCon), convencional + camada hidrofóbica (ScCon-HE), autocondicionante (ScAc), autocondicionante + camada hidrofóbica (ScAc-HE); Allbond Universal convencional (AlCon), convencional + camada hidrofóbica (AlCon-HE), autocondicionante (AlAc), autocondicionante + camada hidrofóbica (AlAc-HE); G-Bond Plus convencional (GbpCon), convencional + camada hidrofóbica (GbpCon-HE), autocondicionante (GbpAc), autocondicionante + camada hidrofóbica (GbpAc-HE) (**Quadro 3**).

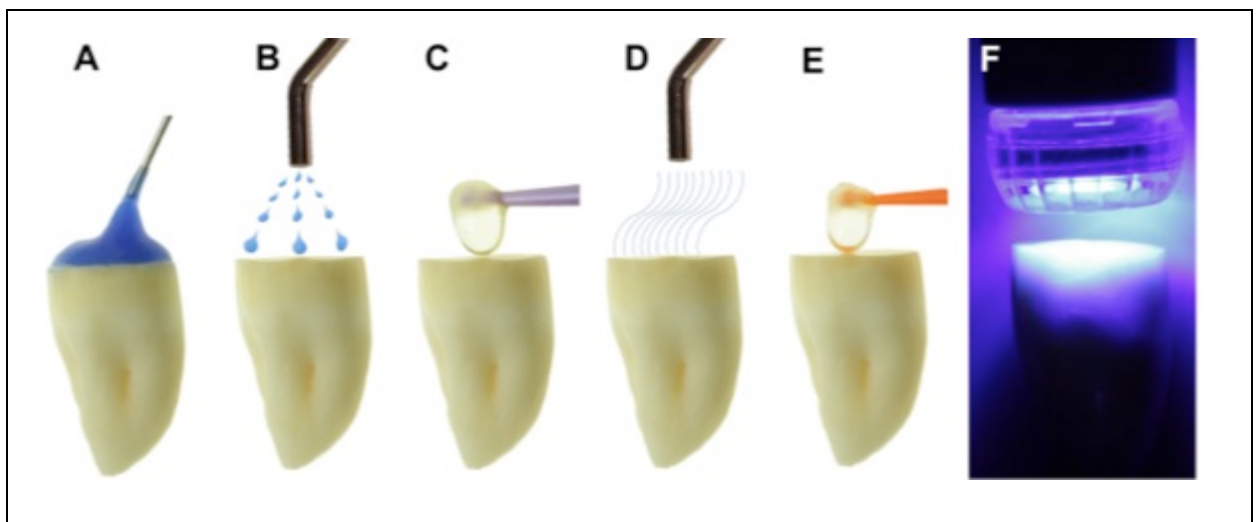


Figura 4 - Estratégia adesiva convencional com aplicação de camada hidrofóbica. (A) condicionamento ácido; (B) lavagem e secagem; (C) Aplicação do adesivo; (D) secagem para evaporação do solvente; (E) aplicação de uma camada de resina hidrofóbica; (F) fotoativação. Vide Quadro 3.

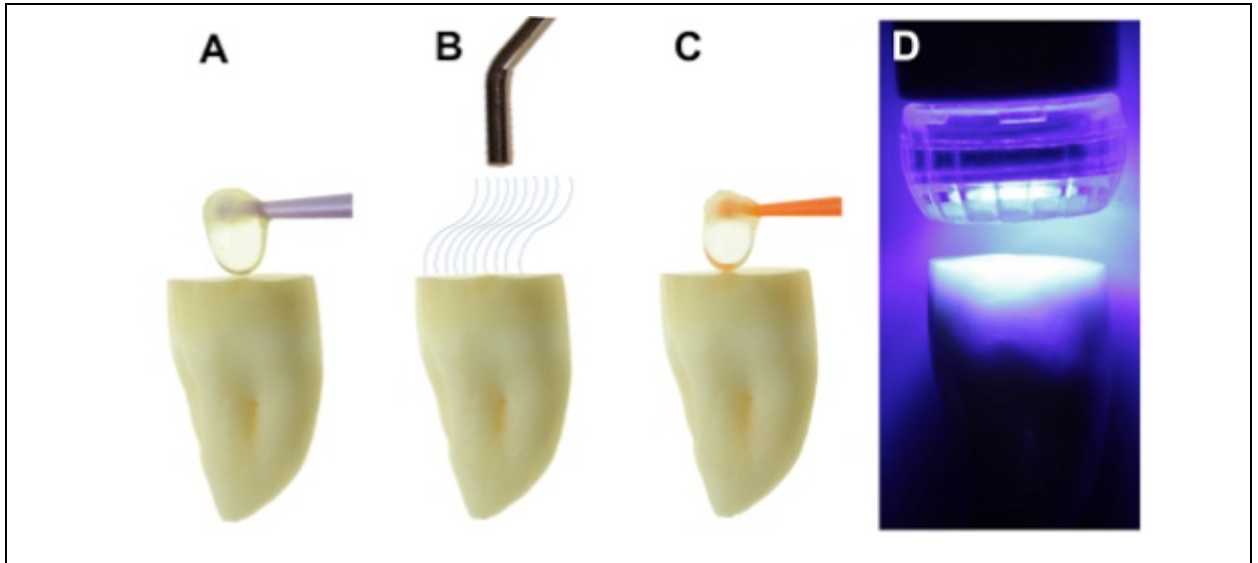


Figura 5 - Estratégia adesiva autocondicionante, a qual dispensa o condicionamento com ácido fosfórico, com aplicação de camada hidrofóbica; (A) aplicação do adesivo; (B) secagem para evaporação do solvente; (C) aplicação de camada de resina hidrofóbica; (D) fotoativação. Vide Quadro 3.

Quadro 2 - Sistemas adesivos e estratégias de aplicação de acordo com as instruções do fabricante para os Experimentos 1 e 2

Sistema adesivo (lote),	Estratégia Autocondicionante	Estratégia convencional
Adper Single Bond 2 (BPBR)	N.A	<ol style="list-style-type: none"> 1. Aplicar condicionador por 15 s 2. Lavar por 10 s 3. Remover o excesso de água 4. Aplicar 2-3 camadas consecutivas do adesivo por 15 s com agitação suave 5. Aplicar suave jato de ar por 5 s 6. Fotoativar por 10 s*
Clearfil SE Bond (Primer: 00954A - Bond: 01416 ^a)	<ol style="list-style-type: none"> 1. Aplicar o <i>primer</i> sobre a superfície do dente e deixar por 20 s 2. Secar com jato de ar para evaporar os componentes voláteis 3. Aplicar <i>bond</i> na superfície do dente, criando uma película uniforme usando um jato de ar suave 4. Fotoativar por 10 s* 	N.A
Peak Universal Adhesive System (Peak SE Primer: 0N062 – Peak LC Bond: Y062)	<ol style="list-style-type: none"> 1. Ativar os dois componentes separados na seringa 2. Aplicar "Peak SE" com microbrush por 20 s esfregando continuamente em dentina. 3. Aplicar 3 s de jato de ar/água ou sucção de alto volume diretamente sobre a preparação para afinar a camada 4. Aplicar uma generosa camada de Peak LC Bond, com agitação suave durante 10 s 5. Afinar e secar a camada usando 10 s com ¼ a ½ da pressão de ar 6. Fotoativar por 10 s* 	<ol style="list-style-type: none"> 1. Fazer condicionamento ácido por 20 s 2. Lavar por 5 s 3. Secar com jato de ar por 2 s 4. Aplicar uma generosa camada de Peak LC Bond, com agitação suave durante 10 s 5. Afinar e secar a camada usando 10 s com ¼ a ½ da pressão de ar 6. Fotoativar por 10 s*
Scotchbond Universal (D-82229)	<ol style="list-style-type: none"> 1. Aplicar o adesivo na preparação com um microbrush e esfrega-lo por 20 s. Se necessário, molhe com adesivo a microbrush durante o tratamento 2. Dirigir um ligeiro jato de ar sobre o líquido durante 5 s aproximadamente até a camada não se mover e o solvente estiver completamente evaporado 3. Fotoativar por 10 s* 	<ol style="list-style-type: none"> 1. Fazer condicionamento ácido por 15 s 2. Lavar por 10 s 3. Secar com jato de ar por 2 s 4. Aplicar adesivo como indicado na estratégia autocondicionante
All-Bond Universal (1200006111)	<ol style="list-style-type: none"> 1. Aplicar duas camadas de adesivo separadamente, esfregar sobre a preparação por 10-15 s por cada camada. Não fotoativar entre as camadas. 2. Evaporar cuidadosamente o excesso de solvente secando com jato de ar durante pelo menos 10 s, não devendo haver nenhum movimento visível do material. A superfície deve ter uma aparência brilhante e uniforme 3. Fotoativar por 10 s* 	<ol style="list-style-type: none"> 1. Fazer condicionamento ácido por 15 s 2. Lavar bem 3. Remover excessos de água com pellets absorventes ou sucção de alta potência por 1-2 s 4. Aplicar adesivo como indicado na estratégia autocondicionante

* aparelho LED de 1200 mW/cm² de intensidade (Radium-cal, SDI Limited, Bayswater, Victoria, Austrália)

Quadro 3 - Materiais adesivos e estratégias de aplicação dos sistemas adesivos utilizados no Experimento 3

Material adesivo (lote)	Estratégia de aplicação			
	Autocondicionante		Convencional	
	Sem camada hidrofóbica	Com camada hidrofóbica	Sem camada hidrofóbica	Com camada hidrofóbica
Heliobond - HE (N37749)				
Scotchbond Universal Adhesive - SBU (448716)	<ol style="list-style-type: none"> 1. Aplicar o adesivo na preparação com um microbrush e esfregue-o por 20 s. Se necessário, molhe novamente com adesivo o microbrush durante o tratamento 2. Dirigir um ligeiro jato de ar sobre o líquido durante 5 s aproximadamente até que a camada não se mova e o solvente esteja completamente evaporado 3. Fotoativar por 10 s* 	<ol style="list-style-type: none"> 1. Repetir os passos da estratégia autocondicionante 2. Aplicar uma camada muito fina de Heliobond com microbrush sobre a superfície dentinária 3. Aplicar jato de ar para conseguir uma camada fina 4. Fotoativar por 10 s* 	<ol style="list-style-type: none"> 1. Condicionamento com ácido fosfórico por 15 s 2. Lavar por 10 s 3. Secar com ar durante 5 s 4. Aplicar adesivo como na estratégia autocondicionante 	<ol style="list-style-type: none"> 1. Repetir os passos da estratégia convencional 2. Aplicar uma camada muito fina de Heliobond com microbrush sobre a superfície dentinária 3. Aplicar jato de ar para conseguir uma camada fina 4. Fotoativar por 10 s*
All-Bond Universal - ABU (1200002722)	<ol style="list-style-type: none"> 1. Aplicar duas camadas de adesivo separadamente, esfregar sobre a preparação por 10-15 s por cada camada. Não fotoativar entre as camadas. 2. Evaporar cuidadosamente o excesso de solvente secando com jato de ar durante pelo menos 10 s, não devendo haver nenhum movimento visível do material. A superfície deve ter uma aparência brilhante e uniforme 3. Fotoativar por 10 s* 	<ol style="list-style-type: none"> 1. Aplicar da mesma forma autocondicionante 2. Aplicar uma camada muito fina de Heliobond com microbrush sobre a superfície dentária 3. Jato de ar para conseguir uma camada fina 4. Fotoativar por 10 s* 	<ol style="list-style-type: none"> 1. Fazer condicionamento ácido por 15 s 2. Lavar bem 3. Remover excessos de água com pellets absorventes ou sucção de alta potência por 1-2 s 4. Aplique adesivo como indicado na estratégia autocondicionante 	<ol style="list-style-type: none"> 1. Repetir os passos da estratégia convencional 2. Aplicar uma camada muito fina de Heliobond com microbrush sobre a superfície dentinária 3. Aplicar jato de ar para conseguir uma camada fina 4. Fotoativar por 10 s*
G-Bond Plus - GBP (1102221)	<ol style="list-style-type: none"> 1. Aplicar usando um microbrush 2. Deixar em repouso durante 10 s após o fim da aplicação 3. Secar cuidadosamente por 5 s com ar livre de óleo sob pressão de ar máxima, utilizando sucção a vácuo 4. Fotoativar por 10 s* 	<ol style="list-style-type: none"> 1. Repetir os passos da estratégia autocondicionante 2. Aplicar uma camada muito fina de Heliobond com microbrush sobre a superfície dentária 3. Jato de ar para se obter camada fina 4. Fotoativar por 10 s* 	<ol style="list-style-type: none"> 1. Aplicar ácido fosfórico gel 34% por 10 s (Scotchbond Universal Etchant) 2. Lavar por 5s e secar suavemente 3. Aplicar o adesivo como na estratégia de autocondicionante <p>NOTA: O fabricante não indica condicionamento com ácido fosfórico em dentina</p>	<ol style="list-style-type: none"> 1. Aplicar da mesma forma como na estratégia convencional 2. Aplicar uma camada muito fina de Heliobond com microbrush sobre a superfície dentinaria 3. Jato de ar para conseguir uma camada fina 4. Fotoativar por 10 s*

* Aparelho LED de 1200 mW/cm² de intensidade (Ratii-cal, SDI Limited, Bayswater, Victoria, Austrália)

Os sistemas adesivos foram aplicados em conformidade com as instruções dos respectivos fabricantes, a partir das quais foram desenhadas as variáveis adesivas de todos os experimentos. As únicas exceções ocorreram no Experimento 3, pois alguns grupos receberam a camada final de resina hidrofóbica, e, ainda no mesmo experimento, pois se realizou o condicionamento com ácido fosfórico para o material G-bond Plus. Todos os procedimentos adesivos e restauradores foram realizados por um único operador em ambiente controlado (24°C).

Os sistemas adesivos foram fotoativados usando um aparelho LED de 1200 mW/cm² de intensidade (Radii-cal, SDI Limited, Bayswater, Victoria, Austrália). Após os procedimentos adesivos, todos os dentes foram restaurados com uma resina composta microhíbrida de restauração (Opallis, FGM Produtos Odontológicos, Joinville, Santa Catarina, Brasil) em dois incrementos de 2 mm sendo cada incremento fotoativado por 40 s com o mesmo aparelho LED (**Figura 6**).

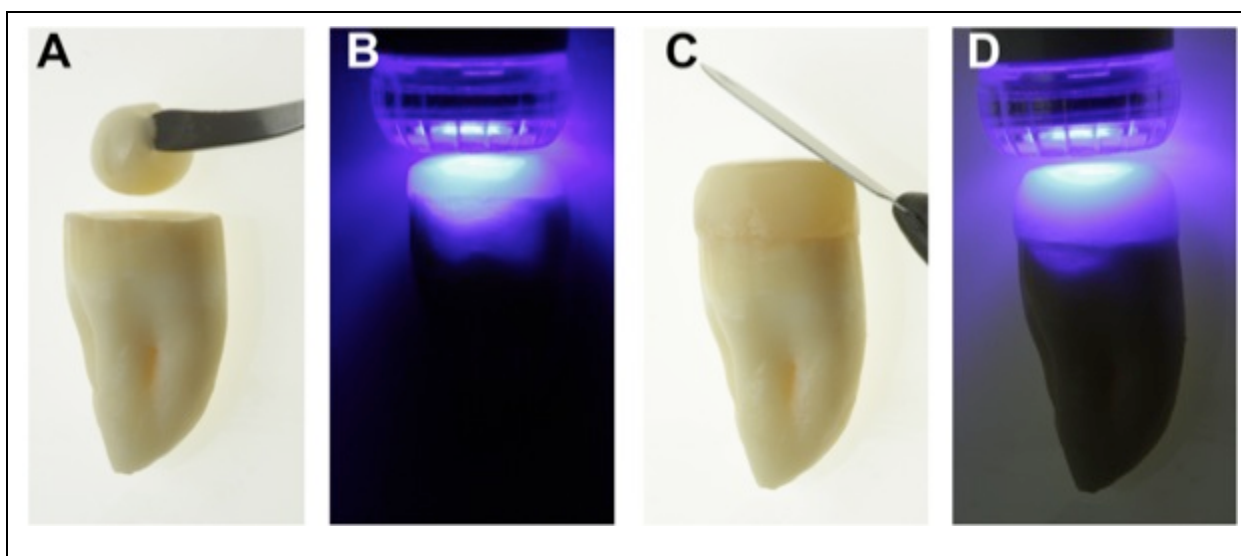


Figura 6 – Restauração dos dentes após aplicação das variáveis adesivas. (A) Colocação da resina composta; (B) fotoativação do primeiro incremento; (C) Colocação do incremento final; (D) fotoativação do segundo incremento.

Os dentes restaurados foram armazenados em água destilada a 37° C por 24 h e levados a uma máquina de corte (Isomet 1000, Buehler Ltd., Lake Bluff, Illinois, EUA). Com ajuda de um disco diamantado (15 HC-4in, Buehler Ltd., Lake Bluff, Illinois, EUA) sob irrigação a 300 rpm os dentes foram seccionados em duas seqüências de cortes perpendiculares uma no eixo x e outra no eixo y, obtendo-se 25-30 “palitos” (**Figura 7**) com uma área de secção transversal de $0,8 \pm 0,1 \text{ mm}^2$,

medida com um paquímetro digital (Digimatic Caliper, Mitutoyo, Kanagawa, Tóquio, Japão, **Figura 8B**).

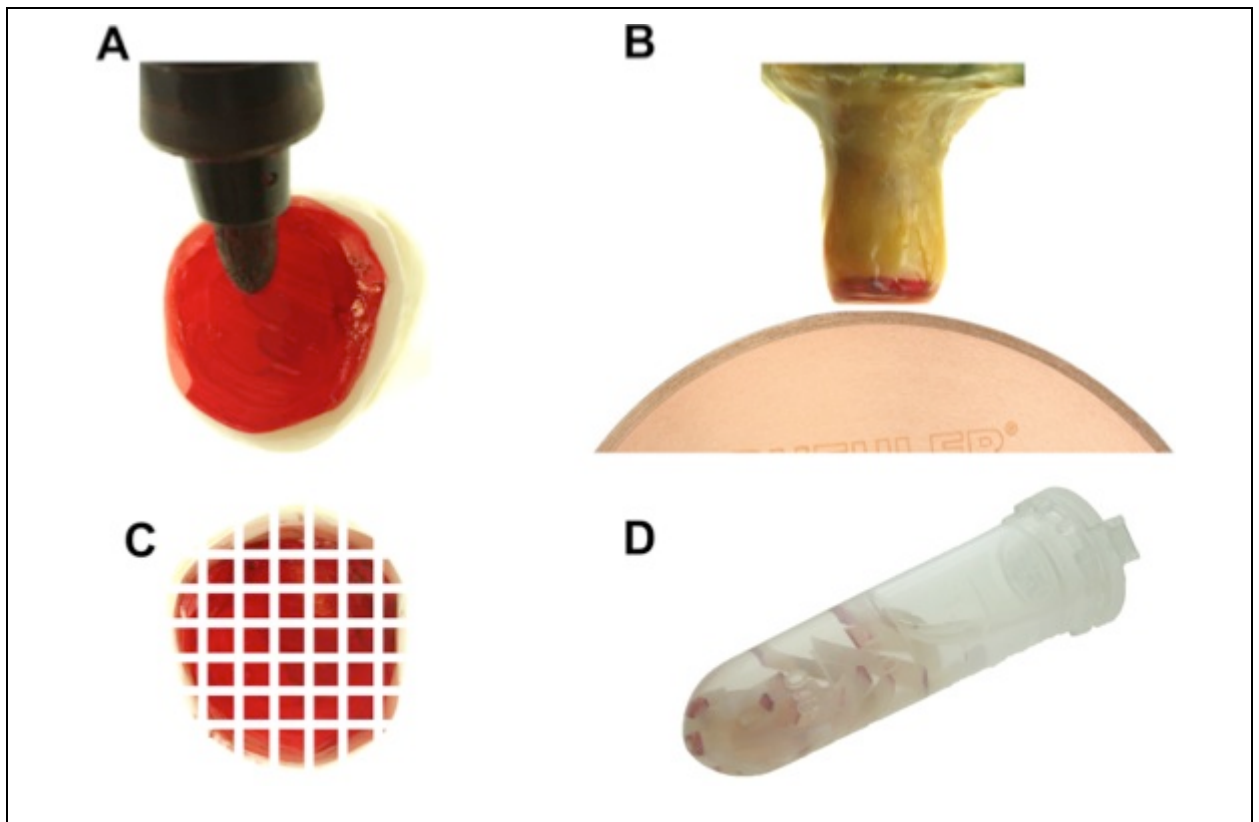


Figura 7 - Sequência de corte para a obtenção dos palitos. (A) superfície oclusal das coroas de resina compostas pintadas com caneta marcadora permanente de cores diferentes para identificação dos palitos de cada grupo. (B) dente fixado com cera pegajosa ao dispositivo da máquina de corte montada com um disco diamantado; (C) duas sequências perpendiculares de corte, uma no eixo x e outra no eixo y, permitiram a obtenção dos palitos para o teste de resistência de união; (D) em *ependorfs*, os palitos de cada dente foram armazenados em água destilada por 24 h.

Nos Experimentos 1 e 3 a totalidade dos palitos foi utilizada imediatamente após as 24 h de armazenamento. No Experimento 2, metade dos palitos de cada dente foi selecionada aleatoriamente para serem testados imediatamente (24 h) e o restante após 6 meses de armazenamento em água destilada. Alguns palitos de cada tempo de avaliação foram aleatoriamente selecionados e distribuídos nos testes de grau de conversão ($n = 3$) e nanoinfiltração ($n = 3$ no tempo de 24 h e $n = 3$ aos 6 meses) como será descrito.

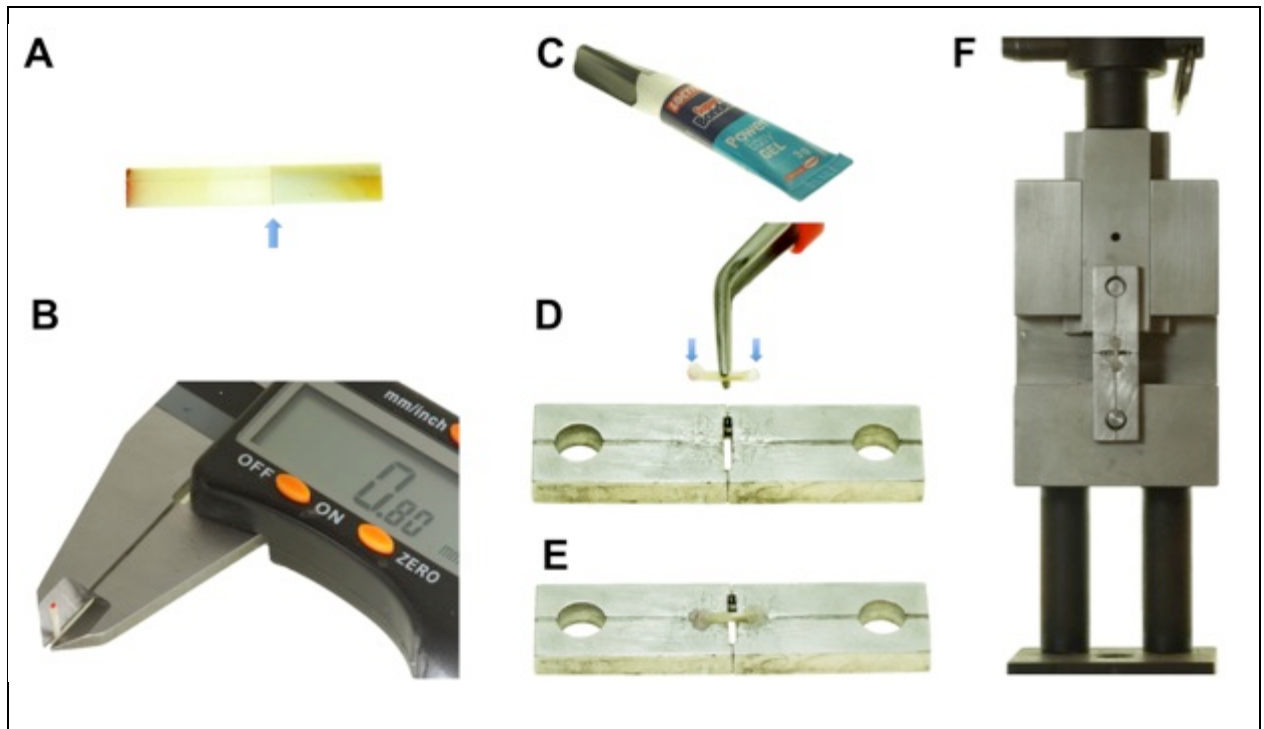


Figura 8 – Sequência para o teste de microtração. (A) a seta azul indica a interface adesiva entre dente e restauração, numa vista lateral do palito. (B) mensuração da largura e da espessura do mesmo palito, com paquímetro digital, para o cálculo da área da interface adesiva. (C) cola de cianocrilato em gel (que permite controle da aplicação). (D) aplicação da cola nas extremidades do palito, protegendo a interface com a pinça; (E) posicionamento do palito nas garras de microtração deixando a interface adesiva no espaço central da garra; (F) adaptação das garras na máquina de ensaios universais após a polimerização da cola.

3.1.3 Teste de resistência de união à microtração (RU)

Os palitos obtidos foram colados individualmente em garras de Geraldeli (Odeme Biotechnology, Joaçaba, Santa Catarina, Brasil) (Perdigao et al.⁶⁴ 2002) aplicando em cada um dos extremos do palito cola de cianocrilato gel (Loctite Superbonder Power Easy Gel, Henckel, São Paulo, São Paulo, Brasil) de forma a não contaminar a interface dente-restauração (**Figura 8**). Para acelerar o tempo de polimerização da cola, sobre sua superfície foi pincelado (Cavibrush, FGM Produtos Odontológicos, Joinville, Santa Catarina, Brasil) líquido de resina acrílica autopolimerizável para prótese removível (Jet, Artigos Odontológicos Clássico, São Paulo, SP, Brasil). Após o tempo de polimerização da cola, as garras com os palitos foram acopladas na máquina de ensaios universal (Kratos Dinamômetros, Cotia, São Paulo, Brasil, **Figura 8F**) e estes foram testados por tração a uma velocidade de 0,5 mm/min com célula de carga de 100 N até rompimento da interface. Os valores de

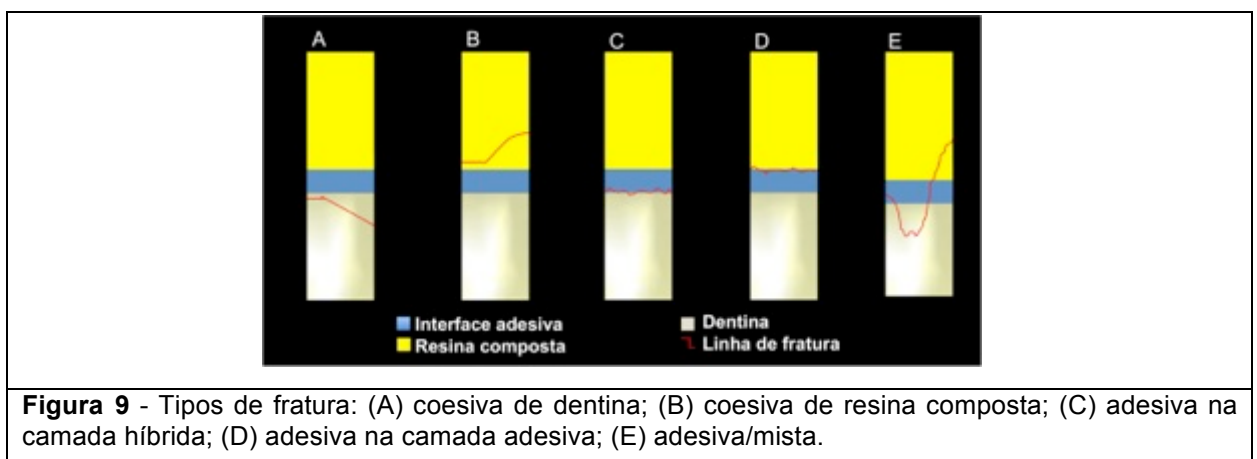
RU de cada palito em Megapascal (MPa) foram calculados pela divisão da tensão máxima a ruptura (N) e a área de secção transversal (mm²) obtida previamente de cada palito.

A unidade de medida foi o dente, assim uma média do valor RU em MPa dos palitos provenientes do mesmo dente foi obtida, totalizando cinco valores médios por grupo experimental.

3.1.4 Tipos de fratura

Utilizando uma lupa esteoreoscópica a 100x de magnificação (Olympus SZ40, Shinjuku-ku, Tóquio, Japão) a falha de cada palito foi classificada segundo o tipo de fratura como (**Figura 9**):

- Adesiva, falha na interface adesiva dente/restauração.
- Coesiva, falha exclusivamente em dentina ou resina composta.
- Adesiva/Mista, falha na interface adesiva dente/restauração, mas que incluiu falha coesiva dos substratos vizinhos.
- Falhas prematuras, palitos que quebraram antes dos testes.



3.1.5 Grau de conversão (GC)

Como descrito anteriormente, três palitos por cada dente foram selecionados aleatoriamente e submetidos ao teste de GC. Previamente ao teste, os palitos foram colados com parafina em *stubs* (**Figura 10**) e levados a uma politriz manual que

permite a irrigação com água constante no momento do polimento, limpando continuamente as partículas geradas. A sequência de lixas de SiC foi #1500; 2000 e 2500 (Buehler Ltd., Lake Bluff, Illinois, EUA) foi aplicada por 30 s cada uma. A seguir os palitos foram lavados em cuba ultrassônica (Dabi Atlante – 3L, Ribeirão Preto, São Paulo, Brasil) por 20 min e armazenados em água destilada a 37° C por 24 h antes da leitura de GC.

A análise de GC foi realizada na camada híbrida com um espectrofotômetro de Raman (Senterra equipment, Bruker Optik GmbH, Ettlingen, Baden-Württemberg, Alemanha). Antes das análises, o equipamento era calibrado para redefinir o zero utilizando padrões de silicone com valores de coeficientes conhecidos. As análises foram realizadas sob os seguintes parâmetros: laser de Neônio com 532 nm de comprimento de onda a 20 mW de potência; resolução espacial de $\approx 3 \mu\text{m}$; resolução espectral de $\approx 5 \text{ cm}^{-1}$, com 30 s de tempo de acumulação e 6 co-adições, em magnificação de 100X para um laser de $\approx 1 \mu\text{m}$ de diâmetro. A análise iniciou obtendo-se os espectros com os picos de referência dos adesivos não polimerizados (*np*). Para isso, cada sistema adesivo teve todos os seus componentes dispensados e misturados em uma placa de vidro, a qual foi levada ao espectrofotômetro para leitura conforme os parâmetros já descritos. A leitura dos adesivos polimerizados (*p*) foi realizada diretamente na camada híbrida. Considerando as duplas ligações dos compostos alifáticos (1639 cm^{-1} , pico *reativo*, aqui são detectadas mudanças) e aromáticos (1609 cm^{-1} , pico *referência*, estável sem mudanças). Para ambas as análises (*np* e *p*) foram determinados os valores da integral (I) dos picos registrados utilizando o software do equipamento Opus Spectroscopy 6.5 (Bruker Optik GmbH, Ettlingen, Baden-Württemberg, Alemanha). Assim, o GC de duplas ligações do monômero em polímero foi calculado com a seguinte fórmula:

$$GC \% = \left[1 - \frac{I \text{ Reativo}(p)_{1639} / I \text{ Referência}(p)_{1609}}{I \text{ Reativo}(np)_{1639} / I \text{ Referência}(np)_{1609}} \right] \times 100$$

As análises espectrais foram obtidas de três sítios aleatoriamente selecionados na camada híbrida de cada palito. A partir disso foi possível a obtenção das médias de cada dente, as quais foram utilizadas na análise estatística ($n = 3$). As leituras iniciais (*np*) também foram realizadas em triplicata.



Figura 10 - *Stub* com palitos lixados previamente à leitura no micro-Raman. Isto facilita a localização da camada híbrida.

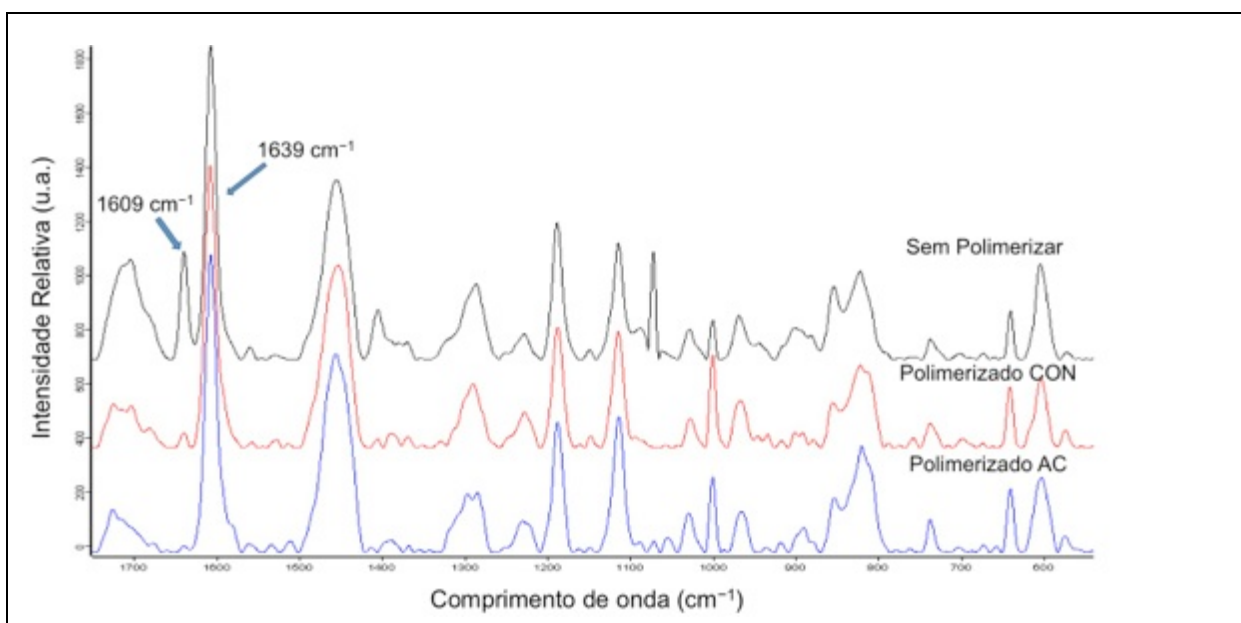


Figura 11 – Espectros representativos obtidos do sistema adesivo Peak Bond Universal não polimerizado, e após sua fotoativação na camada híbrida *in situ* em sua versão ER e SE. As setas indicam os picos de referencia (1609 cm^{-1}) e reativo (1639 cm^{-1}).

3.1.6 Avaliação da Nanoinfiltração (NI) em Microscopia Eletrônica de Varredura (MEV)

Três palitos provenientes de cada dente para os tempos imediato e 6 meses, foram reservados para análise de NI (**Figura 11**). Utilizando eppendorfs (OA12, Odeme Biotechnology, Joaçaba, SC, Brasil), os palitos foram imersos em uma solução aquosa (pH=7,0) de nitrato de prata amoniacal 50% em peso (Tay et al.⁶⁵ 2002) e armazenados por 24 h em ausência de luz. Após o tempo de infiltração, os palitos foram lavados profusamente com água destilada e imersos em solução reveladora (Kodak, Rochester, Nova York, EUA) por 8 h sob luz fluorescente para a redução dos íons de prata ao longo das nanoporosidades da interface de união. Assim, os palitos foram novamente lavados em água corrente e fixados com parafina em *stubs* de alumínio para facilitar o polimento uniforme das amostras, realizado em

uma politriz de bancada (ALM 4, Arotec, Cotia, São Paulo, Brasil) sob escoamento contínuo da água com lixas de granulações 1000, 1200, 1500, 2000, 2500 e pastas diamantadas com granulações decrescentes de 1 μm e 0,25 μm (Buehler Ltd, Lake Bluff, IL, USA). Entre cada lixa e pasta de polimento as amostras foram lavadas em cuba ultrassônica por 8 min seguidos de escoamento de água sobre a superfície por 2 min. As amostras foram deixadas, então, por 30 min em cuba ultrassônica. Para o processo de desidratação foram deixadas em um dessecador contendo sílica coloidal por 24 h para, então, serem metalizadas em carbono/ouro (MED 010, Balzers Union, Balzers, Liechtenstein).

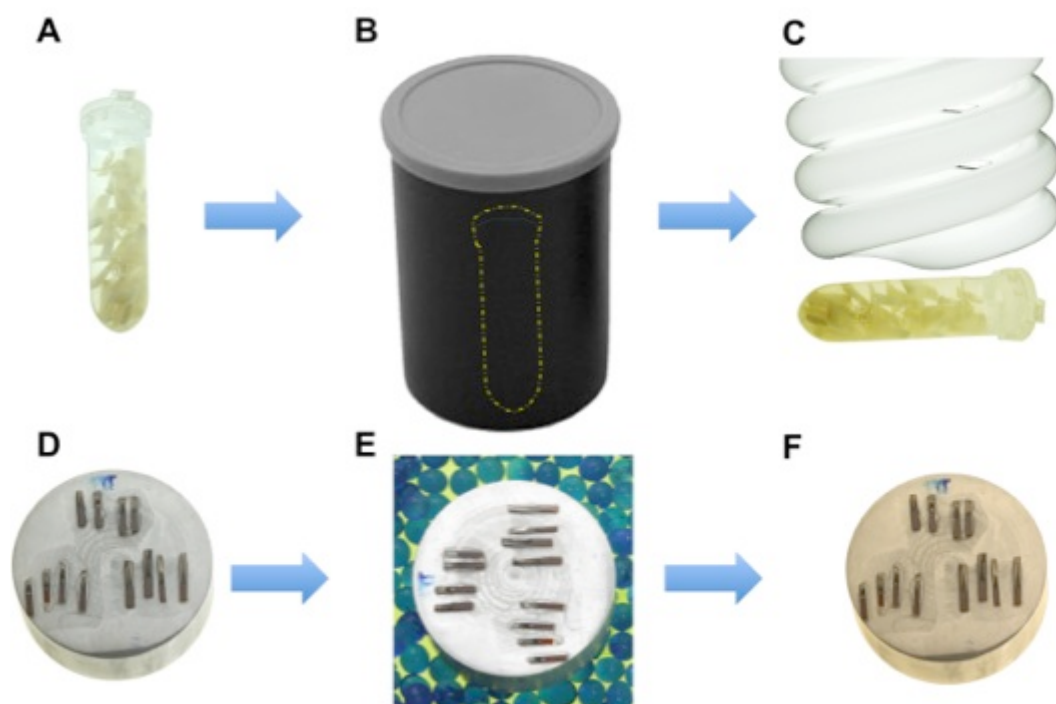


Figura 12 - Preparação de amostras para NI: (A) palitos imersos em uma solução aquosa de nitrato de prata (pH=7,0); (B) armazenados por 24 h sob total escuridão; (C) após lavagem, imersos em solução reveladora por 8 h sob luz fluorescente; (D) palitos revelados, montados em *stubs* de alumínio para facilitar o polimento uniforme; (E) palitos dessecados em sílica por 24 h; (F) palitos metalizados para a análise de MEV.

As interfaces resina-dentina foram analisadas num microscópio eletrônico de varredura de emissão de campo (LEO 435 VP, LEO Electron Microscopy Ltd., Cambridge, United Kingdom) em modo de elétrons retroespalhados. Um operador que desconhecia a origem dos palitos a serem analisados realizou três fotomicrografias com aumento 1200X da interface dente-restauração de cada um dos palitos, sendo uma no centro e outras duas 3 μm à esquerda e à direita do centro. A porcentagem relativa de nitrato de prata nas áreas ocupadas pela camada híbrida e

camada de adesivo foram consideradas na mensuração automatizada da NI utilizando um software UTHSCSA ImageTool 3.0 software (Department of Dental Diagnostic Science, University of Texas, San Antonio, Texas, EUA) operado por apenas um avaliador, o qual desconhecia a que grupo experimental pertencia a imagem. Assim, foram obtidas as médias dos valores percentuais de NI dos palitos provenientes do mesmo dente para fins estatísticos.

3.1.7 Medição do pH

O pH dos sistemas adesivos foi mensurado com um pH-metro E520 (Metrohm, Appenzell Exterior, Herisau, Suíça). O equipamento foi calibrado em soluções tampão de pH 4,0 e 7,0. Para a extração dos íons de hidrogênio dos sistemas adesivos foram realizadas três misturas por cada adesivo com água destilada na proporção 1:9 em peso, as quais foram homogeneizadas com agitação manual e armazenadas por 12 h em escuridão. A seguir foram realizadas três leituras por cada mistura. Entre cada medição, o eletrodo do equipamento foi lavado por escoamento com álcool 90° por 30 s, seguido de água destilada por 30 s, finalizando com uma medição de ambas as soluções tampão (4,0 e 7,0) para conferir sua estabilidade. A média de valores por adesivo foi utilizada para fins estatísticos.



Figura 13 - Análise de pH. Vista do eletrodo totalmente submerso na solução para a leitura do pH do adesivo após 12 h de armazenamento em escuridão.

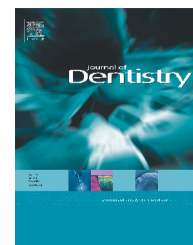
4. ARTIGOS

4.1 Capítulo 1 (do experimento 1)

IMMEDIATE BONDING PROPERTIES OF UNIVERSAL ADHESIVES TO DENTINE

Available online at www.sciencedirect.com

SciVerse ScienceDirect

journal homepage: www.intl.elsevierhealth.com/journals/jden

Immediate bonding properties of universal adhesives to dentine

Miguel Angel Muñoz^{a,b}, Issis Luque^a, Viviane Hass^a, Alessandra Reis^a,
Alessandro Dourado Loguercio^{a,*}, Nara Hellen Campanha Bombarda^a

^aSchool of Dentistry, Department of Restorative Dentistry, State University of Ponta Grossa, Ponta Grossa, Paraná, Brazil

^bSchool of Dentistry, Universidad de Valparaíso, Valparaíso, Chile

ARTICLE INFO

Article history:

Received 1 November 2012

Received in revised form

27 February 2013

Accepted 1 March 2013

Keywords:

Microtensile bond strength

Nanoleakage

Degree of conversion

Etch-and-rinse

Self-etch

Universal simplified adhesive systems

ABSTRACT

Objectives: To evaluate the dentine microtensile bond strength (μ TBS), nanoleakage (NL), degree of conversion (DC) within the hybrid layer for etch-and-rinse and self-etch strategies of universal simplified adhesive systems.

Methods: forty caries free extracted third molars were divided into 8 groups for μ TBS ($n = 5$), according to the adhesive and etching strategy: Clearfil SE Bond [CSE] and Adper Single Bond 2 [SB], as controls; Peak Universal Adhesive System, self-etch [PkSe] and etch-and-rinse [PkEr]; Scotchbond Universal Adhesive, self-etch [ScSe] and etch-and-rinse [ScEr]; All Bond Universal, self-etch [AlSe] and etch-and-rinse [AlEr]. After restorations were constructed, specimens were stored in water (37 °C/24 h) and then resin–dentine sticks were prepared (0.8 mm²). The sticks were tested under tension at 0.5 mm/min. Some sticks from each tooth group were used for DC determination by micro-Raman spectroscopy or nanoleakage evaluation (NL). The pH for each solution was evaluated using a pH metre. Data were analyzed with one-way ANOVA and Tukey's test ($\alpha = 0.05$).

Results: For μ TBS, only PkSe and PkEr were similar to the respective control groups ($p > 0.05$). AlSe showed the lowest μ TBS mean ($p < 0.05$). For NL, ScEr, ScSe, AlSe, and AlEr showed the lowest NL similar to control groups ($p < 0.05$). For DC, only ScSe showed lower DC than the other materials ($p < 0.05$).

Conclusions: Performance of universal adhesives was shown to be material-dependent. The results indicate that this new category of universal adhesives used on dentine as either etch-and-rinse or self-etch strategies were inferior as regards at least one of the properties evaluated (μ TBS, NL and DC) in comparison with the control adhesives (CSE for self-etch and SB for etch-and-rinse).

© 2013 Elsevier Ltd. All rights reserved.

1. Introduction

The bonding mechanism of adhesive systems basically involves the replacement of minerals removed from the

hard dental tissue by resin monomers, in such a way that a polymer becomes micro-mechanically interlocked to the dental substrate.¹ However, the adhesive systems available on the market can be classified into two categories: etch-and-rinse (Er) and those applied using self-etch strategies

* Corresponding author at: Universidade Estadual de Ponta Grossa, Pós-graduação em Odontologia, Rua Carlos Cavalcanti, 4748, Bloco M, Sala 64A, Uvaranas, 84030-900 Ponta Grossa, Paraná, Brazil. Tel.: +55 42 3027 7898; fax: +55 42 3220 3741.

E-mail addresses: aloguercio@hotmail.com, alelog@uepg.br (A.D. Loguercio).

Abbreviations: Co, composite; De, dentine; HL, hybrid layer; AL, adhesive layer.

0300-5712/\$ – see front matter © 2013 Elsevier Ltd. All rights reserved.

<http://dx.doi.org/10.1016/j.jdent.2013.03.001>

(Se), in versions of three (only Er), two or one application step.^{2,3}

When using the Er strategy, the first step involves the application of a phosphoric acid gel to both dental substrates, which allows removal of the smear layer, exposure of the collagen fibrils in dentine, and increase in surface area and surface energy in the enamel substrate. The primer is then applied (second step) followed by the bond (third step) resin separately or in a single solution.^{2–4} Irrespective of the number of steps, the main disadvantage of the Er system, mainly two-step versions, is that there is risk of collagen fibre collapse during the process of demineralized dentine drying, which leads to a decrease in bond strength.^{5,6} The collagen collapse is prevented by keeping demineralized dentine moist, which is a difficult task to perform clinically. In fact, adequate moisture depends on both the solvent used in the material⁷ and on the clinician's interpretation of the manufacturer's directions.

The incomplete impregnation of collagen fibers⁸ and the need to protect them against the degrading mechanisms present in the oral cavity environment,^{9,10} led to the development of the second category, an adhesive using the self-etch strategy.

In the Se strategy (one-step or two-step), there is no need to apply a preliminary phosphoric acid gel on dental substrates as dentine demineralization and priming occur simultaneously.^{3,11} The dissolved hydroxyapatite crystals and residual smear layer are incorporated in the hybridized complex.^{3,12} Except for very acidic Se systems,^{13,14} the whole extension of the demineralized dentine depth is impregnated by resin monomers, which may be the reason why Se systems are not associated with the technique sensitivity characteristic of bonding to moist etched dentine.^{7,15,16} This advantage makes Se materials suitable for areas where adequate control of moisture is rather difficult, such as in posterior restorations.

A clear disadvantage of the Se protocol is the reduction in enamel bonding effectiveness.^{17,18} The increase in surface area in intact and ground enamel obtained with Se adhesives is lower than that achieved with phosphoric acid, and it depends on the pH of the Se adhesive.¹⁸ The performance of Se adhesives has improved when these systems were applied to phosphoric acid-treated enamel.^{12,19,20} However, this procedure has been shown to be unsuitable for use on the dentine substrate,^{21–23} because accidental dentine etching may occur during the enamel-etching process, particularly when a low-viscosity etchant is used. The effect of intentionally etching dentine with phosphoric acid prior to the application of self-etch adhesives has been studied.^{21,23–26} The results are controversial and material-dependent.

Considering the differences in professional judgement regarding the selection of the adhesive strategy and number of steps, some manufacturers have released more versatile adhesive systems that include etch-and-rinse (two step) and self-etch (one or two step) options. These new materials are called “Universal”, “Multi-purpose” or “Multi-mode” adhesives.^{23,27} There is little information in the literature about the performance of this new class of universal adhesives.^{23,27} Thus, this study compared the immediate microtensile bond strengths (μ TBS), nanoleakage (NL), *in situ* degree of conversion (DC) of three universal adhesives applied to dentine according to the etch-and-rinse and the self-etch strategies.

The two-step etch-and-rinse, Adper Single Bond 2 (SB, 3M ESPE, St. Paul, MN, USA), and two-step self-etch, Clearfil SE Bond (CSE, Kuraray, Okayama, Japan) were also evaluated as control groups. The following null hypotheses were tested in this study: (1) universal adhesives applied to dentine according to the Er and the Se strategies when compared to their respective control groups do not affect the immediate resin-dentine bond strength; (2) universal adhesives applied to dentine according to the Er and the Se strategies when compared to their respective control groups do not affect the immediate silver nitrate deposition and (3) universal adhesives applied to dentine according to the Er and the Se strategies when compared to their respective control groups do not affect the degree of conversion of the adhesives.

2. Materials and methods

2.1. Tooth selection and preparation

Forty extracted, caries-free human third molars were used. The teeth were collected after obtaining the respective patients' informed consent under a protocol approved by the local Ethics Committee Review Board. The teeth were disinfected in 0.5% chloramine, stored in distilled water and used within six months after extraction. A flat dentine surface was exposed after wet grinding the occlusal enamel on a #180 grit SiC paper. The exposed dentine surfaces were further polished on wet #600-grit silicon-carbide paper for 60 s to standardize the smear layer.

2.2. Experimental design

The teeth were randomly assigned into eight groups ($n = 5$) according to the different bonding strategies of the selected adhesive system. As control materials, the 2-step etch-and-rinse (Er), Adper Single Bond 2 (SB, 3M ESPE, St. Paul, MN, USA); and the 2-step self-etch (Se), Clearfil SE Bond (CSE, Kuraray, Okayama, Japan) were used. The following three universal adhesive systems were tested: Peak Universal Adhesive System (Peak LC Bond and Peak SE Primer, Ultradent Products Inc., South Jordan, UT, USA), applied as a 2-step Er (PkEr) and 2-step Se (PkSe); Scotchbond Universal Adhesive (3M ESPE, St. Paul, MN, USA), applied as a 2-step Er (ScEr) and 1-step Se (ScSe); and All Bond Universal (Bisco Inc., Schaumburg, IL, USA) applied as a 2-step Er (AlEr) and 1-step Se (AlSe).

2.3. Restorative procedure and specimen preparation

The adhesive systems were applied strictly in accordance with the respective manufacturer's instructions, described in Table 1. After the bonding procedures, all teeth received a micro-hybrid composite restoration (Opallis, FGM Produtos Odontológicos, Joinville, SC, Brazil) in two increments of 2 mm. Each increment was light polymerized for 40 s using a LED light curing unit set at 1200 mW/cm² (Radium-cal, SDI Limited, Bayswater, Victoria, Australia).

After the restored teeth had been stored in distilled water at 37 °C for 24 h, the specimens were sectioned longitudinally in the mesio-distal and buccal-lingual directions across the

Table 1 – Adhesive system (batch number), composition and application mode* of the adhesive systems used (*) according to the manufacturer's instructions.

Adhesive system (batch number)	Composition	Self-etch strategy	Etch-and-rinse strategy
Adper Single Bond 2 (BPBR)	<ol style="list-style-type: none"> Etchant: 35% phosphoric acid (Scotchbond Etchant) Adhesive: bis-GMA, HEMA, dimethacrylates, ethanol, water, photoinitiator, methacrylate functional copolymer of polyacrylic and poly(itaconic) acids, 10% by weight of 5 nm-diameter spherical silica particles 	N.A	<ol style="list-style-type: none"> Apply etchant for 15 s Rinse for 10 s Blot excess water Apply 2–3 consecutive coats of adhesive for 15 s with gentle agitation Gently air dry for 5 s Light polymerize for 10 s at 1200 mW/cm²
Clearfil SE Bond (Primer: 00954 A – Bond: 01416A)	<ol style="list-style-type: none"> Primer: water, MDP, HEMA, camphorquinone, hydrophilic dimethacrylate Bonding: MDP, bis-GMA, HEMA, camphorquinone, hydrophobic dimethacrylate, N,N-diethanol <i>p</i>-toluidine bond, colloidal silica 	<ol style="list-style-type: none"> Apply primer to tooth surface and leave in place for 20 s Dry with air stream to evaporate the volatile ingredients Apply bond to the tooth surface and then create a uniform film using a gentle air stream Light polymerize for 10 s at 1200 mW/cm² 	N.A
Peak Universal Adhesive System (Peak SE Primer: ON062 – Peak LC Bond: Y062)	<ol style="list-style-type: none"> Etchant: 35% phosphoric acid (Ultraetch) Peak SE Primer: ethyl alcohol, methacrylic acid, 2-hydroxyethyl methacrylate (Peak SE Primer) Peak LC Bond resin: ethyl alcohol, 2-hydroxyethyl methacrylate (Peak LC Bond) 	<ol style="list-style-type: none"> Initial use of Peak SE requires activation of the two components separated in the syringe Application of the Peak SE with microbrush for 20 s using continuous scrubbing on dentine. Do not scrub enamel Thin/dry for 3 s using air/water syringe or high volume suction directly over preparation Apply a puddle coat of Peak LC Bond and gently agitate for 10 s Thin/dry 10 s using ¼ to ½ air pressure Light polymerize for 10 s at 1200 mW/cm² 	<ol style="list-style-type: none"> Apply etchant for 20 s Rinse for 5 s Air dry 2 s Apply a puddle coat of Peak LC Bond with gentle agitation for 10 s Dry 10 s using ¼ to ½ air pressure Light polymerize for 10 s at 1200 mW/cm²
Scotchbond Universal Adhesive (D-82229)	<ol style="list-style-type: none"> Etchant: 34% phosphoric acid, water, synthetic amorphous silica, polyethylene glycol, aluminium oxide. (Scotchbond Universal Etchant) Adhesive: MDP phosphate monomer, dimethacrylate resins, HEMA, methacrylate-modified polyalkenoic acid copolymer, filler, ethanol, water, initiators, silane 	<ol style="list-style-type: none"> Apply the adhesive to the entire preparation with a microbrush and rub it in for 20 s. If necessary, rewet the disposable applicator during treatment Direct a gentle stream of air over the liquid for about 5 s until it no longer moves and the solvent has evaporated completely Light polymerize for 10 s 	<ol style="list-style-type: none"> Apply etchant for 15 s Rinse for 10 s Air dry 2 s Apply adhesive as for the self-etch mode
All-Bond Universal (1200006111)	<ol style="list-style-type: none"> Etchant Uni-Etch: 32% phosphoric acid, benzalkonium Chloride Adhesive: MDP, bis-GMA, HEMA, ethanol, water, initiators 	<ol style="list-style-type: none"> Apply two separate coats of adhesive, scrubbing the preparation with a microbrush for 10–15 s per coat. Do not light polymerize between coats Evaporate excess solvent by thoroughly air-drying with an air syringe for at least 10 s, there should be no visible movement of the material. The surface should have a uniform glossy appearance Light polymerize for 10 s at 1200 mW/cm² 	<ol style="list-style-type: none"> Apply etchant for 15 s Rinse thoroughly Remove excess water with absorbent pellet or high volume suction for 1–2 s Apply adhesive as for the self-etch mode

bonded interface, using a slow-speed diamond saw (Isomet, Buehler Ltd., Lake Bluff, IL, USA) to obtain 15–20 resin–denture sticks with a cross sectional area of approximately 0.8 mm² measured with a digital calliper (Digimatic Calliper, Mitutoyo, Tokyo, Japan).

All the specimens from each tooth were used for the microtensile bond strength evaluation (μ TBS), except for six that were randomly selected. These six resin–denture bonded specimens were divided for measurement of the *in situ* degree of conversion (DC) and nanoleakage (NL).

2.4. Microtensile bond strength test (μ TBS)

Resin–denture bonded sticks were attached to a Geraldeli's jig²⁸ with cyanoacrylate adhesive and tested under tension (Kratos Dinamometros, Cotia, SP, Brazil) at 0.5 mm/min until failure. The μ TBS values were calculated by dividing the load at failure by the cross-sectional bonding area.

The failure mode of the specimens was classified as cohesive ([C] failure exclusive within denture or resin composite), adhesive ([A] failure at resin/denture interface), or mixed ([M] failure at resin/denture interface, which included cohesive failure of the neighbouring substrates). The classification was performed under a stereomicroscope at 100 \times magnification (Olympus SZ40, Tokyo, Japan). Specimens with premature failures (PF) were included in the tooth mean.

2.5. Degree of conversion *in situ* (DC)

Three resin–denture bonded sticks from each tooth were wet polished with #1500, 2000 and 2500-grit SiC paper. Then they were ultrasonically cleaned for 20 min and stored in water at 37 °C for 24 h, before taking the DC readings. The micro-Raman spectroscopy analysis was performed using the Senterra equipment (Bruker Optik GmbH, Ettlingen, Baden-Württemberg, Germany). The micro-Raman spectrometer was first calibrated by resetting to zero, and then for coefficient values using a silicon specimen. Specimens were analyzed using the following micro-Raman parameters: 20 mW Neon laser with 532 nm wavelength, spatial resolution of \approx 3 μ m, spectral resolution \approx 5 cm⁻¹, accumulation time of 30 s with 6 co-additions, and 110 \times magnification (Olympus UK, London, UK) to a \approx 1 μ m beam diameter. Spectra were taken at the denture–adhesive interface at three different sites within intertubular denture for each resin–denture bonded stick. The average value of the measurements taken from the same tooth was used for statistical purposes. Spectra of unpolymerized adhesives were taken as reference. Post-processing of spectra was performed using the dedicated Opus Spectroscopy Software version 6.5. The ratio of double-bond content of monomer to polymer in the adhesive was calculated according to the following formula:

$$DC (\%) = \left(1 - \frac{R_{(cured)}}{R_{(uncured)}} \right) \times 100$$

where “R” is the ratio of aliphatic and aromatic peak areas at 1639 cm⁻¹ and 1609 cm⁻¹ in polymerized and unpolymerized adhesives. The *in situ* DC of all resin–denture bonded sticks from the same tooth was averaged for statistical purposes.

2.6. Nanoleakage (NL) evaluation

Three resin-bonded sticks, from each tooth, were used for NL evaluation. Ammoniacal silver nitrate was prepared according to the protocol previously described by Tay et al.²⁹ The sticks were placed in the ammoniacal silver nitrate solution in darkness for 24 h, rinsed thoroughly in distilled water, and immersed in photo developing solution for 8 h under a fluorescent light to reduce silver ions into metallic silver grains within voids along the bonded interface. Specimens were polished with a wet #600, 1000, 1200, 1500, 2000 and 2500-grit SiC paper and 1 and 0.25 μ m diamond paste (Buehler Ltd., Lake Bluff, IL, USA) using a polishing cloth. They were ultrasonically cleaned, air dried, mounted on stubs, and coated with carbon-gold (MED 010, Balzers Union, Balzers, Liechtenstein). Resin–denture interfaces were analyzed in a field-emission scanning electron microscope operated in the backscattered mode (LEO 435 VP, LEO Electron Microscopy Ltd., Cambridge, UK).

Three images were captured of each resin–denture bonded stick. The relative percentage of NL within the adhesive and hybrid layers in each specimen was measured in all images using the UTHSCSA ImageTool 3.0 software (Department of Dental Diagnostic Science at The University of Texas Health Science Centre, San Antonio, Texas) by a blinded researcher. The values originating from the same specimen were averaged for statistical purposes. The mean NL of all sticks from the same tooth was averaged for statistical purposes.

2.7. pH measurement

The pH of the adhesive systems was measured with a pH metre E520 (Metrohm, Herisau, Switzerland). Three blends were prepared by mixing each adhesive solution and deionized water in a proportion of 1:9 (by weight). This blend was kept stored in darkness for 12 h to allow uptake of H⁺. For each blend, three readings were performed at room temperature, in darkness and a mean was obtained.

2.8. Statistical analysis

Data from μ TBS, NL, DC and pH were analyzed by one-way ANOVA and Tukey's *post hoc* test at $\alpha = 0.05$.

3. Results

3.1. Microtensile bond strength test

The overall values of all tested properties are shown in Table 2. Significant differences were detected in the mean μ TBS among the adhesives tested ($p < 0.05$). Only the universal adhesive PK, used in both the Er and Se approach, showed mean μ TBS statistically similar to those of the controls SB and CSE respectively ($p > 0.05$). The other universal materials resulted in lower mean μ TBS than those of the Er and Se controls ($p < 0.05$). The lowest mean μ TBS was observed for AISE ($p < 0.05$).

The majority of the specimens (83.1%) showed adhesive/mixed failures. Cohesive failures were observed in 8.3% of the

Table 2 – Microtensile bond strength (μ TBS), nanoleakage (NL) and degree of conversion (DC) values (means \pm standard deviations) of the different experimental groups.

Adhesive system	Application mode	Test		
		μ TBS (MPa)	NL (%)	DC (%)
Adper Single Bond 2	Etch-and-rinse	49.3 \pm 4.6A	12.4 \pm 2.5b	85.4 \pm 4.7 ^a
Clearfil SE Bond	Self-etch	43.0 \pm 4.5A,B	7.6 \pm 2.a	87.7 \pm 2.8 ^a
Peak Universal Adhesive System	Etch-and-rinse	43.6 \pm 4.6A,B	23.4 \pm 5.9c	89.2 \pm 6.3 ^a
	Self-etch	39.9 \pm 4.5B,C	34.4 \pm 11c	78.9 \pm 9.5 ^a
Scotchbond Universal Adhesive	Etch-and-rinse	35.1 \pm 6.6B,C	5.1 \pm 2.5a	88.3 \pm 5.6 ^a
	Self-etch	32.4 \pm 4.5C	5.5 \pm 3.2a	69.1 \pm 9.8 ^b
All Bond Universal	Etch-and-rinse	39.3 \pm 3.7B	9.3 \pm 2.9b	77.9 \pm 0.1 ^a
	Self-etch	13.4 \pm 1.9D	6.2 \pm 4.1a	77.8 \pm 0.1 ^a

* Similar capital (μ TBS), lower (NL) and superscript letter (DC) are not statistically significant ($p < 0.05$)

Table 3 – Number and percentage of specimens (%) according to the fracture mode and premature failures of all experimental group.

Adhesive system	Application mode	Fracture pattern			
		A	C	A/M	PF
Adper Single Bond 2	Etch-and-rinse	50 (56.8)	18 (20.5)	8 (9.1)	12 (13.6)
Clearfil SE Bond	Self-etch	66 (76.7)	4 (4.7)	8 (9.3)	8 (9.3)
Peak Universal Adhesive System	Etch-and-rinse	60 (68.2)	6 (6.8)	12 (13.6)	10 (11.4)
	Self-etch	56 (66.7)	8 (9.5)	12 (14.3)	8 (9.5)
Scotchbond Universal Adhesive	Etch-and-rinse	64 (82.1)	0 (0)	12 (15.4)	2 (2.5)
	Self-etch	58 (76.3)	4 (5.3)	12 (15.8)	2 (2.6)
All Bond Universal	Etch-and-rinse	46 (60.5)	10 (13.2)	16 (21.1)	4 (5.2)
	Self-etch	52 (66.7)	0 (0)	20 (25.6)	6 (7.7)

specimens. A small number of premature failures (8.6%) were observed in the present study (Table 3).

3.2. Nanoleakage evaluation

Significant differences were observed among the adhesives tested ($p = 0.0001$; Table 2 and Fig. 1). PKEr and PKSe adhesives showed the highest percentage of NL ($p < 0.05$). AlEr and AlSe adhesives were statistically similar to their respective control groups (SB and CSE; $p > 0.05$). Scotchbond Universal (ScEr and ScSe) showed the lowest nanoleakage ($p < 0.05$); in the Er approach, the NL was lower than that of SB; and in the Se approach NL was similar to that of CSE.

3.3. Degree of conversion

All materials showed similar DC ($p > 0.05$, Table 2), except for ScSe, which showed a statistically lower DC compared with the other materials ($p = 0.01$).

3.4. pH measurement

The pH of the adhesives is shown in Table 4. SB showed the highest pH (4.1) and the Peak LC Bond showed the lowest (1.2).

4. Discussion

Although all the new universal adhesive systems tested share the same versatility of being used in the Er and Se approaches,

there are differences in their compositions and number of steps, which seems to be the key reason for the different performance of these materials when compared with the controls.

The 2-step Se CSE and, the 1-step Se ScSe and AlSe have a pH within a similar range (around 2). Se adhesives within this pH range partially demineralize dentine, leaving a substantial amount of hydroxyapatite crystals around the collagen fibrils.³⁰ As these three materials have 10-methacryloyloxydecyl dihydrogen phosphate monomer (MDP) in their composition, they bond chemically to dentine.^{31–33} Yoshida et al.³⁴ showed that an effective chemical interaction occurs between MDP and hydroxyapatite forming a stable nano-layer that could form a stronger phase at the adhesive interface, which increases the mechanical strength of the adhesive interface. In addition, stable MDP-Ca salt deposition along with nano-layering may explain the high bond stability,^{34,35} which has previously been proven both in laboratory and clinical research.^{33,36,37}

Based on these similarities one would expect that these two universal adhesives (ScSe and AlSe) would be equivalent to the control 2-step Se CSE; however, this was not observed in the present investigation. The slight difference in terms of μ TBS between CSE and ScSe may be due to the presence of polyalkenoic acid copolymer in the composition of ScSe and this leads to partially reject the 1st. null hypothesis. This copolymer may have competed with the MDP by binding to the calcium of the hydroxyapatite.³⁴ In addition to impairing the bond of MDP to dentine, the polyalkenoic acid copolymer could have prevented

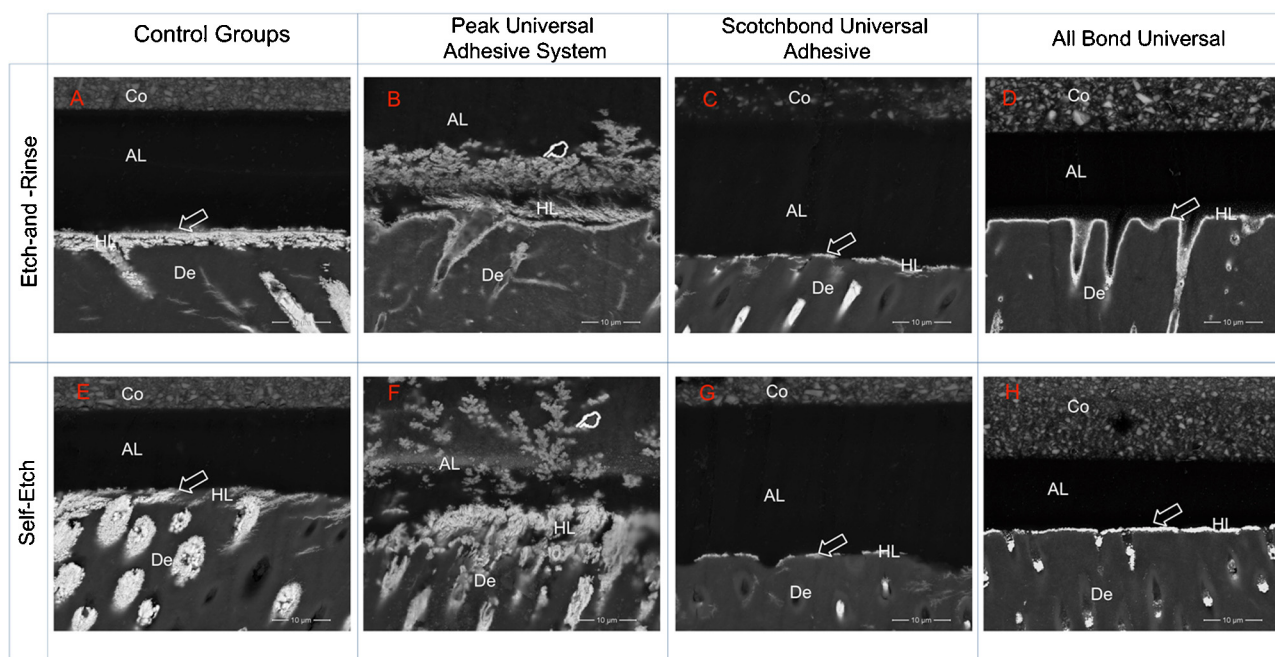


Fig. 1 – Representative backscattered SEM images of the resin–dentine adhesive interfaces of each experimental group (Control etch-and-rinse group = Adper Single Bond 2 and control self-etch group = Clearfil SE Bond). The amount of nanoleakage was lower and practically occurred within the hybrid layer for Scotchbond Universal (C, G); All Bond Universal (D, H) and controls (white arrows) (A, E). For Peak Universal Adhesive System, the amount of nanoleakage was higher than the other materials; with most silver nitrate uptake occurring throughout the entire thickness of the HL (white arrows) and in the AL, forming the so-called “water trees (white hands) (B, F).

monomer approximation during polymerization, due to its high molecular weight. Consequently a lower DC was observed for this material in comparison with the others leads to partially reject the 3rd. null hypothesis. Furthermore, ScSe and ALSe are 1-step self-etch adhesives and this probably makes the MDP concentration lower than that of CSE, which has this monomer incorporated into both the primer and bond components.³⁴

Among all Se adhesives, ALSe showed the poorest performance in terms of μ TBS, which might be attributed to its application mode. ALSe was the only material that was not applied actively on dentine, as the materials were applied in accordance with the respective manufacturer’s directions. Previous literature findings clearly pointed out that active application improves the bonding performance

of Se adhesive systems to dentine.^{38,39} Future studies need to be conducted to test this hypothesis.

Among adhesives that are used with the Se strategy, PKSe provided μ TBS values similar to those of the control CSE. The placement of an additional layer has been shown to increase the performance of 1-step Se materials both *in vitro*^{37,40-42} and clinically.^{43,44} This is also a disadvantage of ScSe and ALSe. In general, 1-step SE adhesives present inferior performance to 2-step Se adhesives.^{45,46} Usually, 1-step Se adhesives result in thinner adhesive layers, which are likely to be prone to polymerization inhibition by oxygen.⁴⁷ With regard to the Er approach, the universal adhesives showed improved bond strength in comparison with the Se strategy, in spite of ScPK being statistically similar to the ScSe. It is known that the smear layer constitutes a true physical barrier and makes it extremely difficult for the bonding and hybrid layer formation to be fully integrated with the dentine.⁴⁸ After preliminary etching with phosphoric acid, the smear layer is removed and superficial dentine is demineralized. This increases impregnation by the adhesive, allowing the creation of a well impregnated hybrid layer, without modifying the potential for interfacial nanoleakage, as has been shown by several authors when a 1-step self-etch adhesive was applied on phosphoric acid etched dentine.^{23,25-27}

NL represents the location of defects at the resin–dentine interface that might serve as the pathway for degradation of resin/dentine bonds over time.⁴⁹ Silver

Table 4 – The pH values (means \pm standard deviations) of each adhesive system.

Adhesive system		pH
Adper Single Bond 2		4.1 \pm 0.02
Clearfil SE Bond	Primer	2.1 \pm 0.01
	Bond	2.6 \pm 0.08
Peak Universal Adhesive System	Peak SE Primer	1.2 \pm 0.01
	Peak LC Bond resin	2.0 \pm 0.04
Scotchbond Universal Adhesive		3.0 \pm 0.05
All Bond Universal		2.4 \pm 0.05

nitrate is capable of occupying the nanometer-sized spaces around naked collagen fibrils, where resin failed to infiltrate, or where residual water has not been displaced by adhesive or even in areas of incomplete monomer conversion.⁴⁹

Among all the adhesives, PKEr and PKSe showed the highest NL at the adhesive interfaces, and this leads to partially reject the 2nd. null hypothesis. In the case of PkSe, Peak SE primer has a low pH value (pH = 1.2), which is within the range of the most aggressive Se adhesives.^{30,45} It has previously been reported that unpolymerized, acidic and aggressive monomers from Se adhesives are able to continue to demineralize the dentine even after polymerization.^{13,14} This is probably the result of hydrolysis of the ester bond from the acid monomer that results in a strong phosphoric acid¹⁴ and might explain the intense silver nitrate deposition within the hybrid layer.

Although one could expect reduced NL within the adhesive layer due to the fact that PKSe is a 2-step Se, the second adhesive layer seems to be as hydrophilic as the primer, since no hydrophobic monomer was listed in the composition of the Peak LC Bond (Table 1). Thus, although increased impregnation of resin monomers might have occurred during the application of an additional adhesive coat, the nature of this impregnation is hydrophilic, which resulted in extensive NL in the hybrid layer. In fact, Peak LC bond (pH = 2.0) may cause an additional etching and an increased conditioning pattern of the dentine substrate. This is likely to result in increased demineralization and production of a hydroxyapatite-denuded, collagen-rich, network,^{45,50,51} increasing the risk of NL^{10,52–54} for PkEr.

Taking all the properties together, none of the materials showed similar behaviour to the controls (a gold standard two-step self-etch Clearfil SE Bond and a gold standard two-step etch-and-rinse Adper Single Bond 2) leads to reject all null hypothesis tested. Future studies need to be conducted to evaluate the long-term performance of this new category of adhesives.

5. Conclusions

Within the limitations of the present study, the results indicate that when the universal adhesives were tested using the self-etch or etch-and-rinse strategy on dentine, they were inferior to the respective controls (Clearfil SE Bond, a two-step etch-and-rinse or Adper Single Bond 2, a two-step etch-and-rinse) with respect to at least one of the properties tested (microtensile bond strength, nanoleakage and *in situ* degree of conversion).

Acknowledgements

This study was partially supported by the National Council for Scientific and Technological Development (CNPq) (grants # 301937/2009-5 and # 301891/2010-9). The authors would like to thank to the manufacturers for the donation of adhesive systems employed in this study.

REFERENCES

1. Nakabayashi N, Kojima K, Masuhara E. The promotion of adhesion by the infiltration of monomers into tooth substrates. *Journal of Biomedical Materials Research* 1982;16:265–73.
2. Van Meerbeek B, Perdigao J, Lambrechts P, Vanherle G. The clinical performance of adhesives. *Journal of Dentistry* 1998;26:1–20.
3. De Munck J, Van Landuyt K, Peumans M, Poitevin A, Lambrechts P, Braem M, et al. A critical review of the durability of adhesion to tooth tissue: methods and results. *Journal of Dental Research* 2005;84:118–32.
4. Van Meerbeek B, Inokoshi S, Braem M, Lambrechts P, Vanherle G. Morphological aspects of the resin–dentin interdiffusion zone with different dentin adhesive systems. *Journal of Dental Research* 1992;71:1530–40.
5. Spencer P, Swafford JR. Unprotected protein at the dentin–adhesive interface. *Quintessence International* 1999;30:501–7.
6. Tay FR, Gwinnett JA, Wei SH. Micromorphological spectrum from overdrying to overwetting acid-conditioned dentin in water-free acetone-based, single-bottle primer/adhesives. *Dental Materials* 1996;12:236–44.
7. Reis A, Loguercio AD, Azevedo CL, de Carvalho RM, da Julio Singer M, Grande RH. Moisture spectrum of demineralized dentin for adhesive systems with different solvent bases. *Journal of Adhesive Dentistry* 2003;5:183–92.
8. Sano H, Shono T, Takatsu T, Hosoda H. Microporous dentin zone beneath resin-impregnated layer. *Operative Dentistry* 1994;19:59–64.
9. Burrow MF, Satoh M, Tagami J. Dentin bond durability after three years using a dentin bonding agent with and without priming. *Dental Materials* 1996;12:302–7.
10. Hashimoto M, Ohno H, Kaga M, Endo K, Sano H, Oguchi H. In vivo degradation of resin–dentin bonds in humans over 1 to 3 years. *Journal of Dental Research* 2000;79:1385–91.
11. Miyazaki M, Onose H, Moore BK. Analysis of the dentin–resin interface by use of laser Raman spectroscopy. *Dental Materials* 2002;18:576–80.
12. Van Meerbeek B, De Munck J, Mattar D, Van Landuyt K, Lambrechts P. Microtensile bond strengths of an etch&rinse and self-etch adhesive to enamel and dentin as a function of surface treatment. *Operative Dentistry* 2003;28:647–60.
13. Carvalho RM, Chersoni S, Frankenberger R, Pashley DH, Prati C, Tay FR. A challenge to the conventional wisdom that simultaneous etching and resin infiltration always occurs in self-etch adhesives. *Biomaterials* 2005;26:1035–42.
14. Wang Y, Spencer P. Continuing etching of an all-in-one adhesive in wet dentin tubules. *Journal of Dental Research* 2005;84:350–4.
15. Tay FR, Gwinnett JA, Wei SH. Relation between water content in acetone/alcohol-based primer and interfacial ultrastructure. *Journal of Dentistry* 1998;26:147–56.
16. Perdigao J. Dentin bonding-variables related to the clinical situation and the substrate treatment. *Dental Materials* 2010;26:e24–37.
17. Kanemura N, Sano H, Tagami J. Tensile bond strength to and SEM evaluation of ground and intact enamel surfaces. *Journal of Dentistry* 1999;27:523–30.
18. Pashley DH, Tay FR. Aggressiveness of contemporary self-etching adhesives part II: etching effects on unground enamel. *Dental Materials* 2001;17:430–44.
19. Rotta M, Bresciani P, Moura SK, Grande RH, Hilgert LA, Baratieri LN, et al. Effects of phosphoric acid pretreatment and substitution of bonding resin on bonding effectiveness of self-etching systems to enamel. *Journal of Adhesive Dentistry* 2007;9:537–45.

20. Frankenberger R, Lohbauer U, Roggendorf MJ, Naumann M, Taschner M. Selective enamel etching reconsidered: better than etch-and-rinse and self-etch? *Journal of Adhesive Dentistry* 2008;**10**:339–44.
21. Van Landuyt KL, Peumans M, De Munck J, Lambrechts P, Van Meerbeek B. Extension of a one-step self-etch adhesive into a multi-step adhesive. *Dental Materials* 2006;**22**:533–44.
22. Van Landuyt KL, Kanumilli P, De Munck J, Peumans M, Lambrechts P, Van Meerbeek B. Bond strength of a mild self-etch adhesive with and without prior acid-etching. *Journal of Dentistry* 2006;**34**:77–85.
23. Hanabusa M, Mine A, Kuboki T, Momoi Y, Van Ende A, Van Meerbeek B, et al. Bonding effectiveness of a new ‘multi-mode’ adhesive to enamel and dentine. *Journal of Dentistry* 2012.
24. Erhardt MC, Osorio E, Aguilera FS, Proenca JP, Osorio R, Toledano M. Influence of dentin acid-etching and NaOCl-treatment on bond strengths of self-etch adhesives. *American Journal of Dentistry* 2008;**21**:44–8.
25. Proenca JP, Polido M, Osorio E, Erhardt MC, Aguilera FS, Garcia-Godoy F, et al. Dentin regional bond strength of self-etch and total-etch adhesive systems. *Dental Materials* 2007;**23**:1542–8.
26. Taschner M, Nato F, Mazzoni A, Frankenberger R, Falconi M, Petschelt A, et al. Influence of preliminary etching on the stability of bonds created by one-step self-etch bonding systems. *European Journal of Oral Sciences* 2012;**120**:239–48.
27. Perdigao J, Sezinando A, Monteiro PC. Laboratory bonding ability of a multi-purpose dentin adhesive. *American Journal of Dentistry* 2012;**25**:153–8.
28. Perdigao J, Geraldini S, Carmo AR, Dutra HR. In vivo influence of residual moisture on microtensile bond strengths of one-bottle adhesives. *Journal of Esthetic and Restorative Dentistry* 2002;**14**:31–8.
29. Tay FR, Pashley DH, Suh BI, Carvalho RM, Itthagarun A. Single-step adhesives are permeable membranes. *Journal of Dentistry* 2002;**30**:371–82.
30. Tay FR, Pashley DH. Aggressiveness of contemporary self-etching systems I: depth of penetration beyond dentin smear layers. *Dental Materials* 2001;**17**:296–308.
31. Inoue S, Koshiro K, Yoshida Y, De Munck J, Nagakane K, Suzuki K, et al. Hydrolytic stability of self-etch adhesives bonded to dentin. *Journal of Dental Research* 2005;**84**:1160–4.
32. Waidyasekera K, Nikaido T, Weerasinghe DS, Ichinose S, Tagami J. Reinforcement of dentin in self-etch adhesive technology: a new concept. *Journal of Dentistry* 2009;**37**:604–9.
33. Peumans M, De Munck J, Van Landuyt KL, Poitevin A, Lambrechts P, Van Meerbeek B. Eight-year clinical evaluation of a 2-step self-etch adhesive with and without selective enamel etching. *Dental Materials* 2010;**26**:1176–84.
34. Yoshida Y, Yoshihara K, Nagaoka N, Hayakawa S, Torii Y, Ogawa T, et al. Self-assembled nano-layering at the adhesive interface. *Journal of Dental Research* 2012;**91**:376–81.
35. Yoshihara K, Yoshida Y, Nagaoka N, Fukegawa D, Hayakawa S, Mine A, et al. Nano-controlled molecular interaction at adhesive interfaces for hard tissue reconstruction. *Acta Biomaterialia* 2010;**6**:3573–82.
36. Toledano M, Osorio R, Osorio E, Aguilera FS, Yamauti M, Pashley DH, et al. Durability of resin–dentin bonds: effects of direct/indirect exposure and storage media. *Dental Materials* 2007;**23**:885–92.
37. Feitosa VP, Leme AA, Sauro S, Correr-Sobrinho L, Watson TF, Sinhoreti MA, et al. Hydrolytic degradation of the resin–dentine interface induced by the simulated pulpal pressure, direct and indirect water ageing. *Journal of Dentistry* 2012.
38. Pleffken PR, de Almeida Lourenco AP, Torres CR, Buhler Borges A. Influence of application methods of self-etching adhesive systems on adhesive bond strength to dentin. *Journal of Adhesive Dentistry* 2011;**13**:517–25.
39. Loguercio AD, Stanislawczuk R, Mena-Serrano A, Reis A. Effect of 3-year water storage on the performance of one-step self-etch adhesives applied actively on dentine. *Journal of Dentistry* 2011;**39**:578–87.
40. Frankenberger R, Perdigao J, Rosa BT, Lopes M. No-bottle” vs “multi-bottle” dentin adhesives—a microtensile bond strength and morphological study. *Dental Materials* 2001;**17**:373–80.
41. Albuquerque M, Pegoraro M, Mattei G, Reis A, Loguercio AD. Effect of double-application or the application of a hydrophobic layer for improved efficacy of one-step self-etch systems in enamel and dentin. *Operative Dentistry* 2008;**33**:564–70.
42. Reis A, Albuquerque M, Pegoraro M, Mattei G, Bauer JR, Grande RH, et al. Can the durability of one-step self-etch adhesives be improved by double application or by an extra layer of hydrophobic resin? *Journal of Dentistry* 2008;**36**:309–15.
43. Loguercio AD, Reis A. Application of a dental adhesive using the self-etch and etch-and-rinse approaches: an 18-month clinical evaluation. *Journal of American Dental Association* 2008;**139**:53–61.
44. Reis A, Leite TM, Matte K, Michels R, Amaral RC, Geraldini S, et al. Improving clinical retention of one-step self-etching adhesive systems with an additional hydrophobic adhesive layer. *Journal of American Dental Association* 2009;**140**:877–85.
45. Van Meerbeek B, De Munck J, Yoshida Y, Inoue S, Vargas M, Vijay P, et al. Buonocore memorial lecture adhesion to enamel and dentin: current status and future challenges. *Operative Dentistry* 2003;**28**:215–35.
46. Perdigao J. New developments in dental adhesion. *Dental Clinics of North America* 2007;**51**:333–57. viii.
47. Nunes TG, Ceballos L, Osorio R, Toledano M. Spatially resolved photopolymerization kinetics and oxygen inhibition in dental adhesives. *Biomaterials* 2005;**26**:1809–17.
48. Kenschima S, Reis A, Uceda-Gomez N, Tancredo Lde L, Filho LE, Nogueira FN, et al. Effect of smear layer thickness and pH of self-etching adhesive systems on the bond strength and gap formation to dentin. *Journal of Adhesive Dentistry* 2005;**7**:117–26.
49. Sano H. Microtensile testing, nanoleakage, and biodegradation of resin–dentin bonds. *Journal of Dental Research* 2006;**85**:11–4.
50. Liu Y, Tjaderhane L, Breschi L, Mazzoni A, Li N, Mao J, et al. Limitations in bonding to dentin and experimental strategies to prevent bond degradation. *Journal of Dental Research* 2011;**90**:953–68.
51. Brackett MG, Li N, Brackett WW, Sword RJ, Qi YP, Niu LN, et al. The critical barrier to progress in dentine bonding with the etch-and-rinse technique. *Journal of Dentistry* 2011;**39**:238–48.
52. Wang Y, Spencer P. Quantifying adhesive penetration in adhesive/dentin interface using confocal Raman microspectroscopy. *Journal of Biomedical Materials Research* 2002;**59**:46–55.
53. Wang Y, Spencer P. Hybridization efficiency of the adhesive/dentin interface with wet bonding. *Journal of Dental Research* 2003;**82**:141–5.
54. Spencer P, Wang Y, Walker MP, Wieliczka DM, Swafford JR. Interfacial chemistry of the dentin/adhesive bond. *Journal of Dental Research* 2000;**79**:1458–63.

4.2 Capítulo 2 (do experimento 2)

LONGEVITY OF BONDING PROPERTIES OF UNIVERSAL ADHESIVES TO DENTIN

OPERATIVE DENTISTRY

ONLINE MANUSCRIPT SUBMISSION AND PEER REVIEW

[AUTHOR INSTRUCTIONS](#) [REVIEWER INSTRUCTIONS](#) [ONLINE JOURNAL](#)

To ensure proper functionality of this site, both JavaScript and Cookies must be enabled.

[HOME](#)

Detailed Status Information

Manuscript #	14-055-L
Current Revision #	0
Submission Date	2014-02-20
Current Stage	Potential Reviewers Assigned
Title	Longevity of Bonding Properties of Universal Adhesives to Dentin
Running Title	Longevity of f Universal Adhesives to Dentin
Manuscript Type	Laboratory Research
Corresponding Author	Alessandro Loguerio (Ponta Grossa State University)
Contributing Authors	Miguel Muñoz , Issis Luque-Martinez , Pamela Malaquias , Viviane Hass , Alessandra Reis , Nara Campanha
Financial Disclosure	I have no relevant financial interests in this manuscript.
Abstract	<p>Objectives: To evaluate the dentine microtensile bond strength (μTBS) and nanoleakage (NL) for etch-and-rinse and self-etch strategies of universal adhesive systems in the immediate time (IM) and after 6 months of storage (6M). Methods: forty caries-free extracted third molars were divided into 8 groups for μTBS (n = 5), according to the adhesive and etching strategy: Clearfil SE Bond [CSE] and Adper Single Bond 2 [SB], as controls; Peak Universal, self-etch [PkSe] and etch-and-rinse [PkEr]; Scotchbond Universal Adhesive, self-etch [ScSe] and etch-and-rinse [ScEr]; All Bond Universal, self-etch [AlSe] and etch-and-rinse [AlEr]. After composite restorations, specimens were stored in water (37°C/24 h) and longitudinally sectioned to obtain resin-dentine bonded sticks (0.8 mm²). Half of the sticks were tested under tension at 0.5 mm/min in the IM and the other half in 6M. Some sticks from each tooth group and time evaluations were used for nanoleakage evaluation (NL). Data were analyzed with two-way repeated measure ANOVA and Tukey's test ($\alpha = 0.05$). Results: Despite higher μTBS in the immediate time, PkSe and PkEr, similar to the respective control groups (IM; $p > 0.05$), these strategies showed higher NL pattern and lower μTBS after 6M ($p < 0.05$). ScSe and ScEr showed intermediary μTBS values, but it was maintained after 6 months ($p > 0.05$). For Al, AlSe showed the lowest μTBS mean ($p < 0.05$), but μTBS and NL values did not degrade ($p > 0.05$). However, AlEr, despite a higher immediate μTBS mean, showed signs of degradation after 6 months ($p < 0.05$). Conclusions: Universal adhesives that contain MDP showed better results after 6 months of water storage.</p>
Associate Editor	Assigned
Key Words	Microtensile bond strength, nanoleakage, degree of conversion, etch-and-rinse, self-etch,

1 Longevity of Bonding Properties of Universal Adhesives to Dentin

2 Abstract

3 Objectives: To evaluate the dentine microtensile bond strength (μ TBS) and
4 nanoleakage (NL) for etch-and-rinse and self-etch strategies of universal adhesive
5 systems in the immediate time (IM) and after 6 months of storage (6M). Methods:
6 forty caries-free extracted third molars were divided into 8 groups for μ TBS (n = 5),
7 according to the adhesive and etching strategy: Clearfil SE Bond [CSE] and Adper
8 Single Bond 2 [SB], as controls; Peak Universal, self-etch [PkSe] and etch-and rinse
9 [PkEr]; Scotchbond Universal Adhesive, self-etch [ScSe] and etch-and-rinse [ScEr];
10 All Bond Universal, self-etch [AlSe] and etch-and-rinse [AlEr]. After composite
11 restorations, specimens were stored in water (37°C/24 h) and longitudinally sectioned
12 to obtain resin–dentine bonded sticks (0.8 mm²). Half of the sticks were tested under
13 tension at 0.5 mm/min in the IM and the other half in 6M. Some sticks from each tooth
14 group and time evaluations were used for nanoleakage evaluation (NL). Data were
15 analyzed with two-way repeated measure ANOVA and Tukey's test ($\alpha = 0.05$).
16 Results: Despite higher μ TBS in the immediate time, PkSe and PkEr, similar to the
17 respective control groups (IM; $p > 0.05$), these strategies showed higher NL pattern
18 and lower μ TBS after 6M ($p < 0.05$). ScSe and ScEr showed intermediary μ TBS
19 values, but it was maintained after 6 months ($p > 0.05$). For Al, AlSe showed the lowest
20 μ TBS mean ($p < 0.05$), but μ TBS and NL values did not degrade ($p > 0.05$). However,
21 AlEr, despite a higher immediate μ TBS mean, showed signs of degradation after 6
22 months ($p < 0.05$). Conclusions: Universal adhesives that contain MDP showed better
23 results after 6 months of water storage.

24 Keywords: Microtensile bond strength; nanoleakage; degree of conversion; etch-and-
25 rinse; self-etch; universal adhesive systems

26

27 **Introduction**

28 Current adhesive materials simplify the bonding procedures by reducing the number
29 of application steps/time required for application. This simpler protocol makes them
30 less technique-sensitive and allows for better application standardization.¹ This is the
31 possibility some of the factors responsible for the large increase in the use of self-etch
32 adhesives among clinicians.²

33 Self-etch materials (also known as non-rinsing adhesives or etch-and-dry
34 adhesives) do not require a separate acid step as demineralization and priming
35 occurs simultaneously.³ The preliminary use of phosphoric acid increases the
36 probability of clinical errors due to the need of rinsing and adequate management of
37 dentin moisture.⁴ Contrary to the etch-and-rinse approach, self-etch adhesives do not
38 remove but incorporate the smear layer in the hybridized complex. Although a
39 complete resin infiltration through is not observed for some acidic self-etch systems,⁵
40 ⁶ some studies report lower incidence of post-operative sensitivity after placement of
41 direct composite posterior restorations.⁷

42 On the other hand, some drawbacks may be listed for these self-etch
43 materials. They do not produce an enamel conditioning pattern as retentive as that
44 produced by phosphoric acid,^{8, 9} which is likely responsible for the higher rates of
45 marginal discoloration in the enamel margins of cervical restorations.¹⁰ Selective
46 enamel etching on the enamel margins with phosphoric acid is the most current
47 accepted technique to solve this problem, showing good results either in vitro¹¹⁻¹³ and
48 in vivo studies.¹⁴

49 One clinical problem of this technique is that accidental dentin etching may

50 occur, particularly in small cavities¹⁵ or when a low-viscosity acid is used.¹⁶ The
51 preliminary etching of dentin with phosphoric acid before application of self-etch
52 application shows controversial results and seems to be material-dependent.¹⁵⁻¹⁸ For
53 one-step self-etch materials, the previous phosphoric acid etching can increase the
54 bond strength of these adhesives to dentin.^{15, 17, 18}

55 Based on that, one may conclude that clinical application of the adhesive
56 systems is limited not only by the bonding strategy, but also by their number of clinical
57 steps. Although simplified adhesives (2-step ER and 1-step SE) are suitable for direct
58 clinical procedures due to their fast and easy application, they may not be adequate
59 in luting procedures when chemically-cured cements are employed, in the face of the
60 possibility of chemical and physical incompatibilities between the adhesive system
61 and the luting materials.^{19, 20}

62 Keeping this concept in mind, a novel family of bonding systems known as
63 “universal” or “multi-mode” adhesives^{21, 22} was recently launched in the market. They
64 are one-step self-etch adhesives that can be associated or not with phosphoric acid
65 etching, which gives the dentist a more versatile adhesive system.²³ The main
66 difference between the oldest self-etch systems and these newest ones is the
67 presence of monomers capable to produce chemical adhesion to the dental
68 substrates.^{21, 22} This is expected to increase the durability of the bonds produced with
69 simplified self-etch adhesive, which has been considered less durable in in vitro and
70 in vivo studies.^{10, 24} To the extent of our knowledge, the longevity of bonds produced
71 by universal adhesives is still scarce in the literature,²⁵ as well as their immediate
72 bonding properties.^{16, 21-23}

73 Thus, the aim of this study was to evaluate the immediate and 6-month

74 microtensile bond strength (μ TBS) and nanoleakage (NL) of universal adhesive
75 systems used in the etch-and-rinse and self-etch approaches. The following null
76 hypotheses were tested in this study: (1) the immediate and 6-month μ TBS and
77 nanoleakage of universal adhesives used as etch-and-rinse or self-etch or will be
78 similar.

79 **Materials and methods**

80 Tooth selection and preparation

81 Forty extracted caries-free human third molars were used. The teeth were
82 collected after obtaining the respective patients' informed consent under a protocol
83 approved by the local Ethics Committee Review Board. The teeth were disinfected in
84 0.5% chloramine, stored in distilled water, and used within six months after extraction.
85 A flat dentine surface was exposed after wet-grinding the occlusal enamel on a #180-
86 grit SiC paper. The exposed dentine surfaces were further polished on wet #600-grit
87 silicon-carbide paper for 60 s to standardize the smear layer.

88 Experimental design

89 The teeth were randomly assigned into eight groups ($n = 5$) according to the
90 different bonding strategies of the selected adhesive system. As control materials, the
91 2-step etch-and-rinse (Er), Adper Single Bond 2 (SB, 3M ESPE, St. Paul, MN, USA);
92 and the 2-step self-etch (Se), Clearfil SE Bond (CSE, Kuraray, Okayama, Japan)
93 were used. The following three universal adhesive systems were tested: Peak
94 Universal Adhesive System (Peak LC Bond and Peak SE Primer Ultradent Products
95 Inc., South Jordan, UT, USA), applied as a 2-step Er (PkEr) and 2-step Se (PkSe);
96 Scotchbond Universal Adhesive (3M ESPE, St. Paul, MN, USA), applied as a 2-step

97 Er (ScEr) and 1-step Se (ScSe); and All Bond Universal (Bisco Inc., Schaumburg, IL,
98 USA) applied as a 2-step Er (AlEr) and 1-step Se (AlSe).

99 Restorative procedure and specimen preparation

100 The adhesive systems were applied in accordance with the respective
101 manufacturer's instructions; however, with vigorous rubbing action,²⁶ described in
102 Table 1. After the bonding procedures, all teeth received a microhybrid composite
103 restoration (Opallis, FGM Produtos Odontológicos, Joinville, SC, Brazil) in two
104 increments of 2 mm. Each increment was light polymerized for 40 s using an LED
105 light curing unit set at 1200 mW/cm² (Radii-cal, SDI Limited, Bayswater, Victoria,
106 Australia).

107 After the restored teeth had been stored in distilled water at 37 °C for 24 h, the
108 specimens were sectioned longitudinally in the mesio-distal and buccal-lingual
109 directions across the bonded interface using a slow-speed diamond saw (Isomet,
110 Buehler Ltd., Lake Bluff, IL, USA) to obtain 25–30 resin–denture sticks with a cross
111 sectional area of approximately 0.8 mm² measured with a digital caliper (Digimatic
112 Caliper, Mitutoyo, Tokyo, Japan).

113 The halves of specimens from each tooth were used for the immediate
114 microtensile bond strength evaluation (μ TBS) and another one after 6 months of
115 water storage (37 °C). The same amount of distilled water was used throughout the
116 study period.

117 Microtensile bond strength test (μ TBS)

118 Resin–denture bonded sticks were attached to a Geraldeli's jig²⁷ with
119 cyanoacrylate adhesive and tested under tension (Kratos Dinamometros, Cotia, SP,

120 Brazil) at 0.5 mm/min until failure. The μ TBS values were calculated by dividing the
121 load at failure by the cross-sectional bonding area.

122 The failure mode of the specimens was classified as cohesive ([C] failure
123 exclusive within dentine or resin composite), adhesive ([A] failure at resin/dentine
124 interface), or mixed ([M] failure at resin/dentine interface, which included cohesive
125 failure of the neighboring substrates). The classification was performed under a
126 stereomicroscope at 100 \times magnification (Olympus SZ40, Tokyo, Japan). Specimens
127 with premature failures (PF) were included in the tooth mean.

128 Nanoleakage (NL) evaluation

129 Three resin-bonded sticks from each tooth in each evaluation time were used
130 for NL. Ammoniacal silver nitrate was prepared according to the protocol previously
131 described by Tay et al.²⁸ The sticks were placed in the ammoniacal silver nitrate
132 solution in darkness for 24 h, rinsed thoroughly in distilled water, and immersed in
133 photo developing solution for 8 h under a fluorescent light to reduce silver ions into
134 metallic silver grains within voids along the bonded interface. Specimens were
135 polished with a wet #600, 1000, 1200, 1500, 2000 and 2500-grit SiC paper and 1 and
136 0.25 μ m diamond paste (Buehler Ltd., Lake Bluff, IL, USA) using a polishing cloth.
137 They were ultrasonically cleaned, air dried, mounted on stubs, and coated with
138 carbon-gold (Shimadzu IC 50, Tóquio, Japão). Resin–dentine interfaces were
139 analyzed in a field-emission scanning electron microscope operated in the
140 backscattered mode (SSX-550, Shimadzu, Tóquio, Japão).

141 Three images were captured of each resin–dentine bonded stick. The relative
142 percentage of NL within the adhesive and hybrid layers in each specimen was
143 measured in all images using the UTHSCSA ImageTool 3.0 software (Department of

144 Dental Diagnostic Science at The University of Texas Health Science Centre, San
145 Antonio, Texas) by a blinded researcher. Values originating from the same specimen
146 were averaged for statistical purposes. The mean NL of all sticks from the same tooth
147 was averaged for statistical purposes.

148 Statistical analysis

149 The experimental unit in the current study was the hemi-tooth, since half of the
150 sample was tested immediately and the other half tested only after 6 months. The
151 μ TBS and NL of all sticks from the same hemi-tooth were averaged for statistical
152 purposes. The μ TBS (MPa) and NL (%) means for every testing group was
153 expressed as the average of five hemi-teeth used per group. The premature failures
154 from MTBS were not included in the hemi-tooth mean. The data of μ TBS (MPa) and
155 NL (%) were subjected to a two-way repeated measures ANOVA (Adhesive vs.
156 Storage time) and a post hoc test (Tukey's post hoc test at $\alpha = 0.05$) for pair-wise
157 comparisons.

158 Results

159 The percentage of specimens with premature failure and the frequency of each
160 fracture pattern mode are shown in Table 2. For all experimental conditions, a lower
161 percentage of premature (5.7 % on average) and cohesive failures (4.6% on average)
162 of the specimens were observed. The majority of the specimens showed adhesive or
163 adhesive/mixed failures.

164 The overall μ TBS values of all adhesives are shown in Table 3. The cross-product
165 Adhesive vs. Storage time was statistically significant ($p = 0.001$). Despite PkSe and
166 PkEr showing higher immediate μ TBS similar to controls (CSE and SB; $p > 0.05$), a

167 significant decrease of μ TBS values occurred after 6 months of water storage ($p <$
168 0.05). Adhesives ScSe, ScEr, AISe and AIEr showed inferior immediate μ TBS in
169 comparison with controls (CSE and SB; $p < 0.05$), but only when applied in the Se
170 mode (ScSe and AISe) there is not significant decrease of μ TBS values after 6
171 months of water storage ($p > 0.05$). In the Er mode, only AI showed significant and
172 lower μ TBS values after 6 months of water storage ($p < 0.05$).

173 The overall NL values of all adhesives are shown in Table 4. The cross-
174 product Adhesive vc. Storage time was statistically significant ($p = 0.001$). Only PkSe
175 and PkEr in both modes of application showed higher and significant amounts of NL
176 in immediate time ($p < 0.05$), and this NL pattern significantly increased after 6
177 months of water storage ($p < 0.05$). Sb and AI when applied in Se and Er mode
178 showed a lower NL pattern in both periods of time and similar to controls (CSE and
179 SB; $p > 0.05$).

180 **Discussion**

181 The results of the present study demonstrated that the universal adhesive
182 tested had a heterogeneous behavior, since some diminished the bond over the
183 course of time and some did not.

184 The adhesive Peak, in spite of presenting excellent bond strength results,
185 formed an adhesive and hybrid layer with a large quantity of NL in the immediate time
186 interval. NL represents the location of defects at the resin-dentin interface that may
187 serve as pathways for degradation of the resin/dentin bond over time.²⁹ Silver nitrate
188 is capable of occupying nanometric-sized spaces present around the exposed
189 collagen fibrils where the monomers were unable to infiltrate or where residual water

190 was not displaced by the adhesive, or even in areas with incomplete monomer
191 conversion²⁹, factors preponderant for the degradation of the bond interface.

192 This large quantity of silver nitrate, in the case of PkSe, was owing to the fact
193 that the primer of the system has a very low pH (Peak SE Primer - pH = 1.2) when
194 compared with those of the other adhesives (Scotchbond Universal=3; Allbond
195 Universal=2.4)¹⁶ characterizing this material within the category of the more
196 aggressive Se.^{30, 31} It has been related that acid and unpolymerized monomers
197 present in acidic adhesive do not completely infiltrate into the region of dentin that
198 was demineralize,^{5, 6} probably the result of hydrolysis of the ester bond of the acidic
199 monomer that results in a strong phosphoric acid⁵ which may also explain the intense
200 deposition of silver nitrate within the hybrid layer of this adhesive.

201 Furthermore, in the case of PkSe, only in this material does the manufacturer
202 indicate the application of Peak LC Bond, compared to the other universal adhesives.
203 Various in vitro^{13, 32-34} and clinical studies^{35, 36} have shown that the application of an
204 additional layer increases the performance of 1-step Se adhesives, provided that this
205 is a layer with a hydrophobic nature. An additional layer of a traditional Bond
206 incorporates non-solvated hydrophobic monomers at the bond interface, which
207 diminishes the relative concentration of solvents retained and non-reacted monomers
208 in the adhesive layer³⁷, making it less permeable^{38, 39} and have less propensity to the
209 effects of degradation over the course of time.^{40, 41}

210 However, Peak LC Bond appears to be as hydrophilic as Peak SE Primer,
211 since no hydrophobic monomers were listed in the composition of Peak LC Bond
212 (Table 1). This high level of hydrophilicity must be responsible for the increase in the
213 patterns of degradation observed for this adhesive in the present study. Indeed, it is

214 important for Peak LC Bond to be hydrophilic to be applied in the Er technique.
215 Nevertheless, in the case of PkEr, in which the primer is not applied, Peak LC Bond
216 (pH=2.0) may have caused an additional etching and an increase in the pattern of
217 conditioning of the dentinal substrate. This probably resulted in an increase in
218 demineralization and collagen exposure,^{31, 42} thereby increasing the NL⁴³ for PkEr, as
219 may be seen in Figure 1.

220 On the other hand, in spite of Sc and Al being one-step Se adhesives and
221 therefore highly hydrophilic, for 3 of the 4 groups tested, no degradation occurred at
222 the bond interface after 6 months of storage in water. This must be attributed, as
223 described in the introduction, to the presence of monomers capable of producing a
224 chemical bond to the hard structures of teeth,^{21, 22} as opposed to the lack of this
225 compound in Peak.

226 In this case, Sc and Al contain 10-MDP in their composition, as does CSE,
227 which was the first SE adhesive to contain this component. Studies with CSE have
228 demonstrated that 10-MDP allows for a stable chemical bond to dentin over the
229 course of time, both in vitro^{32, 44-46} and in clinical^{14, 47, 48} studies of long-term. This
230 compound forms a stable nanolayer together with a deposition of stable MDP-Ca
231 salts at the adhesive interface,⁴⁹ which increases its mechanical strength^{49, 50}.
232 However, in spite of the presence of MDP, the adhesives had different behaviors.

233 Please note that in spite of its stability, the μ TBS values of ScSe and AlSe in
234 the immediate time interval were not equivalent to those of the control CSE. ScSe
235 and AlSe are one-step adhesives and this probably leads to the concentration of MDP
236 being lower than it is in CSE, which has MDP incorporated into both the *primer* and
237 the bond.⁴⁹ Moreover, it has been demonstrated that the presence of HEMA, a

238 component of Sc and Al may compete with MDP by the bond to the calcium of
239 hydroxyapatite, thereby harming the chemical bond of MDP to dentin.⁴⁹ Nevertheless,
240 both ScSe and AlSe demonstrated excellent results with regard to bond stability.

241 The difference between ScSe and AlSe may be explained by two factors:
242 presence of the copolymer of polyalkenoic acid in Sc; and the high quantity of solvent
243 of Al. Sc contains a specific polyalkenoic acid copolymer (PAC) used in resin-
244 modified glass ionomer Vitrebond (3M ESPE). PAC bonds chemically and
245 spontaneously to hydroxyapatite in glass ionomer materials,⁵¹ and a recent study
246 demonstrated that the presence of PAC showed more enhanced bond strength than a
247 PAC-free adhesive with the same composition.^{47, 51} Yoshida⁴⁹ hypothesized that PAC
248 may compete with the MDP present in Sc; however, if we compare the longevity
249 results of Sc (MDP + PAC) in comparison with SB (PAC), two materials with similar
250 compositions, the only difference being the presence of MDP in the former, we may
251 observe that the association MDP-PAC enhanced the results observed, since both
252 ScSe and ScEr demonstrated stable results after 6 months of storage in water.

253 Whereas Al contains more solvent when compared with Sc (30-60 wt% and
254 10-15 wt%0 respectively.^{52, 53} This will lead to a larger quantity of residual solvent
255 retained in the hybrid layer and adhesive layer,⁵⁴ preventing the formation of a
256 polymer with high reticulation,⁵⁵⁻⁵⁷ thus diminishing the degree of conversion,³⁸
257 reducing the resin-dentin bond strength⁵⁸⁻⁶¹ and increasing the permeability of the
258 adhesive layer after polymerization.^{62, 63} Consequently, the polymers will be more
259 susceptible to degradation over time.^{3, 64}

260 This may explain the lower results of AlSe in the immediate time interval in
261 comparison with the other adhesives, and this is in agreement with a recent study

262 published by Munoz et al.,¹⁶ even when applied actively; whereas in the mentioned
263 study, AI was applied passively.¹⁶ Active/vigorous application improves the immediate
264 and long-term results of the bond to dentin of 1-step SE adhesive systems⁶⁵⁻⁶⁹
265 because it increases the penetration of monomers into dentin and solvent
266 evaporation. Agitation will also improve the efficacy of polymerization by improving
267 the chemical interaction of the adhesive with the dental substrate, particularly for the
268 acid SE adhesives.^{43, 70} In addition, unreacted acid monomers present in the
269 superficial layer of the adhesive may be taken to a basal area of dentin, increasing
270 demineralization of the substrate, diffusion of monomers, and improving the
271 interaction with the smear layer and subjacent dentin.⁶⁵⁻⁶⁷

272 As only AI/Er demonstrated degradation of the bond strength values over the
273 course of time after 6 months of evaluation, we could hypothesize that the presence
274 of PAC is more important for Er adhesives than for Se. Some authors have indicated
275 that the function of PAC is to improve the stability to humidity,^{71, 72} a crucial factor for
276 Er adhesives, which due to dentin demineralization has a more sensitive technique
277 when compared with that of the Se adhesives.²

278 Although there are various universal adhesive systems available on the
279 market, among the materials evaluated, only those that contain MDP presented better
280 bond longevity characterized by the maintenance of bond strength and nanoleakage
281 pattern, as observed for the 2-step Se adhesive (CSE) tested.

282

283 **Reference**

284

285 1. Van Meerbeek B, Van Landuyt K, De Munck J, Hashimoto M, Peumans M,
286 Lambrechts P, Yoshida Y, Inoue S & Suzuki K (2005) Technique-sensitivity of
287 contemporary adhesives *Dental Materials Journal* **24(1)** 1-13.

288 2. Perdigao J (2007) New developments in dental adhesion *Dental clinics of North*
289 *America* **51(2)** 333-357.

290 3. De Munck J, Van Landuyt K, Peumans M, Poitevin A, Lambrechts P, Braem M &
291 Van Meerbeek B (2005) A critical review of the durability of adhesion to tooth tissue:
292 methods and results *Journal of Dental Research* **84(2)** 118-132.

293 4. Reis A, Loguercio AD, Azevedo CL, de Carvalho RM, da Julio Singer M & Grande
294 RH (2003) Moisture spectrum of demineralized dentin for adhesive systems with
295 different solvent bases *The Journal of Adhesive Dentistry* **5(3)** 183-192.

296 5. Wang Y & Spencer P (2005) Continuing etching of an all-in-one adhesive in wet
297 dentin tubules *Journal of Dental Research* **84(4)** 350-354.

298 6. Carvalho RM, Chersoni S, Frankenberger R, Pashley DH, Prati C & Tay FR (2005)
299 A challenge to the conventional wisdom that simultaneous etching and resin
300 infiltration always occurs in self-etch adhesives *Biomaterials* **26(9)** 1035-1042,
301 10.1016/j.biomaterials.2004.04.003.

302 7. Unemori M, Matsuya Y, Akashi A, Goto Y & Akamine A (2004) Self-etching
303 adhesives and postoperative sensitivity *American Journal of Dentistry* **17(3)** 191-195.

304 8. Pashley DH & Tay FR (2001) Aggressiveness of contemporary self-etching
305 adhesives. Part II: etching effects on unground enamel *Dental Materials* **17(5)** 430-
306 444.

307 9. Kanemura N, Sano H & Tagami J (1999) Tensile bond strength to and SEM
308 evaluation of ground and intact enamel surfaces *Journal of Dentistry* **27(7)** 523-530.

309 10. Hafer M, Schneider H, Rupf S, Busch I, Fuchss A, Merte I, Jentsch H, Haak R &
310 Merte K (2013) Experimental and clinical evaluation of a self-etching and an etch-
311 and-rinse adhesive system *The Journal of Adhesive Dentistry* **15(3)** 275-286,
312 10.3290/j.jad.a29633.

- 313 11. Van Meerbeek B, De Munck J, Mattar D, Van Landuyt K & Lambrechts P (2003)
314 Microtensile bond strengths of an etch&rinse and self-etch adhesive to enamel and
315 dentin as a function of surface treatment *Operative dentistry* **28(5)** 647-660.
- 316 12. Rotta M, Bresciani P, Moura SK, Grande RH, Hilgert LA, Baratieri LN, Loguercio
317 AD & Reis A (2007) Effects of phosphoric acid pretreatment and substitution of
318 bonding resin on bonding effectiveness of self-etching systems to enamel *The*
319 *Journal of Adhesive Dentistry* **9(6)** 537-545.
- 320 13. Frankenberger R, Perdigao J, Rosa BT & Lopes M (2001) "No-bottle" vs "multi-
321 bottle" dentin adhesives--a microtensile bond strength and morphological study
322 *Dental Materials* **17(5)** 373-380.
- 323 14. Peumans M, De Munck J, Van Landuyt KL, Poitevin A, Lambrechts P & Van
324 Meerbeek B (2010) Eight-year clinical evaluation of a 2-step self-etch adhesive with
325 and without selective enamel etching *Dental Materials* **26(12)** 1176-1184.
- 326 15. Taschner M, Nato F, Mazzoni A, Frankenberger R, Falconi M, Petschelt A &
327 Breschi L (2012) Influence of preliminary etching on the stability of bonds created by
328 one-step self-etch bonding systems *European Journal of Oral Sciences* **120(3)** 239-
329 248.
- 330 16. Munoz MA, Luque I, Hass V, Reis A, Loguercio AD & Bombarda NH (2013)
331 Immediate bonding properties of universal adhesives to dentine *Journal of Dentistry*
332 **41(5)** 404-411.
- 333 17. Proenca JP, Polido M, Osorio E, Erhardt MC, Aguilera FS, Garcia-Godoy F,
334 Osorio R & Toledano M (2007) Dentin regional bond strength of self-etch and total-
335 etch adhesive systems *Dental Materials* **23(12)** 1542-1548.
- 336 18. Erhardt MC, Osorio E, Aguilera FS, Proenca JP, Osorio R & Toledano M (2008)
337 Influence of dentin acid-etching and NaOCl-treatment on bond strengths of self-etch
338 adhesives *American Journal of Dentistry* **21(1)** 44-48.
- 339 19. Tay FR, Suh BI, Pashley DH, Prati C, Chuang SF & Li F (2003) Factors
340 contributing to the incompatibility between simplified-step adhesives and self-cured or
341 dual-cured composites. Part II. Single-bottle, total-etch adhesive *The Journal of*
342 *Adhesive Dentistry* **5(2)** 91-105.
- 343 20. Tay FR, Pashley DH, Yiu CK, Sanares AM & Wei SH (2003) Factors contributing
344 to the incompatibility between simplified-step adhesives and chemically-cured or dual-

- 345 cured composites. Part I. Single-step self-etching adhesive *The Journal of Adhesive*
346 *Dentistry* **5(1)** 27-40.
- 347 21. Perdigao J, Sezinando A & Monteiro PC (2012) Laboratory bonding ability of a
348 multi-purpose dentin adhesive *American Journal of Dentistry* **25(3)** 153-158.
- 349 22. Hanabusa M, Mine A, Kuboki T, Momoi Y, Van Ende A, Van Meerbeek B & De
350 Munck J (2012) Bonding effectiveness of a new 'multi-mode' adhesive to enamel and
351 dentine *Journal of Dentistry* **40(6)** 475-484.
- 352 23. Perdigao J, Munoz M, Sezinando A, Luque-Martinez I, Staichak R, Reis A &
353 Loguercio A (2013) Immediate adhesive properties to dentin and enamel of a
354 universal adhesive associated with a hydrophobic resin coat *Operative dentistry*, in
355 press.
- 356 24. Heintze SD, Ruffieux C & Rousson V (2010) Clinical performance of cervical
357 restorations--a meta-analysis *Dental Materials* **26(10)** 993-1000.
- 358 25. Marchesi G, Frassetto A, Mazzoni A, Apolonio F, Diolosa M, Cadenaro M,
359 Pashley DH, Tay F & Breschi L (2013) Adhesive performance of a multi-mode
360 adhesive system: 1-Year in vitro study *Journal of Dentistry*, in press.
- 361 26. Dal-Bianco K, Pellizzaro A, Patzlaft R, de Oliveira Bauer JR, Loguercio AD & Reis
362 A (2006) Effects of moisture degree and rubbing action on the immediate resin-dentin
363 bond strength *Dental Materials* **22(12)** 1150-1156.
- 364 27. Perdigao J, Geraldeli S, Carmo AR & Dutra HR (2002) In vivo influence of
365 residual moisture on microtensile bond strengths of one-bottle adhesives *Journal of*
366 *Esthetic and Restorative Dentistry* **14(1)** 31-38.
- 367 28. Tay FR, Pashley DH, Suh BI, Carvalho RM & Itthagarun A (2002) Single-step
368 adhesives are permeable membranes *Journal of Dentistry* **30(7-8)** 371-382.
- 369 29. Sano H (2006) Microtensile testing, nanoleakage, and biodegradation of resin-
370 dentin bonds *Journal of Dental Research* **85(1)** 11-14.
- 371 30. Tay FR & Pashley DH (2001) Aggressiveness of contemporary self-etching
372 systems. I: Depth of penetration beyond dentin smear layers *Dental Materials* **17(4)**
373 296-308.

- 374 31. Van Meerbeek B, De Munck J, Yoshida Y, Inoue S, Vargas M, Vijay P, Van
375 Landuyt K, Lambrechts P & Vanherle G (2003) Buonocore memorial lecture.
376 Adhesion to enamel and dentin: current status and future challenges *Operative*
377 *Dentistry* **28(3)** 215-235.
- 378 32. Feitosa VP, Leme AA, Sauro S, Correr-Sobrinho L, Watson TF, Sinhoreti MA &
379 Correr AB (2012) Hydrolytic degradation of the resin-dentine interface induced by the
380 simulated pulpal pressure, direct and indirect water ageing *Journal of Dentistry*, in
381 press.
- 382 33. Albuquerque M, Pegoraro M, Mattei G, Reis A & Loguercio AD (2008) Effect of
383 double-application or the application of a hydrophobic layer for improved efficacy of
384 one-step self-etch systems in enamel and dentin *Operative Dentistry* **33(5)** 564-570,
385 10.2341/07-145.
- 386 34. Reis A, Albuquerque M, Pegoraro M, Mattei G, Bauer JR, Grande RH, Klein-
387 Junior CA, Baumhardt-Neto R & Loguercio AD (2008) Can the durability of one-step
388 self-etch adhesives be improved by double application or by an extra layer of
389 hydrophobic resin? *Journal of Dentistry* **36(5)** 309-315.
- 390 35. Reis A, Leite TM, Matte K, Michels R, Amaral RC, Geraldeli S & Loguercio AD
391 (2009) Improving clinical retention of one-step self-etching adhesive systems with an
392 additional hydrophobic adhesive layer *Journal of the American Dental Association*
393 **140(7)** 877-885.
- 394 36. Loguercio AD & Reis A (2008) Application of a dental adhesive using the self-etch
395 and etch-and-rinse approaches: an 18-month clinical evaluation *Journal of the*
396 *American Dental Association* **139(1)** 53-61.
- 397 37. Breschi L, Mazzoni A, Ruggeri A, Cadenaro M, Di Lenarda R & De Stefano
398 Dorigo E (2008) Dental adhesion review: aging and stability of the bonded interface
399 *Dental Materials* **24(1)** 90-101.
- 400 38. Cadenaro M, Antonioli F, Sauro S, Tay FR, Di Lenarda R, Prati C, Biasotto M,
401 Contardo L & Breschi L (2005) Degree of conversion and permeability of dental
402 adhesives *European Journal of Oral Sciences* **113(6)** 525-530.
- 403 39. Breschi L, Cadenaro M, Antonioli F, Sauro S, Biasotto M, Prati C, Tay FR & Di
404 Lenarda R (2007) Polymerization kinetics of dental adhesives cured with LED:
405 correlation between extent of conversion and permeability *Dental Materials* **23(9)**
406 1066-1072.

- 407 40. Reis A, Grande RH, Oliveira GM, Lopes GC & Loguercio AD (2007) A 2-year
408 evaluation of moisture on microtensile bond strength and nanoleakage *Dental*
409 *Materials* **23(7)** 862-870.
- 410 41. Reis A, de Carvalho Cardoso P, Vieira LC, Baratieri LN, Grande RH & Loguercio
411 AD (2008) Effect of prolonged application times on the durability of resin-dentin bonds
412 *Dental Materials* **24(5)** 639-644.
- 413 42. Liu Y, Tjaderhane L, Breschi L, Mazzoni A, Li N, Mao J, Pashley DH & Tay FR
414 (2011) Limitations in bonding to dentin and experimental strategies to prevent bond
415 degradation *Journal of Dental Research* **90(8)** 953-968.
- 416 43. Wang Y & Spencer P (2003) Hybridization efficiency of the adhesive/dentin
417 interface with wet bonding *Journal of Dental Research* **82(2)** 141-145.
- 418 44. Toledano M, Osorio R, Osorio E, Aguilera FS, Yamauti M, Pashley DH & Tay F
419 (2007) Durability of resin-dentin bonds: effects of direct/indirect exposure and storage
420 media *Dental Materials* **23(7)** 885-892.
- 421 45. Waidyasekera K, Nikaido T, Weerasinghe DS, Ichinose S & Tagami J (2009)
422 Reinforcement of dentin in self-etch adhesive technology: a new concept *Journal of*
423 *Dentistry* **37(8)** 604-609.
- 424 46. Inoue S, Koshiro K, Yoshida Y, De Munck J, Nagakane K, Suzuki K, Sano H &
425 Van Meerbeek B (2005) Hydrolytic stability of self-etch adhesives bonded to dentin
426 *Journal of Dental Research* **84(12)** 1160-1164.
- 427 47. Perdigao J, Kose C, Mena-Serrano A, De Paula E, Tay L, Reis A & Loguercio A
428 (2013) A New Universal Simplified Adhesive: 18-Month Clinical Evaluation *Operative*
429 *Dentistry*, in press.
- 430 48. Mena-Serrano A, Kose C, De Paula EA, Tay LY, Reis A, Loguercio AD &
431 Perdigao J (2013) A new universal simplified adhesive: 6-month clinical evaluation
432 *Journal of Esthetic and Restorative Dentistry* **25(1)** 55-69.
- 433 49. Yoshida Y, Yoshihara K, Nagaoka N, Hayakawa S, Torii Y, Ogawa T, Osaka A &
434 Meerbeek BV (2012) Self-assembled Nano-layering at the Adhesive Interface *Journal*
435 *of Dental Research* **91(4)** 376-381.
- 436 50. Yoshihara K, Yoshida Y, Nagaoka N, Fukegawa D, Hayakawa S, Mine A,
437 Nakamura M, Minagi S, Osaka A, Suzuki K & Van Meerbeek B (2010) Nano-

- 438 controlled molecular interaction at adhesive interfaces for hard tissue reconstruction
439 *Acta biomaterialia* **6(9)** 3573-3582.
- 440 51. Mitra SB, Lee CY, Bui HT, Tantbirojn D & Rusin RP (2009) Long-term adhesion
441 and mechanism of bonding of a paste-liquid resin-modified glass-ionomer *Dental*
442 *Materials* **25(4)** 459-466.
- 443 52. Chen L & Suh BI (2013) Effect of hydrophilicity on the compatibility between a
444 dual-curing resin cement and one-bottle simplified adhesives *The Journal of Adhesive*
445 *Dentistry* **15(4)** 325-331.
- 446 53. 3MESPE (2013)
447 [http://multimedia.3m.com/mws/mediawebserver?mwsId=SSSSSuUn_zu8l00xMY_eMYt1Nv70k](http://multimedia.3m.com/mws/mediawebserver?mwsId=SSSSSuUn_zu8l00xMY_eMYt1Nv70k17zHvu9lxtD7SSSSSS--)
448 [17zHvu9lxtD7SSSSSS--](http://multimedia.3m.com/mws/mediawebserver?mwsId=SSSSSuUn_zu8l00xMY_eMYt1Nv70k17zHvu9lxtD7SSSSSS--); Retrieved online december 6, 2013.
- 449 54. Yiu CK, Pashley EL, Hiraishi N, King NM, Goracci C, Ferrari M, Carvalho RM,
450 Pashley DH & Tay FR (2005) Solvent and water retention in dental adhesive blends
451 after evaporation *Biomaterials* **26(34)** 6863-6872.
- 452 55. Ye Q, Spencer P, Wang Y & Misra A (2007) Relationship of solvent to the
453 photopolymerization process, properties, and structure in model dentin adhesives
454 *Journal of Biomedical Materials Research Part A* **80(2)** 342-350.
- 455 56. Loguercio AD, Loeblein F, Cherobin T, Ogliairi F, Piva E & Reis A (2009) Effect of
456 solvent removal on adhesive properties of simplified etch-and-rinse systems and on
457 bond strengths to dry and wet dentin *The Journal of Adhesive Dentistry* **11(3)** 213-
458 219.
- 459 57. Paul SJ, Leach M, Rueggeberg FA & Pashley DH (1999) Effect of water content
460 on the physical properties of model dentine primer and bonding resins *Journal of*
461 *Dentistry* **27(3)** 209-214.
- 462 58. Hass V, Folkuenig MS, Reis A & Loguercio AD (2011) Influence of adhesive
463 properties on resin-dentin bond strength of one-step self-etching adhesives *Journal of*
464 *Adhesive Dentistry* **13(5)** 417-424.
- 465 59. Reis A, Ferreira SQ, Costa TR, Klein-Junior CA, Meier MM & Loguercio AD
466 (2010) Effects of increased exposure times of simplified etch-and-rinse adhesives on
467 the degradation of resin-dentin bonds and quality of the polymer network *European*
468 *Journal of Oral Sciences* **118(5)** 502-509.

- 469 60. Takahashi H, Sato H, Uno S, Pereira PN & Sano H (2002) Effects of mechanical
470 properties of adhesive resins on bond strength to dentin *Dental Materials* **18(3)** 263-
471 268.
- 472 61. Cho BH & Dickens SH (2004) Effects of the acetone content of single solution
473 dentin bonding agents on the adhesive layer thickness and the microtensile bond
474 strength *Dental Materials* **20(2)** 107-115.
- 475 62. Malacarne J, Carvalho RM, de Goes MF, Svizero N, Pashley DH, Tay FR, Yiu CK
476 & Carrilho MR (2006) Water sorption/solubility of dental adhesive resins *Dental*
477 *Materials* **22(10)** 973-980.
- 478 63. Ito S, Hashimoto M, Wadgaonkar B, Svizero N, Carvalho RM, Yiu C, Rueggeberg
479 FA, Foulger S, Saito T, Nishitani Y, Yoshiyama M, Tay FR & Pashley DH (2005)
480 Effects of resin hydrophilicity on water sorption and changes in modulus of elasticity
481 *Biomaterials* **26(33)** 6449-6459, 10.1016/j.biomaterials.2005.04.052.
- 482 64. Breschi L, Mazzoni A, Ruggeri A, Cadenaro M, Di Lenarda R & De Stefano
483 Dorigo E (2008) Dental adhesion review: aging and stability of the bonded interface
484 *Dental Materials* **24(1)** 90-101.
- 485 65. do Amaral RC, Stanislawczuk R, Zander-Grande C, Michel MD, Reis A &
486 Loguercio AD (2009) Active application improves the bonding performance of self-
487 etch adhesives to dentin *Journal of Dentistry* **37(1)** 82-90.
- 488 66. do Amaral RC, Stanislawczuk R, Zander-Grande C, Gagler D, Reis A & Loguercio
489 AD (2010) Bond strength and quality of the hybrid layer of one-step self-etch
490 adhesives applied with agitation on dentin *Operative Dentistry* **35(2)** 211-219.
- 491 67. Miyazaki M, Platt JA, Onose H & Moore BK (1996) Influence of dentin primer
492 application methods on dentin bond strength *Operative Dentistry* **21(4)** 167-172.
- 493 68. Pleffken PR, de Almeida Lourenco AP, Torres CR & Buhler Borges A (2011)
494 Influence of application methods of self-etching adhesive systems on adhesive bond
495 strength to dentin *The Journal of Adhesive Dentistry* **13(6)** 517-525.
- 496 69. Loguercio AD, Stanislawczuk R, Mena-Serrano A & Reis A (2011) Effect of 3-year
497 water storage on the performance of one-step self-etch adhesives applied actively on
498 dentine *Journal of Dentistry* **39(8)** 578-587.
- 499 70. Zhang Y & Wang Y (2013) Effect of application mode on interfacial morphology
500 and chemistry between dentine and self-etch adhesives *Journal of Dentistry* **41(3)**
501 231-240.

502 71. Van Meerbeek B, Conn LJ, Jr., Duke ES, Eick JD, Robinson SJ & Guerrero D
503 (1996) Correlative transmission electron microscopy examination of
504 nondemineralized and demineralized resin-dentin interfaces formed by two dentin
505 adhesive systems *Journal of Dental Research* **75(3)** 879-888.

506 72. Spencer P, Wang Y, Walker MP, Wieliczka DM & Swafford JR (2000) Interfacial
507 chemistry of the dentin/adhesive bond *Journal of Dental Research* **79(7)** 1458-1463.

508

509

510

511 **Legend of Figure**

512 **Figure 1.** Representative backscatter SEM images of the resin-dentin adhesive
513 interfaces of each experimental group for immediate and 6-month times. (Control
514 etch-and-rinse group= Adper Single Bond 2 and control self-etch group=Clearfil SE
515 Bond). For the Peak Universal Adhesive System, the amount of nanoleakage was
516 higher than the other materials and increased after 6 months for strategies ER (B, F)
517 and SE (J, N). The amount of nanoleakage was lower and stable after 6M within the
518 hybrid layer for Scotchbond Universal (C, G, K, O); AllBond Universal (D, H, L, P) and
519 controls (white arrows) (A, E, I, M). Abbreviations: Co, composite; De, dentine; HL,
520 hybrid layer; AL, adhesive layer; SE, self-etch; ER, etch-and-rinse

521

522

523

Table 1. Adhesive system (batch number), composition and application mode* of the adhesive systems used (*) according to the manufacturer's instructions.

Adhesive (batch number),	Composition	Self-etch strategy	Etch-and-rinse strategy
Adper Single Bond 2 (BPBR)	1. Etchant: 35% phosphoric acid (Scotchbond Etchant) 2. Adhesive: Bis-GMA, HEMA, dimethacrylates, ethanol, water, photoinitiator, methacrylate functional copolymer of polyacrylic and polyitaconic acids, 10% by weight of 5 nm-diameter spherical silica particles	N.A	1. Apply etchant for 15 s 2. Rinse for 10 s 3. Blot excess water 4. Apply 2-3 consecutive coats of adhesive for 15 s with gentle agitation 5. Gently air thin for 5 s 6. Light-cure for 10 s at 1200 mW/cm2
Clearfil SE Bond (Primer: 00954A - Bond: 01416 ^a)	1. Primer: water, MDP, HEMA, camphorquinone, hydrophilic dimethacrylate 2. Bonding: MDP, Bis-GMA, HEMA, camphorquinone, hydrophobic dimethacrylate, N,N-diethanol p-toluidine bond, colloidal silica	1. Apply primer to tooth surface and leave in place for 20 s 2. Dry with air stream to evaporate the volatile ingredients 3. Apply bond to the tooth surface and then create a uniform film using a gentle air stream 4. Light-cure for 10 s at 1200 mW/cm2	N.A
Peak Universal Adhesive System (Peak SE Primer: ON062 – Peak LC Bond: Y062)	1. Etchant: 35% phosphoric acid (Ultraetch) 2. Primer: ethyl alcohol, methacrylic acid, 2-hydroxyethyl methacrylate (Peak SE Primer) 3. Adhesive: Ethyl alcohol, 2-hydroxyethyl methacrylate (Peak LC Bond)	1. Initial use of Peak SE requires activation of the two components separated in the syringe 2. Application of the Peak SE with microbrush for 20 s using continuous scrubbing on dentin. Do not scrub enamel 3. Thin/dry for 3 s using air/water syringe or high volume suction directly over preparation 4. Apply a puddle coat of Peak LC Bond with gently agitate for 10 s 5. Thin/dry 10 s using ¼ to ½ air pressure 6. Light polymerize for 10 s at 1200mW/cm2	1. Apply etchant for 20 s 2. Rinse for 5 s 3. Air dry 2 s 4. Apply a puddle coat of Peak LC Bond with gently agitate for 10 s 5. Thin/dry 10 s using ¼ to ½ air pressure 6. Light-cure for 10 s at 1200mW/cm2
Scotchbond Universal Adhesive (D-82229)	1. Etchant: 34% phosphoric acid, water, synthetic amorphous silica, polyethylene glycol, aluminum oxide. (Scotchbond Universal Etchant) 2. Adhesive: MDP Phosphate monomer, dimethacrylate resins, HEMA, methacrylate-modified polyalkenoic acid copolymer, filler, ethanol, water, initiators, silane	1. Apply the adhesive to the entire preparation with a microbrush and rub it in for 20 s. If necessary, rewet the disposable applicator during treatment 2. Direct a gentle stream of air over the liquid for about 5 s until it no longer moves and the solvent is evaporated completely 3. Light-cure for 10 s	1. Apply etchant for 15 s 2. Rinse for 10 s 3. Air dry 2 s 4. Apply adhesive as for the self-etch mode
All-Bond Universal (1200006111)	1. Etchant Uni-Etch: 32% phosphoric acid, benzalkonium chloride. 2. Adhesive: MDP, Bis-GMA, HEMA, ethanol, water, initiators	1. Apply two separate coats of adhesive, scrubbing the preparation with a microbrush for 10-15 s per coat. Do not light cure between coat. Do not light polymerize between coats. 2. Evaporate excess solvent by thoroughly air-drying with an air syringe for at least 10 s, there should be no visible movement of the material. The surface should have a uniform glossy appearance 3. Light cure for 10 s at 1200 mW/cm2	1. Apply etchant for 15 s 2. Rinse thoroughly 3. Remove excess water with absorbent pellet or high volume suction for 1-2 s 4. Apply adhesive as for the self-etch mode

Table 2. Number of specimens (%) according to fracture mode and the premature failure of all experimental groups.

Adhesive system	Application mode	Time	Fracture pattern			
			A	C	A/M	PF
Adper Single Bond 2	ER control	Immediate	51(73.9)	10(14.5)	6(8.7)	2(2.9)
		6 month	49(73.1)	4(6.0)	9(13.4)	5(7.5)
Clearfil SE Bond	SE control	Immediate	50 (74.6)	3(4.5)	10(14.9)	4(6.0)
		6 month	53(80.3)	2(3.0)	8(12.1)	3(4.6)
Peak Universal	ER	Immediate	56(80)	0(0)	10(14.3)	4(5.7)
		6 month	52(75.4)	2(2.9)	12(17.4)	3(4.3)
	SE	Immediate	58(82.8)	3(4.3)	7(10)	2(2.9)
		6 month	48(69.6)	1(1.5)	13(18.8)	7(10.1)
Scotchbond Universal	ER	Immediate	53(79.1)	2(3.0)	10(14.9)	2(3.0)
		6 month	50(78.1)	0(0)	13(20.3)	1(1.6)
	SE	Immediate	47 (71.2)	3(4.6)	11(16.7)	5(7.5)
		6 month	51(73.9)	1(1.4)	14(20.3)	3(4.4)
Allbond Universal	ER	Immediate	49(71)	8(11.6)	7(10.1)	5(7.3)
		6 month	49(73.1)	2(3.0)	12(17.9)	4(6.0)
	SE	Immediate	51(72.85)	4(5.7)	9(12.9)	6(8.6)
		6 month	47(69.1)	5(7.4)	10(14.7)	6(8.8)

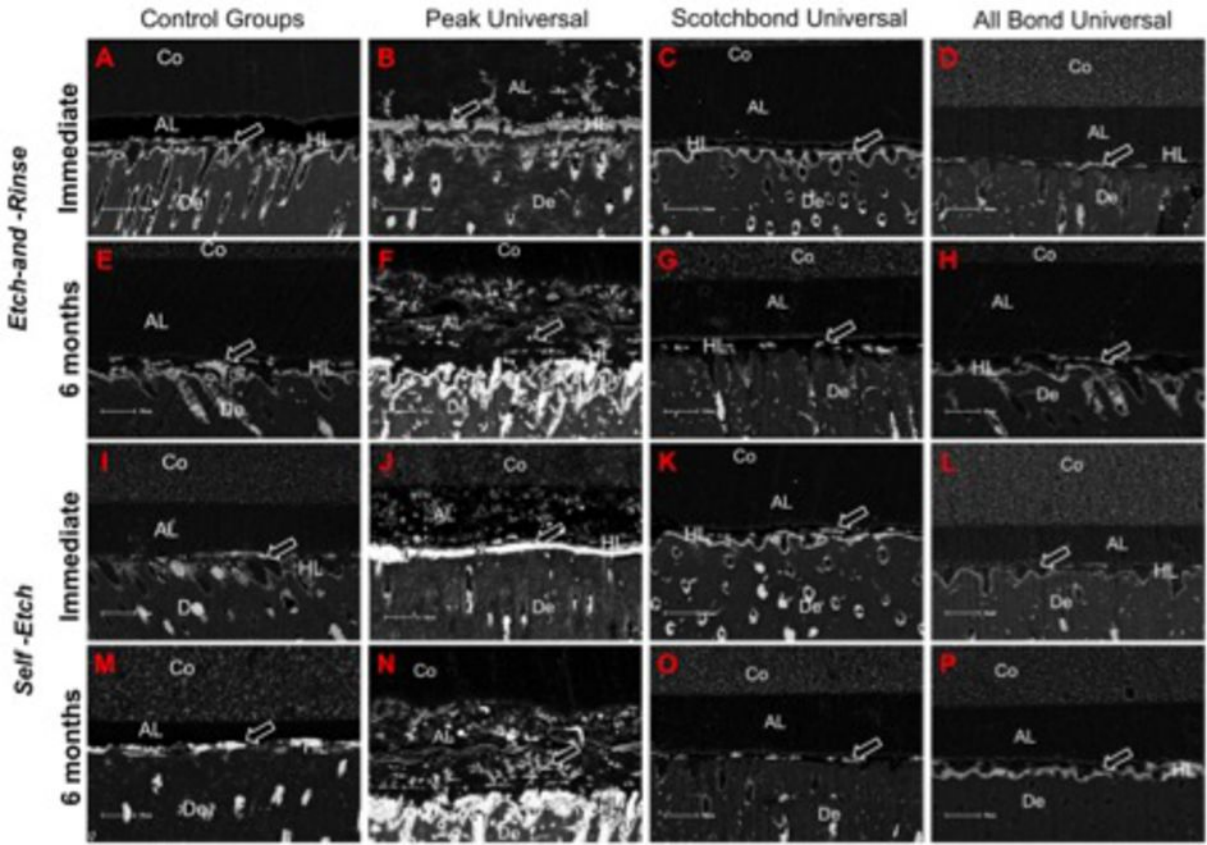
A – adhesive fracture mode; C – cohesive fracture mode; A/M – adhesive/mixed fracture mode; PF – premature failure

Table 3 – Microtensile bond strength (μ TBS) values (means \pm standard deviations) of the different experimental groups

Time	Adhesive System							
	Adper Single Bond 2	Clearfil SE Bond	Peak Universal SE	Peak Universal ER	Scotchbond Universal SE	Scotchbond Universal ER	Allbond Universal SE	Allbond Universal ER
Immediate	47.6 \pm 5.5 a	42.9 \pm 4.4 a, b	39.5 \pm 5.1 b	44.3 \pm 1.6 a, b	33.3 \pm 3.2 c	34.7 \pm 4.6 b, c	20.9 \pm 4.1 e	38.5 \pm 4 b
6 month	38.8 \pm 5.7 b	36.2 \pm 2.7 b, c	27.9 \pm 4.9 d	34.2 \pm 4.2 c	33.6 \pm 5.8 c	34.6 \pm 6.2 c	20.4 \pm 4.8 e	28.1 \pm 4.3 c

Table 4 – Nanoleakage (NL) values (means \pm standard deviations) of the different experimental groups

Time	Adhesive System							
	Adper Single Bond 2	Clearfil SE Bond	Peak Universal Bond SE	Peak Universal Bond ER	ScotchBond Universal SE	ScotchBond Universal ER	Allbond Universal SE	Allbond Universal ER
Immediate	13.1 \pm 2.0 b	7.5 \pm 2.9 a, b	31.6 \pm 3.1 c	23.9 \pm 4.2 c	5.1 \pm 2.1 a	5.3 \pm 1.1 a	6.0 \pm 3.9 a	9.4 \pm 1.8 b
6 month	14.7 \pm 4.1 b	8.6 \pm 4.1 a, b	42.2 \pm 2.6 d	34.4 \pm 4.1 d	4.7 \pm 2.8 a	5.4 \pm 2.0 a	5.9 \pm 1.4 a	8.9 \pm 3.1 a, b

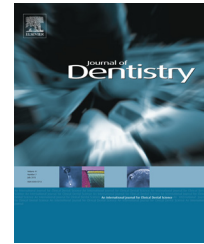


4.3 Capítulo 3 (do experimento 3)

INFLUENCE OF A HYDROPHOBIC RESIN COAT ON THE BONDING EFFICACY OF THREE UNIVERSAL ADHESIVES

Available online at www.sciencedirect.com

ScienceDirect

journal homepage: www.intl.elsevierhealth.com/journals/jden

Influence of a hydrophobic resin coating on the bonding efficacy of three universal adhesives

Miguel Angel Muñoz^{a,b}, Ana Sezinando^c, Issis Luque-Martinez^a,
 Anna Luiza Szesz^a, Alessandra Reis^a, Alessandro D. Loguercio^a,
 Nara Hellen Campanha^a, Jorge Perdigão^{d,*}

^a School of Dentistry, Department of Restorative Dentistry, State University of Ponta Grossa, Ponta Grossa, Paraná, Brazil

^b School of Dentistry, Universidad de Valparaíso, Valparaíso, Chile

^c School of Dentistry, University Rey Juan Carlos, Alcorcón, Madrid, Spain

^d Division of Operative Dentistry, Department of Restorative Sciences, University of Minnesota, Minneapolis, MN, USA

ARTICLE INFO

Article history:

Received 26 October 2013

Accepted 28 January 2014

Available online xxx

Keywords:

Microtensile bond strength

Nanoleakage

Degree of conversion

Etch-and-rinse

Self-etch

Universal simplified adhesive systems

Hydrophobic resin coat

ABSTRACT

Objectives: To evaluate the effect of an additional hydrophobic resin coating (HE) on the resin–dentine microtensile bond strengths (μ TBS), nanoleakage (NL), and *in situ* degree of conversion (DC) of three universal adhesives used in the etch-and-rinse (ER) and the self-etch (SE) modes.

Methods: Sixty caries-free extracted third molars were divided into 12 groups according to the combination of the factors adhesive (All-Bond Universal [ABU]; G-Bond Plus [GBP] and Scotchbond Universal [SBU]), adhesive strategy (ER and SE), and the use of HE (Heliobond; yes or no). After restorations were constructed, specimens were stored in water (37 °C/24 h) and sectioned into resin–dentine beams (0.8 mm²) to be tested under tension (0.5 mm/min). Selected beams from each tooth were used for DC quantification and for NL evaluation. Data from each adhesive were analyzed with two-way ANOVA and Tukey's test ($\alpha = 0.05$).

Results: ABU and GBP resulted in higher μ TBS in the ER mode. The use of HE increased the μ TBS of ABU and GBP only in the SE mode. Lower NL was observed for SBU and ABU in the ER mode + HE, and for GBP in the SE mode + HE. SBU and GBP showed higher DC when used in the ER mode, which was increased with HE application. The DC of ABU was similar in all conditions.

Conclusions: The conversion of 1-step SE to 2-step SE may increase the μ TBS and DC of current universal adhesives. The reduction in the NL is more dependent on the adhesive composition than on the bonding strategy.

© 2014 Published by Elsevier Ltd.

* Corresponding author at: Division of Operative Dentistry, Department of Restorative Sciences, University of Minnesota, Minneapolis, MN 55455, USA.

E-mail addresses: perdi001@umn.edu, alelog@uepg.br (J. Perdigão).

<http://dx.doi.org/10.1016/j.jdent.2014.01.013>

0300-5712/© 2014 Published by Elsevier Ltd.

1. Introduction

For each of the two current adhesive strategies dental manufacturers have introduced simplified adhesive systems, known as 2-step etch-and-rinse [ER] and 1-step self-etch [SE] adhesives,^{1,2} making them more attractive to the clinician and reducing the sensitivity of the application technique. This task of simplification was possible through the inclusion of hydrophilic monomers and the increase in the amount of solvents, to make adhesives compatible with the inherent wet dentine substrate.^{3,4} Recently, newly universal or multi-mode adhesive systems were introduced with manufacturers' claims that one monomer solution can be used for either adhesive strategy^{5,6} without compromising the bonding effectiveness,⁷ therefore being able to replace existing simplified adhesives.

The increased amount of solvents and hydrophilic monomers in the adhesive formulations lead to greater amount of residual solvents entrapped in the adhesive layer.⁴ The accumulation of hydrophilic monomers and especially residual solvents may hinder the formation of a high cross-linking polymer,^{8–10} decreasing the degree conversion (DC)¹¹ which may reduce resin–dentine bond strengths,^{12–15} and increase the permeability of the adhesive layer after polymerization.^{16,17} Consequently, the resulting polymers will be more susceptible to degradation over time.^{2,18}

One of the methods used to circumvent these drawbacks includes the application of an additional layer of a hydrophobic resin coating over the polymerized simplified adhesive.¹⁹ This extra resin coat aims at increasing the thickness and uniformity of the adhesive layer, as well as to reduce the fluid flow across the adhesive interface.^{19,20} Excellent *in vitro* and clinical results have been reported after placement of an hydrophobic resin coating over 1-step SE adhesives^{21,22}; however this technique has not been tested with new universal or multi-mode adhesives. Thus, the aim of this study was to compare the immediate resin–dentine microtensile bond strengths, nanoleakage and *in situ* degree of conversion of three universal adhesives with or without an additional hydrophobic resin coating. We tested the null hypothesis that the application of a hydrophobic resin coat on the cured adhesives will not influence the selected properties of the universal adhesives systems used in both adhesive Q2 strategies (see Fig. 1).

2. Materials and methods

Sixty caries-free extracted human third molars were disinfected in 0.5% chloramine, stored in distilled water and used within six months after extraction. The teeth were collected after obtaining the patients' informed consent under a protocol approved by the local Ethics Committee Review Board under the protocol number 17878/11.

A flat occlusal dentine surface was exposed in all teeth after wet-grinding the occlusal enamel with # 180 grit SiC paper. The exposed dentine surfaces were further polished with wet # 600-grit silicon-carbide paper for 60 s to standardize the smear layer.²³

2.1. Experimental design and specimen preparation

The specimens were randomly assigned to twelve experimental conditions ($n = 5$) resulting from the combination of the factors “adhesive system” (Scotchbond Universal Adhesive [SBU, 3M ESPE, St. Paul, MN, USA, also known as Single Bond Universal in some countries], All-Bond Universal [ABU, Bisco Inc., Schaumburg, IL, USA] and G-Bond Plus [GBP, GC Corporation Tokyo, Japan – also known as G-aenial Bond]) (Table 1); “adhesive strategy” (etch-and-rinse [ER] or self-etch [SE]); and hydrophobic resin coating (yes or no; HE, Heliobond, Ivoclar Vivadent, Schaan, Liechtenstein).

The adhesive systems were applied according to the respective manufacturers' instructions (Table 1), except for G-Bond Plus, for which the manufacturer does not recommend dentine etching with phosphoric acid (Table 1). Composite resin crowns were built with a nanofilled composite (Filtek Z350 XT, 3M ESPE, St. Paul, MN, USA; also named Filtek Supreme XTE or Filtek Supreme Plus in some countries) in two increments of 2 mm each. Each increment was light-cured for 40 s using a LED light-curing unit set at 1200 mW/cm² (Radii-cal, SDI Limited, Bayswater, Victoria, Australia).

After storage in distilled water for 24 h at 37 °C, the specimens were longitudinally sectioned in mesio-distal and buccal-lingual directions across the bonded interface with a slow-speed diamond saw (Isomet Ltd., Buehler Ltd., Lake Bluff, IL, USA) to obtain resin–dentine beams with a cross sectional area of approximately 0.8 mm² measured with a digital calliper (Digimatic Caliper, Mitutoyo, Tokyo, Japan). All specimens from each tooth were used for the μ TBS test, except four that were randomly selected for measurement of nanoleakage (NL) and *in situ* DC.

2.2. Microtensile bond strength (μ TBS)

The resin–dentine bonded beams were attached to a Geraldeli jig²⁴ (Odeme Biotechnology, Joaçaba, SC, Brazil) with cyanoacrylate adhesive and tested under tension (Model 5565, Instron Co., Canton, MA, USA) at 0.5 mm/min until failure. The μ TBS values were calculated by dividing the load at failure by the cross-sectional bonding area.

The failure mode was classified as cohesive [C] failure (exclusively within dentine or resin composite), adhesive [A] failure (at the resin/dentine interface), or mixed [M] failure (at the resin/dentine interface that included cohesive failure of the neighbouring substrates). The failure mode analysis was performed under a stereomicroscope at 100 \times magnification (Olympus SZ40, Tokyo, Japan). Specimens with premature failures (PF) were included in the tooth mean as zero MPa and those with cohesive failures were excluded.

2.3. Nanoleakage (NL)

Two resin-bonded beams from each tooth were used for NL evaluation. Ammoniacal silver nitrate was prepared according to the protocol previously described by Tay et al.²⁵ The beams were placed in the ammoniacal silver nitrate solution in darkness for 24 h, rinsed thoroughly in distilled water, and immersed in photo developing solution for 8 h under a fluorescent light to reduce silver ions into metallic silver

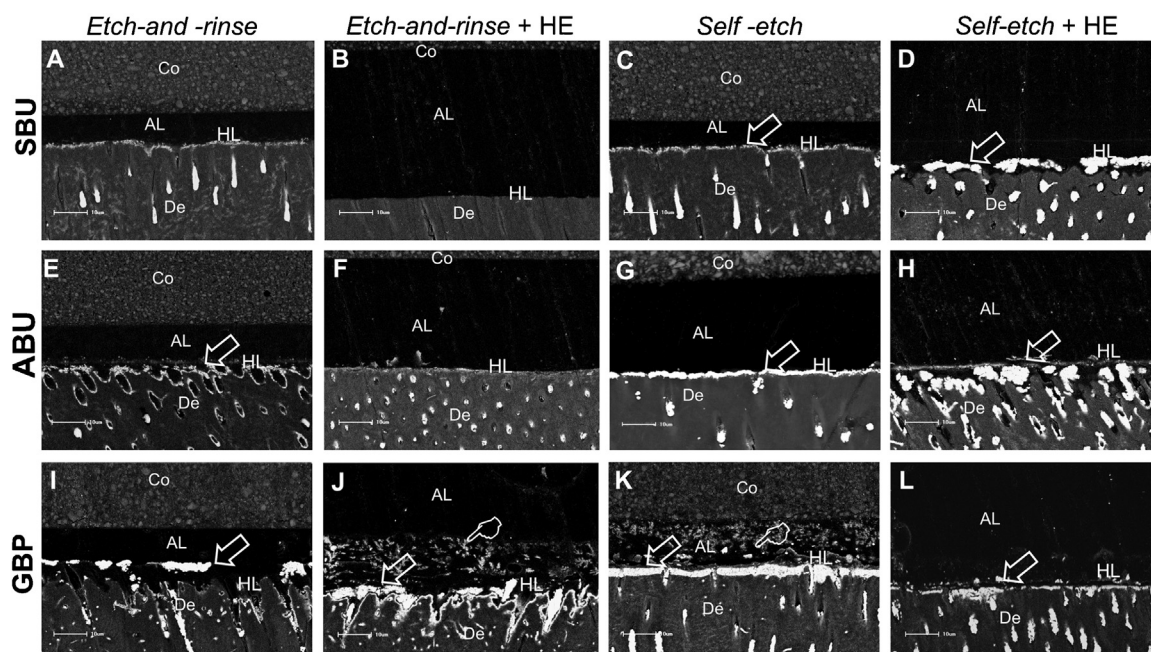


Fig. 1 – Representative backscattered SEM micrographs of the resin–dentine adhesive interfaces of each universal adhesive system (Scotchbond Universal Adhesive [SBU]; All-Bond Universal [ABU] and G-Bond Plus [GBP]), using two bonding strategies (etch-and-rinse [ER] and self-etch [SE]) with or without an additional hydrophobic resin coating [HE]. For SBU and ABU, lower NL was observed in the ER strategy when HE was used (B and F) compared with their controls (A and E). However, for the SE strategy, nanoleakage occurred similarly within the hybrid layer (HL) for both ABU and SBU using or not HE (white arrows) (C, D, G and H). GBP showed higher amount of nanoleakage under the ER strategy in the hybrid layer (white arrows) (I), when compared with SBU and ABU; most silver nitrate uptake occurred throughout the entire thickness of the HL (white arrows) and in the adhesive layer (AL), forming the so-called “water trees” (white pointers) (J and K). Only in the SE strategy with HE, the nanoleakage was lower and similar to SBU and ABU (D, H and L).

grains. Specimens were polished with wet 600-, 1000-, 1200-, 1500-, 2000- and 2500-grit SiC paper and 0.25 μm diamond paste (Buehler Ltd., Lake Bluff, IL, USA) using a polishing cloth. Specimens were then ultrasonically cleaned, air dried, mounted on Al stubs, and coated with carbon-gold (MED 010, Balzers Union, Balzers, Liechtenstein). Resin–dentine interfaces were analyzed in a field-emission scanning electron microscope operated in the backscattered mode (LEO 435 VP, LEO Electron Microscopy Ltd., Cambridge, UK).

Three micrographs of each resin–dentine bonded beam were obtained. A blinded operator measured the relative percentage of NL of the adhesive interface in each micrograph with the UTHSCSA ImageTool 3.0 software (University of Texas Health Science Center, San Antonio, TX, USA). The values originating from the same specimen were averaged for statistical purposes. The mean NL of all beams from the same tooth was averaged for statistical purposes.

2.4. Degree of conversion in situ (DC)

The adhesive interface of two resin–dentine bonded beam from each tooth was wet polished with 1500-, 2000-, and 2500-grit SiC paper for 15 s each. Then they were ultrasonically cleaned for 20 min in distilled water and stored in water for 24 h at 37 °C. The micro-Raman spectrometer (Senterra, Bruker Optik GmbH, Ettlingen, Baden–Württemberg, Germany)

was first calibrated for zero and then for coefficient values using a silicon specimen. The following micro-Raman parameters were employed: 20 mW Neon laser with 532 nm wavelength, spatial resolution of $\approx 3 \mu\text{m}$, spectral resolution $\approx 5 \text{ cm}^{-1}$, accumulation time of 30 s with 6 co-additions, and magnification of 100 \times (Olympus UK, London, UK) to a $\approx 1 \mu\text{m}$ beam diameter. Spectra were taken at the dentine–adhesive and enamel–adhesive interface at three different sites for each specimen and an average per tooth was performed. Spectra of uncured adhesives were taken as reference. Post-processing of spectra was performed using the dedicated Opus Spectroscopy Software version 6.5 (Bruker Optik GmbH, Ettlingen, Baden Württemberg, Germany). The ratio of double-bond content of monomer to polymer in the adhesive was calculated according to the following formula:

$$\text{DC (\%)} = \left(1 - \frac{R_{(\text{cured})}}{R_{(\text{uncured})}} \right) \times 100$$

where “R” is the ratio of aliphatic and aromatic peak areas at 1639 cm^{-1} and 1609 cm^{-1} in cured and uncured adhesives.

2.5. Statistical analysis

The Kolmogorov–Smirnov test was used to assess whether the data followed a normal distribution. The Barlett’s test was computed to determine if the assumption of equal variances

Table 1 – Adhesive materials (batch number), composition and application mode of the adhesive systems used.

Materials/batch number	Composition	Application mode			
		Self-etch		Etch-and-rinse	
		Without hydrophobic resin coating	With hydrophobic resin coating	Without hydrophobic resin coating	With hydrophobic resin coating
Heliobond – HE (N37749)	bis-GMA, TEGDMA, initiators, stabilizers				
Scotchbond Universal Adhesive – SBU (448716)	1. Etchant: 34% phosphoric acid, water, synthetic amorphous silica, polyethylene glycol, aluminium oxide (Scotchbond Universal Etchant) 2. Adhesive: MDP phosphate monomer, dimethacrylate resins, HEMA, methacrylate-modified polyalkenoic acid copolymer, filler, ethanol, water, initiators, and silane	1. Apply the adhesive to the entire preparation with a microbrush and rub it in for 20 s 2. Direct a gentle stream of air over the liquid for about 5 s until it no longer moves and the solvent is evaporated completely 3. Light-cure for 10 s	1. Apply the same self-etch mode 2. Apply a very thin layer of Heliobond with a microbrush on the dental surface 3. An air blower to achieve an optimally thin layer 4. Light-cure for 10 s	1. Apply etchant for 15 s 2. Rinse for 10 s 3. Air dry 5 s 4. Apply adhesive as for the self-etch mode	1. Apply the same etch-and-rinse mode 2. Apply a very thin layer of Heliobond with a microbrush on the dental surface 3. An air blower to achieve an optimally thin layer 4. Light-cure for 10 s
All-Bond Universal – ABU (1200002722)	1. Etchant: 35% phosphoric acid, benzalkonium chloride (SELECT HV-Etch with BAC) 2. Adhesive: MDP, bis-GMA, HEMA, ethanol, water, initiators	1. Apply two separate coats of adhesive, scrubbing the preparation with a microbrush for 10–15 s per coat. Do not light cure between coats 2. Evaporate excess solvent by thoroughly air-drying with an air syringe for at least 10 s, there should be no visible movement of the material. The surface should have a uniform glossy appearance 3. Light cure for 10 s	1. Apply the same self-etch mode 2. Apply a very thin layer of Heliobond with a microbrush on the dental surface 3. An air blower to achieve an optimally thin layer 4. Light-cure for 10 s	1. Apply etchant for 15 s 2. Rinse thoroughly 3. Remove excess water with absorbent pellet or high volume suction for 1–2 s 4. Apply adhesive as for the self-etch mode	1. Apply the same etch-and-rinse mode 2. Apply a very thin layer of Heliobond with a microbrush on the dental surface 3. An air blower to achieve an optimally thin layer 4. Light-cure for 10 s
G-Bond Plus – GBP (1102221)	Acetone, dimethacrylate, 4 methacryloxyethyltrimellitate anhydride, phosphoric acid ester monomer, silicon dioxide, photo initiator, distilled water Note: the manufacturer does not recommend dentine conditioning with phosphoric acid	1. Apply using a microbrush 2. Leave undisturbed for 10 s after the end of application 3. Dry thoroughly for 5 s with oil free air under maximum air pressure. Use vacuum suction to prevent splatter of the adhesive 4. Light-cure for 10 s	1. Apply the same self-etch mode 2. Apply a very thin layer of Heliobond with a microbrush on the dental surface 3. An air blower to achieve an optimally thin layer 4. Light-cure for 10 s	1. Apply 34% phosphoric acid gel for 10 s (Scotchbond Universal Etchant) 2. Rinse for 5 s and gently dry 3. Apply adhesive as for the self-etch mode	1. Apply the same etch-and-rinse mode 2. Apply a very thin layer of Heliobond with a microbrush on the dental surface 3. An air blower to achieve an optimally thin layer 4. Light-cure for 10 s
HEMA, 2-hydroxyethyl methacrylate; MDP, methacryloyloxydecyl dihydrogen phosphate; Bis-GMA, bisphenol glycidyl methacrylate; TEGDMA, triethylene glycol dimethacrylate.					

175
176
177
178
179
180
181
182
183
184
185
186
187
188
189
190
191
192

192
193
194
195
196
197
198
199
200
201
202
203
204
205
206
207
208
209

Please cite this article in press as: Muñoz MA, et al. Influence of a hydrophobic resin coating on the bonding efficacy of three universal adhesives. Journal of Dentistry (2014), <http://dx.doi.org/10.1016/j.jdent.2014.01.013>

Table 2 – Number (%) of specimens according to fracture pattern mode for the experimental groups.

Adhesive system	Application mode	Hydrophobic bond	Fracture pattern			
			A	C	A/M	PF
SBU	ER	Without	48 (70.6)	0 (0.0)	14 (20.6)	6 (8.8)
		With	45 (95.8)	1 (2.1)	0 (0.0)	1 (2.1)
	SE	Without	51 (83.6)	2 (3.3)	6 (9.8)	2 (3.3)
		With	55 (80.9)	4 (5.9)	1 (1.5)	8 (11.7)
ABU	ER	Without	42 (80.8)	0 (0.0)	8 (15.4)	2 (3.8)
		With	47 (92.2)	2 (3.9)	2 (3.9)	0 (0.0)
	SE	Without	50 (79.4)	0 (0.0)	10 (15.9)	3 (4.7)
		With	48 (80.0)	1 (1.7)	7 (11.7)	4 (6.6)
GBP	ER	Without	48 (77.5)	2 (3.2)	8 (12.9)	4 (6.4)
		With	50 (83.3)	1 (1.7)	8 (13.3)	1 (1.7)
	SE	Without	48 (87.3)	1 (1.8)	4 (7.3)	2 (3.6)
		With	52 (94.6)	1 (1.8)	0 (0.0)	2 (3.6)

A, adhesive fracture mode; C, cohesive fracture mode; A/M, adhesive/mixed fracture mode; PF, premature failure.

was valid. After observing the normality of the data distribution and the equality of the variances, data from μ TBS, NL, and *in situ* DC were analyzed separately using two-way ANOVA having the “adhesive strategy” and “hydrophobic resin coating” as independent variables for each adhesive system. Tukey’s *post hoc* test was used for pairwise comparisons at a level of significance of 5%.

3. Results

Most of the specimens showed adhesive/mixed failures. Cohesive and premature failures were observed in few specimens (Table 2).

3.1. Microtensile bond strength (μ TBS)

When used under manufacturers’ directions, higher mean μ TBS were obtained with the ER mode for ABU and GBP (Table 3;

$p = 0.0001$ and 0.01 , respectively). The use of a hydrophobic resin coating increased the μ TBS of the adhesives only in the SE mode (Table 3; $p < 0.01$).

3.2. Nanoleakage (NL)

None of the conditions resulted in nanoleakage-free interfaces (Table 4). Significant reductions in the NL were only observed for SBU and ABU in the ER mode when a hydrophobic resin coating was applied (Table 4; $p = 0.01$ and $p = 0.007$, respectively). For GBP, the highest NL was observed in the SE mode, which was significantly reduced with the hydrophobic resin coating (Table 4; $p = 0.001$).

3.3. Degree of conversion (DC)

When used as per manufacturers’ instructions, GBP and SBU resulted in statistically higher DC when applied in the ER mode than when applied in the SE mode. The addition of a

Table 3 – Microtensile bond strength (μ TBS) values (means \pm standard deviations) of the different experimental groups.*

Adhesives	ER		SE	
	Without coating	With coating	Without coating	With coating
SBU	32.3 \pm 3.7B	34.6 \pm 4.1B	34.7 \pm 5.8B	42.7 \pm 6.1A
ABU	40.8 \pm 5.0b	36.3 \pm 4.4c	22.0 \pm 5.1d	48.4 \pm 5.9a
GBP	20.5 \pm 3.2 ^b	22.6 \pm 4.6 ^b	11.5 \pm 3.3 ^a	22.0 \pm 4.6 ^b

Comparisons are valid only within rows.

* Means identified with the identical letters are statistically similar ($p > 0.05$).

Table 4 – Nanoleakage (NL) values (means \pm standard deviations) of the different experimental groups.*

Adhesives	ER		SE	
	Without coating	With coating	Without coating	With coating
SBU	6.2 \pm 2.3B	1.3 \pm 1.1A	6.7 \pm 3.0B	5.9 \pm 2.2B
ABU	11.2 \pm 2.6c	4.3 \pm 1.1a	7.3 \pm 3.4a,b	10.1 \pm 2.7b,c
GBP	14.8 \pm 3.8 ^b	15.3 \pm 2.3 ^b	39.9 \pm 2.6 ^c	7.4 \pm 2.3 ^a

Comparisons are valid only within rows.

* Means identified with identical letters are statistically similar ($p > 0.05$).

Table 5 – Degree of conversion (DC) values (means ± standard deviations) of the different experimental groups.*

Adhesives	ER		SE	
	Without coating	With coating	Without coating	With coating
SBU	86.2 ± 4.6A,B	92.9 ± 4.3A	63.9 ± 5.2C	81.4 ± 4.6B
ABU	76.3 ± 4.1a	80.9 ± 4.1a	75.2 ± 1.9a	81.9 ± 5.1a
GBP	67.3 ± 4.5 ^c	92.1 ± 3.9 ^a	57.3 ± 4.7 ^d	87.7 ± 3.9 ^b

* Comparisons are valid only within rows.

^a Means identified with identical letters are statistically similar ($p > 0.05$).

hydrophobic resin coating increased the DC of GBP and SBU for either bonding strategy. However, this increase was not statistically significant for the ER strategy. The adhesive strategy and the hydrophobic resin coating did not affect the DC of ABU (Table 5; $p = 0.45$).

4. Discussion

The performance of the universal adhesives used in this study was dependent on the adhesive strategy. The results of the μ TBS testing showed that two of the three adhesive systems showed improved resin–dentine bond strengths when used in the ER mode, following manufacturers' directions. The presence of the smear layer imposes a physical barrier against the penetration of resin monomers.³ Phosphoric acid etching removes the smear layer, promotes superficial dentine demineralization²⁶ and increases the impregnation of resin monomers, which leads to the formation of a thick hybrid layer fully integrated with the dentine.^{5,6,26–28} The findings of this study along with the improved bond strength of 1-step SE adhesives after previous phosphoric acid etching^{5,6,27,28} suggests that the smear layer incorporation into the adhesive interface reduces bonding effectiveness.

We hypothesize that the demineralization by-products, as well as the smear layer remnants, may have acted as contaminants, preventing the polymer chains from getting closer during the polymerization reaction. This would explain why using the same adhesive formulation under identical light-curing protocols for two out of the three adhesive systems, the *in situ* DC was lower when the materials were used in the SE mode.

While the hydrophobic resin coating HE did not improve mean μ TBS under the ER strategy, HE increased mean μ TBS in the SE mode, which reached the magnitude of mean μ TBS obtained in the ER approach. As HE also resulted in an increase in the *in situ* DC of adhesives applied in the SE mode (yet not statistically significant for ABU), we have to partially reject the null hypothesis. Other studies also detected higher DC and lower adhesive permeability for adhesive systems which use an extra hydrophobic resin coating.^{11,29} This was also observed in a recent study, in which Peak Universal Adhesive, a 2-step SE adhesive, was superior in terms of DC than its 1-step counterparts (Scotchbond Universal, All-Bond Universal).⁷

The extra layer of hydrophobic resin adds unsolvated hydrophobic monomers to the adhesive interface, which decreases the relative concentration of retained solvents and unreacted monomers in the adhesive layer.¹⁸ This, in turn,

increases the ultimate tensile strength of the adhesive interface,¹⁰ due to the formation of a more densely packed hybrid layers,²⁰ making it more resistant to the tensile forces during μ TBS testing and less prone to degradation effects over time.^{30,31}

Although earlier studies have reported that the permeability of the adhesive interfaces can be reduced by the application of a hydrophobic resin coating,²⁰ the NL of two out of the three adhesives tested in the SE mode was not significantly influenced in our study when HE was applied. This may be a result of NL being not only related to the permeability of the adhesive layer, but also to submicron defects, such as solvent/water entrapment, incomplete resin infiltration, or areas of inadequate polymerization.^{25,30} For instance, the fact that only GBP applied in the SE mode resulted in a very low DC and a very high NL, lead us to speculate that excessive NL of the GBP interface might be related to incomplete polymerization of the adhesive interface. This may be the reason why the additional hydrophobic resin coating benefited GBP in SE mode, increasing the DC and simultaneously decreasing the NL.

The poorest performance of GBP in all tests in the present study, as well as an earlier study,⁶ may also be attributed to the application mode. Contrary to SBU and ABU, which were applied actively, GBP was the only material for which the manufacturer does not recommend active application. It is consensual that an active adhesive application increases the resin–dentine bond strength.^{32–38} Several factors may account for this effect. It is believed that an increased penetration of monomers into dentine substrates as well as higher solvent evaporation occurs under active application. Agitation also improves the polymerization efficacy through enhancing the chemical interaction with tooth substrate, mainly for acidic SE adhesives.^{35,39} Additionally, unreacted acidic resin monomers from the superficial layer of the adhesive can be taken to the basal portion of dentine, increasing the substrate demineralization, monomer diffusion and better interaction with the smear layer and underlying dentine.^{36–38}

Apart from the application mode, compositional differences between adhesives may play an important role on the adhesive performance. SBU and ABU contain 10-methacryloyloxydecyl dihydrogen phosphate monomer (MDP) capable to chemically interact with the hydroxyapatite by the formation of a stable nano-layering at the adhesive interface^{40–42} GBP has a lower pH value (pH = 1.5) than the other adhesives (SBU = 3; ABU = 2.4).⁷ The hydrolysis of the ester bond from the acid monomer results in a strong phosphoric acid that might increase the demineralization over time and explain the intense NL within the hybrid layer.^{7,43}

An overall analysis of the three adhesives shows that SBU performed generally similar when used in the ER and SE mode. As mentioned in an earlier study,⁷ SBU has polyalkenoic acid copolymer and MDP in its formulation. Although both molecules may compete by binding to the calcium in hydroxyapatite,⁴¹ they are usually associated with improved adhesive performance^{44,45} which may explain the good performance of SBU in both bonding strategies.

In summary, the use of a hydrophobic resin coating may improve the performance of the universal adhesives mainly when applied as SE adhesives.

5. Conclusions

With the limitations of this study the use of an additional hydrophobic resin coating improved the adhesive performance in terms of resin–dentine bond strengths of new universal adhesives when used with the SE strategy. The additional hydrophobic resin coating also improved the DC for both the ER and the SE strategies. The reduction in NL may be more dependent on the adhesive composition than on the adhesive strategy. In spite of the improvements observed for universal adhesives *in vitro*, longevity studies should be performed to validate these initial results.

REFERENCES

1. Peumans M, Kanumilli P, De Munck J, Van Landuyt K, Lambrechts P, Van Meerbeek B. Clinical effectiveness of contemporary adhesives: a systematic review of current clinical trials. *Dental Materials* 2005;**21**:864–81.
2. De Munck J, Van Landuyt K, Peumans M, Poitevin A, Lambrechts P, Braem M, et al. A critical review of the durability of adhesion to tooth tissue: methods and results. *Journal of Dental Research* 2005;**84**:118–32.
3. Van Meerbeek B, De Munck J, Yoshida Y, Inoue S, Vargas M, Vijay P, et al. Buonocore memorial lecture. Adhesion to enamel and dentin: current status and future challenges. *Operative Dentistry* 2003;**28**:215–35.
4. Yiu CK, Pashley EL, Hiraishi N, King NM, Goracci C, Ferrari M, et al. Solvent and water retention in dental adhesive blends after evaporation. *Biomaterials* 2005;**26**:6863–72.
5. Perdigao J, Sezinando A, Monteiro PC. Laboratory bonding ability of a multi-purpose dentin adhesive. *American Journal of Dentistry* 2012;**25**:153–8.
6. Hanabusa M, Mine A, Kuboki T, Momoi Y, Van Ende A, Van Meerbeek B, et al. Bonding effectiveness of a new ‘multi-mode’ adhesive to enamel and dentine. *Journal of Dentistry* 2012;**40**:475–84.
7. Munoz MA, Luque I, Hass V, Reis A, Loguercio AD, Bombarda NH. Immediate bonding properties of universal adhesives to dentine. *Journal of Dentistry* 2013;**41**:404–11.
8. Paul SJ, Leach M, Rueggeberg FA, Pashley DH. Effect of water content on the physical properties of model dentine primer and bonding resins. *Journal of Dentistry* 1999;**27**:209–14.
9. Ye Q, Spencer P, Wang Y, Misra A. Relationship of solvent to the photopolymerization process, properties, and structure in model dentin adhesives. *Journal of Biomedical Materials Research Part A* 2007;**80**:342–50.
10. Loguercio AD, Loeblein F, Cherobin T, Oglari F, Piva E, Reis A. Effect of solvent removal on adhesive properties of simplified etch-and-rinse systems and on bond strengths to dry and wet dentin. *Journal of Adhesive Dentistry* 2009;**11**:213–9.
11. Cadenaro M, Antonioli F, Sauro S, Tay FR, Di Lenarda R, Prati C, et al. Degree of conversion and permeability of dental adhesives. *European Journal of Oral Sciences* 2005;**113**:525–30.
12. Hass V, Folkuenig MS, Reis A, Loguercio AD. Influence of adhesive properties on resin–dentin bond strength of one-step self-etching adhesives. *Journal of Adhesive Dentistry* 2011;**13**:417–24.
13. Reis A, Ferreira SQ, Costa TR, Klein-Junior CA, Meier MM, Loguercio AD. Effects of increased exposure times of simplified etch-and-rinse adhesives on the degradation of resin–dentin bonds and quality of the polymer network. *European Journal of Oral Sciences* 2010;**118**:502–9.
14. Takahashi A, Sato Y, Uno S, Pereira PN, Sano H. Effects of mechanical properties of adhesive resins on bond strength to dentin. *Dental Materials* 2002;**18**:263–8.
15. Cho BH, Dickens SH. Effects of the acetone content of single solution dentin bonding agents on the adhesive layer thickness and the microtensile bond strength. *Dental Materials* 2004;**20**:107–15.
16. Malacarne J, Carvalho RM, de Goes MF, Svizero N, Pashley DH, Tay FR, et al. Water sorption/solubility of dental adhesive resins. *Dental Materials* 2006;**22**:973–80.
17. Ito S, Hashimoto M, Wadgaonkar B, Svizero N, Carvalho RM, Yiu C, et al. Effects of resin hydrophilicity on water sorption and changes in modulus of elasticity. *Biomaterials* 2005;**26**:6449–59.
18. Breschi L, Mazzoni A, Ruggeri A, Cadenaro M, Di Lenarda R, De Stefano Dorigo E. Dental adhesion review: aging and stability of the bonded interface. *Dental Materials* 2008;**24**:90–101.
19. King NM, Tay FR, Pashley DH, Hashimoto M, Ito S, Brackett WW, et al. Conversion of one-step to two-step self-etch adhesives for improved efficacy and extended application. *American Journal of Dentistry* 2005;**18**:126–34.
20. de Andrade e Silva SM, Carrilho MR, Marquezini Junior L, Garcia FC, Manso AP, Alves MC, et al. Effect of an additional hydrophilic versus hydrophobic coat on the quality of dentinal sealing provided by two-step etch-and-rinse adhesives. *Journal of Applied Oral Science* 2009;**17**:184–9.
21. Reis A, Leite TM, Matte K, Michels R, Amaral RC, Geraldini S, et al. Improving clinical retention of one-step self-etching adhesive systems with an additional hydrophobic adhesive layer. *Journal of the American Dental Association* 2009;**140**:877–85.
22. Reis A, Albuquerque M, Pegoraro M, Mattei G, Bauer JR, Grande RH, et al. Can the durability of one-step self-etch adhesives be improved by double application or by an extra layer of hydrophobic resin? *Journal of Dentistry* 2008;**36**:309–15.
23. Pashley DH, Tao L, Boyd L, King GE, Horner JA. Scanning electron microscopy of the substructure of smear layers in human dentine. *Archives of Oral Biology* 1988;**33**:265–70.
24. Perdigao J, Geraldini S, Carmo AR, Dutra HR. *In vivo* influence of residual moisture on microtensile bond strengths of one-bottle adhesives. *Journal of Esthetic and Restorative Dentistry* 2002;**14**:31–8.
25. Tay FR, Pashley DH, Yoshiyama M. Two modes of nanoleakage expression in single-step adhesives. *Journal of Dental Research* 2002;**81**:472–6.
26. Kenshima S, Reis A, Uceda-Gomez N, Tancredo Lde L, Filho LE, Nogueira FN, et al. Effect of smear layer thickness and pH of self-etching adhesive systems on the bond strength and gap formation to dentin. *Journal of Adhesive Dentistry* 2005;**7**:117–26.

- 425 27. Proenca JP, Polido M, Osorio E, Erhardt MC, Aguilera FS, 458
 426 Garcia-Godoy F, et al. Dentin regional bond strength of self- 459
 427 etch and total-etch adhesive systems. *Dental Materials* 460
 428 2007;**23**:1542-8. 461
- 429 28. Tschner M, Nato F, Mazzoni A, Frankenberger R, Falconi M, 462
 430 Petschelt A, et al. Influence of preliminary etching on the 463
 431 stability of bonds created by one-step self-etch bonding 464
 432 systems. *European Journal of Oral Sciences* 2012;**120**:239-48. 465
- 433 29. Breschi L, Cadenaro M, Antonioli F, Sauro S, Biasotto M, 466
 434 Prati C, et al. Polymerization kinetics of dental adhesives 467
 435 cured with LED: correlation between extent of conversion 468
 436 and permeability. *Dental Materials* 2007;**23**:1066-72. 469
- 437 30. Reis A, Grande RH, Oliveira GM, Lopes GC, Loguercio AD. A 470
 438 2-year evaluation of moisture on microtensile bond strength 471
 439 and nanoleakage. *Dental Materials* 2007;**23**:862-70. 472
- 440 31. Reis A, de Carvalho Cardoso P, Vieira LC, Baratieri LN, 473
 441 Grande RH, Loguercio AD. Effect of prolonged application 474
 442 times on the durability of resin-dentin bonds. *Dental* 475
 443 *Materials* 2008;**24**:639-44. 476
- 444 32. Pleffken PR, de Almeida Lourenco AP, Torres CR, Buhler 477
 445 Borges A. Influence of application methods of self-etching 478
 446 adhesive systems on adhesive bond strength to dentin. 479
 447 *Journal of Adhesive Dentistry* 2011;**13**:517-25. 480
- 448 33. Loguercio AD, Stanislawczuk R, Mena-Serrano A, Reis A. 481
 449 Effect of 3-year water storage on the performance of one- 482
 450 step self-etch adhesives applied actively on dentine. *Journal* 483
 451 *of Dentistry* 2011;**39**:578-87. 484
- 452 34. Zhang Y, Wang Y. Distinct photopolymerization efficacy on 485
 453 dentin of self-etch adhesives. *Journal of Dental Research* 486
 454 2012;**91**:795-9. 487
- 455 35. Zhang Y, Wang Y. Effect of application mode on interfacial 488
 456 morphology and chemistry between dentine and self-etch 489
 457 adhesives. *Journal of Dentistry* 2013;**41**:231-40. 490
 491
36. do Amaral RC, Stanislawczuk R, Zander-Grande C, Michel 458
 MD, Reis A, Loguercio AD. Active application improves the 459
 bonding performance of self-etch adhesives to dentin. 460
Journal of Dentistry 2009;**37**:82-90. 461
37. do Amaral RC, Stanislawczuk R, Zander-Grande C, Gagler D, 462
 Reis A, Loguercio AD. Bond strength and quality of the 463
 hybrid layer of one-step self-etch adhesives applied with 464
 agitation on dentin. *Operative Dentistry* 2010;**35**:211-9. 465
38. Miyazaki M, Platt JA, Onose H, Moore BK. Influence of dentin 466
 primer application methods on dentin bond strength. 467
Operative Dentistry 1996;**21**:167-72. 468
39. Wang Y, Spencer P. Hybridization efficiency of the adhesive/ 469
 dentin interface with wet bonding. *Journal of Dental Research* 470
 2003;**82**:141-5. 471
40. Yoshihara K, Yoshida Y, Nagaoka N, Fukegawa D, Hayakawa 472
 S, Mine A, et al. Nano-controlled molecular interaction at 473
 adhesive interfaces for hard tissue reconstruction. *Acta* 474
Biomaterialia 2010;**6**:3573-82. 475
41. Yoshida Y, Yoshihara K, Nagaoka N, Hayakawa S, Torii Y, 476
 Ogawa T, et al. Self-assembled nano-layering at the 477
 adhesive interface. *Journal of Dental Research* 2012;**91**:376-81. 478
42. Inoue S, Koshiro K, Yoshida Y, De Munck J, Nagakane K, 479
 Suzuki K, et al. Hydrolytic stability of self-etch adhesives 480
 bonded to dentin. *Journal of Dental Research* 2005;**84**:1160-4. 481
43. Wang Y, Spencer P. Continuing etching of an all-in-one 482
 adhesive in wet dentin tubules. *Journal of Dental Research* 483
 2005;**84**:350-4. 484
44. Perdigao J, Sezinando A, Monteiro PC. Effect of substrate age 485
 and adhesive composition on dentin bonding. *Operative* 486
Dentistry 2013;**38**:267-74. 487
45. Van Meerbeek B, Yoshihara K, Yoshida Y, Mine A, De Munck 488
 J, Van Landuyt KL. State of the art of self-etch adhesives. 489
Dental Materials 2011;**27**:17-28. 490
 491

5. DISCUSSÃO

Apesar de todos os novos sistemas adesivos universais testados compartilharem a mesma versatilidade de aplicação como CON ou AC, houve diferenças no desempenho destes materiais quando comparados com os controles o que parece estar associado às suas diferentes composições e a seus números de passos. Entretanto, quando se aplicou uma camada hidrofóbica adicional aos adesivos testados, seu desempenho tornou-se dependente principalmente da estratégia adesiva.

Com base nas semelhanças nas suas composições, esperava-se, no tempo imediato, que os adesivos universais ScAc e AIAC fossem equivalentes ao controle CSE, no entanto, isto não foi observado. Estes três materiais possuem o monômero 10-MDP em sua composição, portanto, unem-se quimicamente à dentina (Inoue et al.⁶⁶ 2005; Waidyasekera et al.⁶⁷ 2009; Peumans et al.⁶⁸ 2010). Yoshida e cols. (Yoshida et al.⁶⁹ 2012) mostraram uma eficaz interação química entre o MDP e a hidroxiapatita formando uma nanocamada estável na interface adesiva, aumentando a resistência mecânica da mesma. A deposição de sais estáveis de MDP-Ca junto à nano-camada tem permitido sua alta estabilidade (Yoshida et al.⁶⁹ 2012; Yoshihara et al.⁷⁰ 2010), a qual tem sido comprovada em pesquisas clínicas e laboratoriais (Peumans et al.⁶⁸ 2010; Toledano et al.⁷¹ 2007; Feitosa et al.⁷² 2012). O melhor comportamento em termos de RU do controle CSE, quando comparado com ScAc, pode se dever à presença do copolímero de ácido polialquenoico (PAC) na composição do ScAc. Este copolímero pode ter competido com o MDP pela união ao cálcio da hidroxiapatita prejudicando levemente a união química do MDP à dentina (Yoshida et al.⁶⁹ 2012). Além disso, o copolímero PAC poderia ter impedido a aproximação dos monômeros durante a polimerização devido a seu alto peso molecular, tendo como consequência, um menor GC deste adesivo. Adicionalmente, ScAc e AIAC são adesivos de um passo e isso, provavelmente, faz com que sua concentração de MDP seja menor que a do CSE, que tem este monômero MDP incorporado na composição tanto do *primer* quanto do *bond* (Yoshida et al.⁶⁹ 2012).

Ainda entre os adesivos AC dos experimentos 1 e 2, tanto no tempo imediato, quanto após 6 meses, AIAC mostrou o pior desempenho em termos de RU, o que poderia ser atribuído ao seu conteúdo de solvente. O adesivo AI contém maior

porcentagem de solvente, quando comparado com Sc (30-60% peso e 10-15% peso, respectivamente) (Chen, Suh⁷³ 2013; 3MESPE⁷⁴ 2013). Isto leva a uma maior quantidade de solvente residual retido nas camadas adesiva e híbrida (Yiu et al.⁹ 2005), prejudicando a formação de um polímero com alta reticulação (Ye et al.³⁹ 2007; Loguercio et al.⁴⁰ 2009; Paul et al.⁴¹ 1999), portanto reduzindo o grau de conversão (Cadenaro et al.⁴² 2005) e a resistência de união dente-resina (Hass et al.⁴³ 2011; Reis et al.⁴⁴ 2010; Takahashi et al.⁴⁵ 2002; Cho, Dickens⁴⁶ 2004) e aumentando a permeabilidade da camada adesiva após a polimerização (Malacarne et al.⁴⁷ 2006; Ito et al.⁴⁸ 2005)

Outro fator que pode ter contribuído para o comportamento inferior de AIAC foi seu modo de aplicação. No experimento 1, AIAC foi esfregado mas não aplicado vigorosamente, de acordo com nossa interpretação das instruções do respectivo fabricante, situação habitual na prática clínica. Alguns autores apontam claramente que a aplicação ativa/vigorosa melhora o desempenho da adesão de sistemas adesivos AC à dentina (Pleffken et al.⁷⁵ 2011; Loguercio et al.⁷⁶ 2011). Acredita-se que com a aplicação ativa aumenta-se tanto a penetração dos monômeros na dentina quanto a evaporação do solvente. A agitação também melhoraria a eficácia de polimerização através da melhoria na interação química do adesivo com o substrato dentário, principalmente para adesivos AC ácidos (Wang, Spencer⁵⁰ 2003; Zhang, Wang⁷⁷ 2013). Além disso, monômeros ácidos não reagidos presentes na camada superficial do adesivo podem ser levados para uma área basal da dentina, aumentando a desmineralização do substrato e a difusão dos monômeros, melhorando a interação com a *smear layer* e a dentina subjacente (do Amaral et al.⁷⁸ 2009; do Amaral et al.⁷⁹ 2010; Miyazaki et al.⁸⁰ 1996). Assim, no experimento 2, o AIAC foi aplicado vigorosamente o que aumentou sua RU, porém, se manteve ainda como a mais baixa entre os adesivos universais. Já no experimento 3, o GBP foi o único adesivo cujo fabricante indica especificamente não esfregar e deixar em repouso e, conseqüentemente, apresentou uma pobre performance em todos os testes. Quando GBP foi aplicado ativamente em um recente estudo (Hanabusa et al.²⁹ 2012) mostrou melhores resultados, o que corrobora que uma aplicação ativa de adesivo aumenta a RU resina-dentina e a estabilidade ao longo do tempo (Pleffken et al.⁷⁵ 2011; Loguercio et al.⁷⁶ 2011; Zhang, Wang⁷⁷ 2013; do Amaral et al.⁷⁸ 2009;

Miyazaki et al.⁸⁰ 1996; Zhang, Wang⁸¹ 2012). Futuros estudos precisam ser realizados para testar isoladamente esta hipótese com sistemas adesivos universais.

Entre todos os adesivos usados com a estratégia AC nos experimentos 1 e 2, PkAc foi o único adesivo que, no tempo imediato, apresentou valores de RU semelhantes ao controle CSE e superiores aos demais adesivos estudados, apesar de não apresentar MDP na sua composição. Tanto estudos *in vitro* (Reis et al.⁶² 2008; Feitosa et al.⁷² 2012; Frankenberger et al.⁸² 2001; Albuquerque et al.⁸³ 2008) como clínicos (Reis et al.⁶³ 2009; Loguercio, Reis⁸⁴ 2008) tem mostrado que a aplicação de uma camada adicional melhora o desempenho de adesivos AC de 1-passo, sendo essa uma vantagem do PkAc sobre ScAc e AIAC. Em geral, adesivos AC de 1-passo apresentam um desempenho inferior aos adesivos AC de 2 passos (Van Meerbeek et al.⁸ 2003; Perdigão⁸⁵ 2007). Adesivos AC de 1-passo resultam em finas camadas de adesivo, sendo propensas à inibição de polimerização por oxigênio (Nunes et al.⁸⁶ 2005). Este resultado imediato positivo do adesivo PkAc, entretanto, não foi estável ao longo de seis meses. Como essa degradação ocorreu tanto para a estratégia AC quanto para a CON, as possíveis justificativas para esse comportamento estão discutidas posteriormente.

Os adesivos com MPD normalmente são associados com um melhor desempenho de união (Van Meerbeek et al.⁸⁷ 2011; Perdigão et al.⁸⁸ 2013). No que se refere à estratégia AC, foi verificado em nossos resultados que, após 6 meses, tanto a RU quanto a NI dos adesivos CSE, AIAC e ScAc se mantiveram estáveis. Um dos fatores favoráveis desses materiais é o seu pH em uma faixa em torno de 2, que produz desmineralização parcial da dentina, deixando quantidade substancial de cristais de hidroxiapatita ao redor das fibrilas de colágeno (Tay, Pashley⁸⁹ 2001), favorecendo sua interação química com o MDP. Essa interação, juntamente com os fatores anteriormente citados, poderia explicar o comportamento dos adesivos estudados, segundo a estratégia AC.

No que diz respeito à abordagem CON, esta pareceu ser favorável no tempo imediato, pois os adesivos universais mostraram melhor RU em comparação com a estratégia AC, exceto para ScCon, que foi semelhante a ScAc. Sabe-se que a *smear layer* constitui uma verdadeira barreira física que torna extremamente difícil a conformação de uniões e formação de uma camada híbrida totalmente integrada

com a dentina (Kenshima et al.⁹⁰ 2005). Após o condicionamento com ácido fosfórico, a *smear layer* é removida e a superfície dentinária é desmineralizada. Isto facilitaria a infiltração do adesivo, permitindo a criação de uma camada híbrida, como tem sido demonstrado em adesivos AC de 1-passo aplicados em dentina previamente condicionada com ácido fosfórico (Hanabusa et al.²⁹ 2012; Proenca et al.³¹ 2007; Taschner et al.³² 2012; Perdigao et al.⁶⁴ 2002; Kenshima et al.⁹⁰ 2005). Neste sentido, os resultados do experimento 3, mostraram para o teste de RU que dois (ABU e GBP) dos três sistemas adesivos (ABU, SBU e GBP) melhoraram seus resultados quando utilizados com a estratégia CON, embora o fabricante do GBP não recomende o condicionamento da dentina com ácido fosfórico.

Ainda abordando a RU dos adesivos, se compararmos os resultados após 6 meses do Sc (que contém MDP e PAC) com o SB (que contém somente PAC), podemos observar que a associação MDP-PAC promoveu estabilidade para ScAc e ScCon. Esta estabilidade não pode ser observada no adesivo SB. A mesma falta de estabilidade foi observada para o material AlCon (que contém apenas MDP), o qual demonstrou degradação dos valores de resistência de união após 6 meses. Alguns autores indicam que a função do PAC é melhorar a estabilidade à umidade (Van Meerbeek et al.⁹¹ 1996; Spencer et al.⁹² 2000), que torna sensível a técnica CON, quando comparada com a técnica AC (Perdigao⁸⁵ 2007).

A NI representa a localização de defeitos na interface resina-dentina que podem servir de vias para a degradação da união resina/dentina ao longo do tempo (Sano³⁸ 2006). O nitrato de prata é capaz de ocupar os espaços de tamanho nanométrico presentes ao redor das fibrilas de colágeno expostas, onde os monômeros não conseguiram infiltrar-se, ou onde a água residual não foi deslocada pelo adesivo ou mesmo em áreas com incompleta conversão de monômero (Sano³⁸ 2006). Entre os adesivos dos experimentos 1 e 2, PkCon e PkAc apresentaram as maiores percentagens de NI nas interfaces adesivas e os únicos que aumentaram após 6 meses. Situação similar no tempo imediato foi apresentada no experimento 3 para GBP na estratégia AC. O *primer* do sistema PkAc (Peak SE Primer) e o GBP possuem valores de pH muito baixos (1,2 e 1,5, respectivamente) quando comparados com os outros adesivos (SBU=3; ABU=2,4) (Munoz et al.⁹³ 2013) deixando-os dentro da categoria dos adesivos AC mais agressivos (Van Meerbeek et

al.⁸ 2003; Tay, Pashley⁸⁹ 2001). Tem sido relatado que monômeros ácidos e não polimerizados são capazes de prosseguir desmineralizando a dentina, mesmo após sua polimerização (Carvalho et al.¹⁹ 2005; Wang, Spencer²⁰ 2005). Isto é provavelmente o resultado da hidrólise da união éster do monômero de ácido que resulta em um ácido fosfórico forte (Wang, Spencer²⁰ 2005) o que poderia explicar a intensa deposição de nitrato de prata dentro da camada híbrida destes adesivos.

Embora se pudesse esperar uma reduzida NI no interior da camada de adesivo devido ao fato de PkAc ser de 2 passos, a segunda camada de adesivo parece ser tão hidrofílica como o *primer*, uma vez que monômeros hidrofóbicos não foram listados na composição do *Bond* (Peak LC bond - Quadro 1). Embora o aumento da infiltração de monômeros possa ter ocorrido durante a aplicação de uma camada de adesivo adicional, a natureza desta infiltração é hidrofílica, o que resultou numa extensa NI na camada híbrida. No caso de PkCon, Peak LC Bond (pH=2,0) pode ter provocado um condicionamento adicional e um aumento no padrão de condicionamento do substrato dentinário. Isto provavelmente resultou num aumento da desmineralização e exposição de colágeno (Van Meerbeek et al.⁸ 2003; Liu et al.³⁴ 2011; Brackett et al.⁹⁴ 2011), aumentando a NI (Hashimoto et al.¹⁶ 2000; Wang, Spencer⁵⁰ 2003; Spencer et al.⁹² 2000; Wang, Spencer⁹⁵ 2002) para PkCon.

Com relação à aplicação de uma camada hidrofóbica no experimento 3, esta não melhorou a média de RU para a estratégia CON, porém aumentou a média de RU na estratégia AC (atingindo magnitudes obtidas na estratégia CON), o que tem sido descrito anteriormente em vários estudos com outros adesivos AC (Reis et al.⁶² 2008; Feitosa et al.⁷² 2012; Frankenberger et al.⁸² 2001; Albuquerque et al.⁸³ 2008). Assim também, resultou em um aumento no GC dos adesivos aplicados no modo AC. Outros estudos também detectaram maior GC e menor permeabilidade para os sistemas adesivos que utilizam uma camada hidrofóbica adicional (Cadenaro et al.⁴² 2005; Breschi et al.⁹⁶ 2007). A camada adicional de *Bond* (HE) incorpora monômeros hidrofóbicos não solvatados na interface adesiva, o que diminui a concentração relativa de solventes retidos e monômeros não reagidos na camada adesiva (Breschi et al.³⁵ 2008). Isto, por sua vez, aumenta a resistência máxima à tração da interface adesiva (Loguercio et al.⁴⁰ 2009), devido à formação de uma camada híbrida mais densa ou compactada (de Andrade e Silva et al.⁶¹ 2009), tornando-a mais resistente

às forças de tração durante o teste de RU à microtração e menos propenso a efeitos de degradação ao longo do tempo (Reis et al.⁹⁷ 2007; Reis et al.⁹⁸ 2008).

Embora estudos anteriores tenham relatado que a permeabilidade da interface adesiva pode ser reduzida através da aplicação de uma camada de resina hidrofóbica (de Andrade e Silva et al.⁶¹ 2009), a NI de dois dos três adesivos universais (SBU e ABU) testados no modo AC não foi significativamente influenciada em nosso estudo quando HE foi aplicado. Isto pode ser devido à NI não só se encontrar relacionada à permeabilidade da camada adesiva, mas também a defeitos sub-micrométricos tais como, a retenção de solvente/água, infiltração incompleta de monômeros, ou áreas com polimerização inadequada (Reis et al.⁹⁷ 2007; Tay et al.⁹⁹ 2002). Assim, o fato de apenas o adesivo GBP, quando aplicado na estratégia AC, ter obtido valores muito baixos de GC e muito altos de NI, leva-nos a especular que a NI excessiva poderia estar relacionada com a incompleta polimerização da interface adesiva. Esta pode ser a razão pela qual a camada hidrofóbica beneficiou o adesivo GBP no modo AC, aumentando o GC e diminuindo a NI. O GC para o GBP foi significativamente maior quando uma camada de resina hidrofóbica foi usada. Esta diferença era esperada de acordo com resultados de outros estudos (Sadek et al.¹⁰⁰ 2008; Hamouda et al.¹⁰¹ 2010) onde adesivos AC de 1-passo foram comparados com AC de 2-passos. No entanto, um aumento em GC não foi previamente relatado para um adesivo específico de 1-passo após a adição de uma camada de resina hidrofóbica.

Por outro lado, adesivos AC de 1-passo e CON de 2-passos podem apresentar bolhas por atração de água, produto de osmose através da camada de adesivo polimerizado (Sauro et al.¹⁰² 2009; Tay et al.¹⁰³ 2002). Estas bolhas de osmose são observadas na zona de transição entre a camada de adesivo de AC de 1-passo ou CON de 2-passo e a resina composta (Sauro et al.¹⁰² 2009; Tay et al.¹⁰³ 2002). Esta zona contém monômeros não polimerizados da camada de inibição por oxigênio (Van Landuyt et al.¹⁰⁴ 2008), o que teria resultado na diminuição do GC do GBP. Assim, a camada de resina hidrofóbica teria copolimerizado com a superfície não polimerizada deste adesivo de 1-passo, resultando em maior GC, devido ao consumo de ligações duplas residuais.

Relatos sobre a permeabilidade e sorção de água na camada híbrida e a interface adesiva mostraram uma relação com a hidrofiliabilidade do polímero (Tay, Pashley¹⁰⁵ 2003; Chersoni et al.¹⁰⁶ 2004). As soluções de monômeros de adesivos AC de 1-passo carecem de uma camada de resina não solvatada, que os torna membranas permeáveis (Tay et al.⁶⁵ 2002; Tay et al.¹⁰⁷ 2004; Tay et al.¹⁰⁸ 2004; Tay et al.¹⁰⁹ 2004). Esta estrutura permeável iria permitir uma rápida transudação de fluido dentinal através do adesivo polimerizado (Salz et al.¹¹⁰ 2005), especificamente em adesivos contendo o co-monômero monofuncional 2-HEMA (Hosaka et al.¹¹¹ 2010). Teoricamente, a ausência de HEMA (como em GBP), deveria resultar em estabilidade a longo prazo da interface adesiva (Tay et al.⁶⁵ 2002; Van Landuyt et al.¹⁰⁴ 2008) quando comparado com adesivos contendo HEMA (Takahashi et al.¹¹² 2011) porque a retenção e sorção de água é maior neste último tipo de adesivos (Hosaka et al.¹¹¹ 2010), comprometendo o GC e resultando em uma degradação por hidrólise. Porém, o HEMA atua como um solvente e a sua falta pode resultar na separação de fases na interface adesiva (Van Landuyt et al.¹⁰⁴ 2008; Van Landuyt et al.¹¹³ 2005; Moszner et al.¹¹⁴ 2005) o que tem sido previamente observado (Van Landuyt et al.¹⁰⁴ 2008), podendo ser outro fator limitante para o bom desempenho do adesivo GBP.

A média de RU do GBP foi menor (AC=11,5 MPa; CON=20,5) do que as obtidas por outros grupos de pesquisa (AC=30,5 MPa; CON= 29,4 MPa) (Hanabusa et al.²⁹ 2012; Goracci et al.¹¹⁵ 2011). Tem sido demonstrado que as propriedades mecânicas do GBP são menores do que os de outros adesivos AC de 1-passo (Hosaka et al.¹¹¹ 2010). Takahashi e cols. (Takahashi et al.¹¹⁶ 2002) e Reis e cols. (Reis et al.¹¹⁷ 2005), reportaram que há uma correlação positiva entre a RU à microtração em dentina e a resistência máxima à tração do adesivo, o que explicaria os baixos resultados obtidos com GBP no presente estudo. Adicionalmente, fatores relacionados à metodologia podem ter sido responsáveis por essa diferença nos valores médios de RU. Um estudo relatou valores de RU em dentina inferiores para palito da periferia quando comparados com os provenientes do centro (Loguercio et al.¹¹⁸ 2005). No estudo de Hanabusa e cols. (Hanabusa et al.²⁹ 2012), foram utilizados nove palitos centrais de cada dente para reduzir a variabilidade do substrato, enquanto que em nosso estudo utilizamos palitos de toda a interface. Além disso, a velocidade de tração em nosso estudo foi de 0,5 mm/min, enquanto que

Hanabusa e cols. (Hanabusa et al.²⁹ 2012), usaram 1,0 mm/min. Um estudo comparativo relatou valores superiores de RU a microtração para Clearfil SE Bond, quando foi utilizada uma maior velocidade de tração. Um padrão mais uniforme de tempo-tensão foi observado para 1 mm/min (Poitevin et al.¹¹⁹ 2008). Em outro estudo (Goracci et al.¹¹⁵ 2011), embora os autores tenham utilizado a mesma velocidade de tração de nosso estudo, aplicaram GBP na dentina sob agitação, contrariamente aos nossos procedimentos (Loguercio et al.⁷⁶ 2011; do Amaral et al.⁷⁹ 2010).

Dentro das limitações do presente estudo *in vitro*, considerando os adesivos universais avaliados, apenas os que contém MDP apresentaram melhor longevidade de união à dentina, caracterizado pela manutenção da resistência de união e padrão de nanoinfiltração. Sugere-se a realização de estudos *in vitro*, com maior tempo de acompanhamento, para investigar os efeitos do uso de uma camada de resina hidrofóbica nos adesivos universais. Pode-se esperar alguma disparidade entre os resultados do presente estudo *in vitro* e aqueles obtidos clinicamente. Estudos clínicos randomizados e controlados, ou ainda, baseados na prática clínica (practice-based research), poderiam ser realizados para a comprovação dos desfechos obtidos. Além disso, estudos em outros substratos como esmalte e/ou materiais de restauração seriam necessários para investigar a versatilidade sugerida pelos fabricantes dos adesivos universais testados.

6. CONCLUSÕES

Dentro das limitações do presente estudo, foram possíveis as seguintes conclusões:

1- Os sistemas adesivos universais utilizados nas estratégias convencional e autocondicionante apresentaram um comportamento, no tempo de 24 h, inferior a seus respectivos controles (Clearfill SE Bond e Adper Single Bond 2) em pelo menos uma das propriedades avaliadas (resistência de união, nanoinfiltração e grau de conversão na camada híbrida), sendo dependentes da composição do material para ambas estratégias.

2- Após seis meses, para a estratégia adesiva AC, houve, em geral, estabilidade na resistência de união e nanoinfiltração dos adesivos com adesão química (Clearfil SE Bond, Scotchbond Universal, Allbond Universal).

3- A utilização de uma camada adicional de resina hidrofóbica melhorou o desempenho dos adesivos universais em termos de resistência de união resina-dentina quando usado com a estratégia AC. A camada adicional de resina hidrofóbica também melhorou o grau de conversão, tanto para a estratégia convencional como para a autocondicionante. A redução da nanoinfiltração foi mais dependente da composição do material que da estratégia adesiva. Apesar das melhorias *in vitro* observadas para adesivos universais, estudos de longevidade devem ser realizados para validar esses resultados iniciais.

REFERÊNCIAS¹

1. Demarco FF, Correa MB, Cenci MS, Moraes RR, Opdam NJ. Longevity of posterior composite restorations: not only a matter of materials. *Dental materials*. 2012 Jan;28(1):87-101.
2. Spencer P, Ye Q, Park J, Topp EM, Misra A, Marangos O, et al. Adhesive/Dentin interface: the weak link in the composite restoration. *Annals of biomedical engineering*. 2010 Jun;38(6):1989-2003.
3. Perdigao J, Reis A, Loguercio AD. Dentin adhesion and MMPs: a comprehensive review. *Journal of esthetic and restorative dentistry*. 2013 Aug;25(4):219-41.
4. Tjaderhane L, Nascimento FD, Breschi L, Mazzoni A, Tersariol IL, Geraldeli S, et al. Strategies to prevent hydrolytic degradation of the hybrid layer-A review. *Dental materials*. 2013 Oct;29(10):999-1011.
5. Nakabayashi N, Kojima K, Masuhara E. The promotion of adhesion by the infiltration of monomers into tooth substrates. *Journal of biomedical materials research*. 1982 May;16(3):265-73.
6. Van Meerbeek B, Perdigao J, Lambrechts P, Vanherle G. The clinical performance of adhesives. *Journal of dentistry*. 1998 Jan;26(1):1-20.
7. De Munck J, Van Landuyt K, Peumans M, Poitevin A, Lambrechts P, Braem M, et al. A critical review of the durability of adhesion to tooth tissue: methods and results. *Journal of dental research*. 2005 Feb;84(2):118-32.
8. Van Meerbeek B, De Munck J, Yoshida Y, Inoue S, Vargas M, Vijay P, et al. Buonocore memorial lecture. Adhesion to enamel and dentin: current status and future challenges. *Operative dentistry*. 2003 May-Jun;28(3):215-35.
9. Yiu CK, Pashley EL, Hiraishi N, King NM, Goracci C, Ferrari M, et al. Solvent and water retention in dental adhesive blends after evaporation. *Biomaterials*. 2005 Dec;26(34):6863-72.
10. Van Meerbeek B, Inokoshi S, Braem M, Lambrechts P, Vanherle G. Morphological aspects of the resin-dentin interdiffusion zone with different dentin adhesive systems. *Journal of dental research*. 1992 Aug;71(8):1530-40.

¹ De acordo com as normas do Programa de Pós-graduação em Odontologia da UEPG, baseado no modelo Vancouver. Abreviaturas dos periódicos em conformidade com o Medline.

11. Spencer P, Swafford JR. Unprotected protein at the dentin-adhesive interface. *Quintessence international*. 1999 Jul;30(7):501-7.
12. Tay FR, Gwinnett JA, Wei SH. Micromorphological spectrum from overdrying to overwetting acid-conditioned dentin in water-free acetone-based, single-bottle primer/adhesives. *Dental materials*. 1996 Jul;12(4):236-44.
13. Reis A, Loguercio AD, Azevedo CL, de Carvalho RM, da Julio Singer M, Grande RH. Moisture spectrum of demineralized dentin for adhesive systems with different solvent bases. *The journal of adhesive dentistry*. 2003 Fall;5(3):183-92.
14. Sano H, Shono T, Takatsu T, Hosoda H. Microporous dentin zone beneath resin-impregnated layer. *Operative dentistry*. 1994 Mar-Apr;19(2):59-64.
15. Burrow MF, Satoh M, Tagami J. Dentin bond durability after three years using a dentin bonding agent with and without priming. *Dental materials*. 1996 Sep;12(5):302-7.
16. Hashimoto M, Ohno H, Kaga M, Endo K, Sano H, Oguchi H. In vivo degradation of resin-dentin bonds in humans over 1 to 3 years. *Journal of dental research*. 2000 Jun;79(6):1385-91.
17. Miyazaki M, Onose H, Moore BK. Analysis of the dentin-resin interface by use of laser Raman spectroscopy. *Dental materials*. 2002 Dec;18(8):576-80.
18. Van Meerbeek B, De Munck J, Mattar D, Van Landuyt K, Lambrechts P. Microtensile bond strengths of an etch&rinse and self-etch adhesive to enamel and dentin as a function of surface treatment. *Operative dentistry*. 2003 Sep-Oct;28(5):647-60.
19. Carvalho RM, Chersoni S, Frankenberger R, Pashley DH, Prati C, Tay FR. A challenge to the conventional wisdom that simultaneous etching and resin infiltration always occurs in self-etch adhesives. *Biomaterials*. 2005 Mar;26(9):1035-42.
20. Wang Y, Spencer P. Continuing etching of an all-in-one adhesive in wet dentin tubules. *Journal of dental research*. 2005 Apr;84(4):350-4.
21. Tay FR, Gwinnett JA, Wei SH. Relation between water content in acetone/alcohol-based primer and interfacial ultrastructure. *Journal of dentistry*. 1998 Mar;26(2):147-56.

22. Perdigao J. Dentin bonding-variables related to the clinical situation and the substrate treatment. *Dental materials*. 2010 Feb;26(2):e24-37.
23. Kanemura N, Sano H, Tagami J. Tensile bond strength to and SEM evaluation of ground and intact enamel surfaces. *Journal of dentistry*. 1999 Sep;27(7):523-30.
24. Pashley DH, Tay FR. Aggressiveness of contemporary self-etching adhesives. Part II: etching effects on unground enamel. *Dental materials*. 2001 Sep;17(5):430-44.
25. Rotta M, Bresciani P, Moura SK, Grande RH, Hilgert LA, Baratieri LN, et al. Effects of phosphoric acid pretreatment and substitution of bonding resin on bonding effectiveness of self-etching systems to enamel. *The journal of adhesive dentistry*. 2007 Dec;9(6):537-45.
26. Frankenberger R, Lohbauer U, Roggendorf MJ, Naumann M, Taschner M. Selective enamel etching reconsidered: better than etch-and-rinse and self-etch? *The journal of adhesive dentistry*. 2008 Oct;10(5):339-44.
27. Van Landuyt KL, Peumans M, De Munck J, Lambrechts P, Van Meerbeek B. Extension of a one-step self-etch adhesive into a multi-step adhesive. *Dental materials*. 2006 Jun;22(6):533-44.
28. Van Landuyt KL, Kanumilli P, De Munck J, Peumans M, Lambrechts P, Van Meerbeek B. Bond strength of a mild self-etch adhesive with and without prior acid-etching. *Journal of dentistry*. 2006 Jan;34(1):77-85.
29. Hanabusa M, Mine A, Kuboki T, Momoi Y, Van Ende A, Van Meerbeek B, et al. Bonding effectiveness of a new 'multi-mode' adhesive to enamel and dentine. *Journal of dentistry*. 2012 Feb 28;40(6):475-84.
30. Erhardt MC, Osorio E, Aguilera FS, Proenca JP, Osorio R, Toledano M. Influence of dentin acid-etching and NaOCl-treatment on bond strengths of self-etch adhesives. *American journal of dentistry*. 2008 Feb;21(1):44-8.
31. Proenca JP, Polido M, Osorio E, Erhardt MC, Aguilera FS, Garcia-Godoy F, et al. Dentin regional bond strength of self-etch and total-etch adhesive systems. *Dental materials*. 2007 Dec;23(12):1542-8.
32. Taschner M, Nato F, Mazzoni A, Frankenberger R, Falconi M, Petschelt A, et al. Influence of preliminary etching on the stability of bonds created by one-step self-etch bonding systems. *European journal of oral sciences*. 2012 Jun;120(3):239-48.

33. Perdigao J, Sezinando A, Monteiro PC. Laboratory bonding ability of a multi-purpose dentin adhesive. *American journal of dentistry*. 2012 Jun;25(3):153-58.
34. Liu Y, Tjaderhane L, Breschi L, Mazzoni A, Li N, Mao J, et al. Limitations in bonding to dentin and experimental strategies to prevent bond degradation. *Journal of dental research*. 2011 Aug;90(8):953-68.
35. Breschi L, Mazzoni A, Ruggeri A, Cadenaro M, Di Lenarda R, De Stefano Dorigo E. Dental adhesion review: aging and stability of the bonded interface. *Dental materials*. 2008 Jan;24(1):90-101.
36. Vaidyanathan TK, Vaidyanathan J. Recent advances in the theory and mechanism of adhesive resin bonding to dentin: a critical review. *Journal of biomedical materials research part B: applied biomaterials*. 2009 Feb;88(2):558-78.
37. Hashimoto M. A review--micromorphological evidence of degradation in resin-dentin bonds and potential preventional solutions. *Journal of biomedical materials research part B: applied biomaterials*. 2010 Jan;92(1):268-80.
38. Sano H. Microtensile testing, nanoleakage, and biodegradation of resin-dentin bonds. *Journal of dental research*. 2006 Jan;85(1):11-4.
39. Ye Q, Spencer P, Wang Y, Misra A. Relationship of solvent to the photopolymerization process, properties, and structure in model dentin adhesives. *Journal of biomedical materials research part A*. 2007 Feb;80(2):342-50.
40. Loguercio AD, Loeblein F, Cherobin T, Ogliari F, Piva E, Reis A. Effect of solvent removal on adhesive properties of simplified etch-and-rinse systems and on bond strengths to dry and wet dentin. *The journal of adhesive dentistry*. 2009 Jun;11(3):213-9.
41. Paul SJ, Leach M, Rueggeberg FA, Pashley DH. Effect of water content on the physical properties of model dentine primer and bonding resins. *Journal of dentistry*. 1999;27(3):209-14.
42. Cadenaro M, Antonioli F, Sauro S, Tay FR, Di Lenarda R, Prati C, et al. Degree of conversion and permeability of dental adhesives. *European journal of oral sciences*. 2005 Dec;113(6):525-30.
43. Hass V, Folkuenig MS, Reis A, Loguercio AD. Influence of Adhesive Properties on Resin-Dentin Bond Strength of One-step Self-etching Adhesives. *Journal of adhesive dentistry*. 2011 OCT 2011;13(5):417-24.

44. Reis A, Ferreira SQ, Costa TR, Klein-Junior CA, Meier MM, Loguercio AD. Effects of increased exposure times of simplified etch-and-rinse adhesives on the degradation of resin-dentin bonds and quality of the polymer network. *European journal of oral sciences*. 2010 Oct;118(5):502-9.
45. Takahashi H, Sato H, Uno S, Pereira PN, Sano H. Effects of mechanical properties of adhesive resins on bond strength to dentin. *Dental materials*. 2002;18(3):263-8.
46. Cho BH, Dickens SH. Effects of the acetone content of single solution dentin bonding agents on the adhesive layer thickness and the microtensile bond strength. *Dental materials*. 2004 Feb;20(2):107-15.
47. Malacarne J, Carvalho RM, de Goes MF, Svizero N, Pashley DH, Tay FR, et al. Water sorption/solubility of dental adhesive resins. *Dental materials*. 2006 Oct;22(10):973-80.
48. Ito S, Hashimoto M, Wadgaonkar B, Svizero N, Carvalho RM, Yiu C, et al. Effects of resin hydrophilicity on water sorption and changes in modulus of elasticity. *Biomaterials*. 2005 Nov;26(33):6449-59.
49. Breschi L, Mazzoni A, Ruggeri A, Cadenaro M, Di Lenarda R, De Stefano Dorigo E. Dental adhesion review: aging and stability of the bonded interface. *Dental materials*. 2008 Jan;24(1):90-101.
50. Wang Y, Spencer P. Hybridization efficiency of the adhesive/dentin interface with wet bonding. *Journal of dental research*. 2003 Feb;82(2):141-5.
51. Wang Y, Spencer P, Yao X. Micro-Raman imaging analysis of monomer/mineral distribution in intertubular region of adhesive/dentin interfaces. *Journal of biomedical optics*. 2006 Mar-Apr;11(2):024005.
52. Santini A, Miletic V. Quantitative micro-Raman assessment of dentine demineralization, adhesive penetration, and degree of conversion of three dentine bonding systems. *European journal of oral science*. 2008 Apr;116(2):177-83.
53. Sano H, Takatsu T, Ciucchi B, Horner JA, Matthews WG, Pashley DH. Nanoleakage: leakage within the hybrid layer. *Operative dentistry*. 1995 Jan-Feb;20(1):18-25.
54. De Munck J, Van Meerbeek B, Yoshida Y, Inoue S, Vargas M, Suzuki K, et al. Four-year water degradation of total-etch adhesives bonded to dentin. *Journal of dental research*. 2003 Feb;82(2):136-40.

55. Nishitani Y, Yoshiyama M, Wadgaonkar B, Breschi L, Mannello F, Mazzoni A, et al. Activation of gelatinolytic/collagenolytic activity in dentin by self-etching adhesives. *European journal of oral sciences*. 2006 Apr;114(2):160-6.
56. Pashley DH, Tay FR, Yiu C, Hashimoto M, Breschi L, Carvalho RM, et al. Collagen degradation by host-derived enzymes during aging. *Journal of dental research*. 2004 Mar;83(3):216-21.
57. Tersariol IL, Geraldeli S, Minciotti CL, Nascimento FD, Paakkonen V, Martins MT, et al. Cysteine cathepsins in human dentin-pulp complex. *Journal of endodontics*. 2010 Mar;36(3):475-81.
58. Nascimento FD, Minciotti CL, Geraldeli S, Carrilho MR, Pashley DH, Tay FR, et al. Cysteine cathepsins in human carious dentin. *Journal of dental research*. 2011 Apr;90(4):506-11.
59. Reis A, Carrilho M, Breschi L, Loguercio A. Overview of Clinical Alternatives to Minimize the Degradation of the Resin-dentin Bonds. *Operative dentistry*. 2013 Jul-Aug;38(4):E1-E25.
60. King NM, Tay FR, Pashley DH, Hashimoto M, Ito S, Brackett WW, et al. Conversion of one-step to two-step self-etch adhesives for improved efficacy and extended application. *American journal of dentistry*. 2005 Apr;18(2):126-34.
61. de Andrade e Silva SM, Carrilho MR, Marquezini Junior L, Garcia FC, Manso AP, Alves MC, et al. Effect of an additional hydrophilic versus hydrophobic coat on the quality of dentinal sealing provided by two-step etch-and-rinse adhesives. *Journal of applied oral science*. May-Jun;17(3):184-9.
62. Reis A, Albuquerque M, Pegoraro M, Mattei G, Bauer JR, Grande RH, et al. Can the durability of one-step self-etch adhesives be improved by double application or by an extra layer of hydrophobic resin? *Journal of dentistry*. 2008 May;36(5):309-15.
63. Reis A, Leite TM, Matte K, Michels R, Amaral RC, Geraldeli S, et al. Improving clinical retention of one-step self-etching adhesive systems with an additional hydrophobic adhesive layer. *Journal of the american dental association (1939)*. 2009 Jul;140(7):877-85.
64. Perdigao J, Geraldeli S, Carmo AR, Dutra HR. In vivo influence of residual moisture on microtensile bond strengths of one-bottle adhesives. *Journal of esthetic and restorative dentistry*. 2002;14(1):31-8.

65. Tay FR, Pashley DH, Suh BI, Carvalho RM, Itthagarun A. Single-step adhesives are permeable membranes. *Journal of dentistry*. 2002 Sep-Nov;30(7-8):371-82.
66. Inoue S, Koshiro K, Yoshida Y, De Munck J, Nagakane K, Suzuki K, et al. Hydrolytic stability of self-etch adhesives bonded to dentin. *Journal of dental research*. 2005 Dec;84(12):1160-4.
67. Waidyasekera K, Nikaido T, Weerasinghe DS, Ichinose S, Tagami J. Reinforcement of dentin in self-etch adhesive technology: a new concept. *Journal of dentistry*. 2009 Aug;37(8):604-9.
68. Peumans M, De Munck J, Van Landuyt KL, Poitevin A, Lambrechts P, Van Meerbeek B. Eight-year clinical evaluation of a 2-step self-etch adhesive with and without selective enamel etching. *Dental materials*. 2010 Dec;26(12):1176-84.
69. Yoshida Y, Yoshihara K, Nagaoka N, Hayakawa S, Torii Y, Ogawa T, et al. Self-assembled Nano-layering at the Adhesive Interface. *Journal of dental research*. 2012 Apr;91(4):376-81.
70. Yoshihara K, Yoshida Y, Nagaoka N, Fukegawa D, Hayakawa S, Mine A, et al. Nano-controlled molecular interaction at adhesive interfaces for hard tissue reconstruction. *Acta biomaterialia*. 2010 Sep;6(9):3573-82.
71. Toledano M, Osorio R, Osorio E, Aguilera FS, Yamauti M, Pashley DH, et al. Durability of resin-dentin bonds: effects of direct/indirect exposure and storage media. *Dental materials*. 2007 Jul;23(7):885-92.
72. Feitosa VP, Leme AA, Sauro S, Correr-Sobrinho L, Watson TF, Sinhorette MA, et al. Hydrolytic degradation of the resin-dentine interface induced by the simulated pulpal pressure, direct and indirect water ageing. *Journal of dentistry*. 2012 Sep 19.
73. Chen L, Suh BI. Effect of hydrophilicity on the compatibility between a dual-curing resin cement and one-bottle simplified adhesives. *The journal of adhesive dentistry*. 2013 Aug;15(4):325-31.
74. 3MESPE.
http://multimedia.3m.com/mws/mediawebserver?mwsId=SSSSSuUn_zu8l00xMY_eMYt1Nv70k17zHvu9lxtD7SSSSSS--, 2013.

75. Pleffken PR, de Almeida Lourenco AP, Torres CR, Buhler Borges A. Influence of application methods of self-etching adhesive systems on adhesive bond strength to dentin. *The journal of adhesive dentistry*. 2011 Dec;13(6):517-25.
76. Loguercio AD, Stanislawczuk R, Mena-Serrano A, Reis A. Effect of 3-year water storage on the performance of one-step self-etch adhesives applied actively on dentine. *Journal of dentistry*. 2011 Aug;39(8):578-87.
77. Zhang Y, Wang Y. Effect of application mode on interfacial morphology and chemistry between dentine and self-etch adhesives. *Journal of dentistry*. 2013 Mar;41(3):231-40.
78. do Amaral RC, Stanislawczuk R, Zander-Grande C, Michel MD, Reis A, Loguercio AD. Active application improves the bonding performance of self-etch adhesives to dentin. *Journal of dentistry*. 2009 Jan;37(1):82-90.
79. do Amaral RC, Stanislawczuk R, Zander-Grande C, Gagler D, Reis A, Loguercio AD. Bond strength and quality of the hybrid layer of one-step self-etch adhesives applied with agitation on dentin. *Operative dentistry*. 2010 Mar-Apr;35(2):211-9.
80. Miyazaki M, Platt JA, Onose H, Moore BK. Influence of dentin primer application methods on dentin bond strength. *Operative dentistry*. 1996 Jul-Aug;21(4):167-72.
81. Zhang Y, Wang Y. Distinct photopolymerization efficacy on dentin of self-etch adhesives. *Journal of dental research*. 2012 Aug;91(8):795-9.
82. Frankenberger R, Perdigao J, Rosa BT, Lopes M. "No-bottle" vs "multi-bottle" dentin adhesives--a microtensile bond strength and morphological study. *Dental materials*. 2001 Sep;17(5):373-80.
83. Albuquerque M, Pegoraro M, Mattei G, Reis A, Loguercio AD. Effect of double-application or the application of a hydrophobic layer for improved efficacy of one-step self-etch systems in enamel and dentin. *Operative dentistry*. 2008 Sep-Oct;33(5):564-70.
84. Loguercio AD, Reis A. Application of a dental adhesive using the self-etch and etch-and-rinse approaches: an 18-month clinical evaluation. *Journal of the American Dental Association*. 2008 Jan;139(1):53-61.
85. Perdigao J. New developments in dental adhesion. *Dental clinics of north america*. 2007 Apr;51(2):333-57, viii.

86. Nunes TG, Ceballos L, Osorio R, Toledano M. Spatially resolved photopolymerization kinetics and oxygen inhibition in dental adhesives. *Biomaterials*. 2005 May;26(14):1809-17.
87. Van Meerbeek B, Yoshihara K, Yoshida Y, Mine A, De Munck J, Van Landuyt KL. State of the art of self-etch adhesives. *Dental materials*. 2011 Jan;27(1):17-28.
88. Perdigao J, Sezinando A, Monteiro PC. Effect of substrate age and adhesive composition on dentin bonding. *Operative dentistry*. 2013 May-Jun;38(3):267-74.
89. Tay FR, Pashley DH. Aggressiveness of contemporary self-etching systems. I: Depth of penetration beyond dentin smear layers. *Dental materials*. 2001 Jul;17(4):296-308.
90. Kenshima S, Reis A, Uceda-Gomez N, Tancredo Lde L, Filho LE, Nogueira FN, et al. Effect of smear layer thickness and pH of self-etching adhesive systems on the bond strength and gap formation to dentin. *The journal of adhesive dentistry*. 2005 Summer;7(2):117-26.
91. Van Meerbeek B, Conn LJ, Jr., Duke ES, Eick JD, Robinson SJ, Guerrero D. Correlative transmission electron microscopy examination of nondemineralized and demineralized resin-dentin interfaces formed by two dentin adhesive systems. *Journal of dental research*. 1996 Mar;75(3):879-88.
92. Spencer P, Wang Y, Walker MP, Wieliczka DM, Swafford JR. Interfacial chemistry of the dentin/adhesive bond. *Journal of dental research*. 2000 Jul;79(7):1458-63.
93. Munoz MA, Luque I, Hass V, Reis A, Loguercio AD, Bombarda NH. Immediate bonding properties of universal adhesives to dentine. *Journal of dentistry*. 2013 May;41(5):404-11.
94. Brackett MG, Li N, Brackett WW, Sword RJ, Qi YP, Niu LN, et al. The critical barrier to progress in dentine bonding with the etch-and-rinse technique. *Journal of dentistry*. 2011 Mar;39(3):238-48.
95. Wang Y, Spencer P. Quantifying adhesive penetration in adhesive/dentin interface using confocal Raman microspectroscopy. *Journal of biomedical materials research*. 2002 Jan;59(1):46-55.
96. Breschi L, Cadenaro M, Antonioli F, Sauro S, Biasotto M, Prati C, et al. Polymerization kinetics of dental adhesives cured with LED: correlation between extent of conversion and permeability. *Dental materials*. 2007 Sep;23(9):1066-72.

97. Reis A, Grande RH, Oliveira GM, Lopes GC, Loguercio AD. A 2-year evaluation of moisture on microtensile bond strength and nanoleakage. *Dental materials*. 2007 Jul;23(7):862-70.
98. Reis A, de Carvalho Cardoso P, Vieira LC, Baratieri LN, Grande RH, Loguercio AD. Effect of prolonged application times on the durability of resin-dentin bonds. *Dental materials*. 2008 May;24(5):639-44.
99. Tay FR, Pashley DH, Yoshiyama M. Two modes of nanoleakage expression in single-step adhesives. *Journal of dental research*. 2002 Jul;81(7):472-6.
100. Sadek FT, Calheiros FC, Cardoso PE, Kawano Y, Tay F, Ferrari M. Early and 24-hour bond strength and degree of conversion of etch-and-rinse and self-etch adhesives. *American journal of dentistry*. 2008 Feb;21(1):30-4.
101. Hamouda IM, Al-Khodary AM, El Shami FM. Degree of conversion and antimicrobial activity of etch-and-rinse versus self-etching adhesives. *The journal of adhesive dentistry*. 2010 Feb;12(1):33-8.
102. Sauro S, Mannocci F, Toledano M, Osorio R, Thompson I, Watson TF. Influence of the hydrostatic pulpal pressure on droplets formation in current etch-and-rinse and self-etch adhesives: a video rate/TSM microscopy and fluid filtration study. *Dental materials*. 2009 Nov;25(11):1392-402.
103. Tay FR, King NM, Chan KM, Pashley DH. How can nanoleakage occur in self-etching adhesive systems that demineralize and infiltrate simultaneously? *The journal of adhesive dentistry*. 2002 Winter;4(4):255-69.
104. Van Landuyt KL, Snauwaert J, Peumans M, De Munck J, Lambrechts P, Van Meerbeek B. The role of HEMA in one-step self-etch adhesives. *Dental materials*. 2008 Oct;24(10):1412-9.
105. Tay FR, Pashley DH. Have dentin adhesives become too hydrophilic? *Journal canadian dental association*. 2003 Dec;69(11):726-31.
106. Chersoni S, Suppa P, Breschi L, Ferrari M, Tay FR, Pashley DH, et al. Water movement in the hybrid layer after different dentin treatments. *Dental materials*. 2004 Nov;20(9):796-803.
107. Tay FR, Pashley DH, Suh B, Carvalho R, Miller M. Single-step, self-etch adhesives behave as permeable membranes after polymerization. Part I. Bond strength and morphologic evidence. *American journal of dentistry*. 2004 Aug;17(4):271-8.

108. Tay FR, Pashley DH, Garcia-Godoy F, Yiu CK. Single-step, self-etch adhesives behave as permeable membranes after polymerization. Part II. Silver tracer penetration evidence. *American journal of dentistry*. 2004 Oct;17(5):315-22.
109. Tay FR, Lai CN, Chersoni S, Pashley DH, Mak YF, Suppa P, et al. Osmotic blistering in enamel bonded with one-step self-etch adhesives. *Journal of dental research*. 2004 Apr;83(4):290-5.
110. Salz U, Zimmermann J, Zeuner F, Moszner N. Hydrolytic stability of self-etching adhesive systems. *The journal of adhesive dentistry*. 2005 Summer;7(2):107-16.
111. Hosaka K, Nakajima M, Takahashi M, Itoh S, Ikeda M, Tagami J, et al. Relationship between mechanical properties of one-step self-etch adhesives and water sorption. *Dental materials*. 2010 Apr;26(4):360-7.
112. Takahashi M, Nakajima M, Hosaka K, Ikeda M, Foxton RM, Tagami J. Long-term evaluation of water sorption and ultimate tensile strength of HEMA-containing/-free one-step self-etch adhesives. *Journal of dentistry*. 2011 Jul;39(7):506-12.
113. Van Landuyt KL, De Munck J, Snauwaert J, Coutinho E, Poitevin A, Yoshida Y, et al. Monomer-solvent phase separation in one-step self-etch adhesives. *Journal of dental research*. 2005 Feb;84(2):183-8.
114. Moszner N, Salz U, Zimmermann J. Chemical aspects of self-etching enamel-dentin adhesives: a systematic review. *Dental materials*. 2005 Oct;21(10):895-910.
115. Goracci C, Margvelashvili M, Apicella D, Sedda M, Magni E, Ferrari M. Influence of resin composite mechanical properties on adhesive microtensile bond strength to dentin. *The journal of adhesive dentistry*. 2011 Aug;13(4):323-31.
116. Takahashi A, Sato Y, Uno S, Pereira PN, Sano H. Effects of mechanical properties of adhesive resins on bond strength to dentin. *Dental materials*. 2002 May;18(3):263-8.
117. Reis A, Grandi V, Carlotto L, Bortoli G, Patzlaff R, Rodrigues Accorinte Mde L, et al. Effect of smear layer thickness and acidity of self-etching solutions on early and long-term bond strength to dentin. *Journal of dentistry*. 2005 Aug;33(7):549-59.
118. Loguercio AD, Uceda-Gomez N, Carrilho MR, Reis A. Influence of specimen size and regional variation on long-term resin-dentin bond strength. *Dental materials*. 2005 Mar;21(3):224-31.

119. Poitevin A, De Munck J, Van Landuyt K, Coutinho E, Peumans M, Lambrechts P, et al. Critical analysis of the influence of different parameters on the microtensile bond strength of adhesives to dentin. *The journal of adhesive dentistry*. 2008 Feb;10(1):7-16.

PARECER CONSUBSTANCIADO DO CEP

DADOS DO PROJETO DE PESQUISA

Título da Pesquisa: EFEITO DE DIFERENTES PROTOCOLOS DE APLICAÇÃO DE ADESIVOS UNIVERSAIS NAS PROPRIEDADES E LONGEVIDADE DA UNIÃO À DENTINA

Pesquisador: Nara Hellen Campanha Bombarda

Área Temática:

Versão: 4

CAAE: 23656513.0.0000.0105

Instituição Proponente: Universidade Estadual de Ponta Grossa

Patrocinador Principal: Financiamento Próprio

DADOS DO PARECER

Número do Parecer: 631.282

Data da Relatoria: 15/04/2014

Apresentação do Projeto:

Vários fatores podem afetar a longevidade de restaurações, entretanto, a cárie adjacente as margens das restaurações ainda é um dos maiores fatores que ocasionam a troca das restaurações.(Demarco et al.2012) Spencer et al.(Spencer et al.2 2010) indicam que, esta falha deve estar relacionada a falta de selamento da interface dente/restauração, o que faz com que haja falta de resistência desta interface aos agentes internos (enzimas proteolíticas) e externos (fluidos e microorganismos, entre outros) que possam ocasionar o início da falha.(Perdigão et al.3 2013; Tjaderhane et al.4 2013) Esta interface adesiva dente/restauração se conforma pela aplicação e interação de um sistema adesivo sobre os tecidos dentários tendo como mecanismo de união basicamente a substituição de minerais removidos dos tecidos dentários duros por monômeros resinosos, de tal modo que se transforma em um polímero interligado micromecanicamente com o substrato dental.(Nakabayashi et al.5 1982)Os atuais sistemas adesivos presentes no mercado podem ser classificados dentro de duas categorias segundo sua estratégia de aplicação clínica: a primeira, denominada de convencionais (CON) ou de condicionamento ácido (etch-andrinse no idioma Inglês) e a segunda de autocondicionantes (AC) (self-etch no idioma Inglês.Reis et al. (Reis et al.59 2013) indicaram diferentes alternativas clínicas que podem ser usadas para diminuir a degradação da interface de união a dentina, e assim,

Endereço: Av. Gen. Carlos Cavalcanti, nº 4748 bl M sala 12

Bairro:

CEP: 84.030-900

UF: PR

Município: PONTA GROSSA

Telefone: (42-)3220-3108

Fax: (42-)3220-3102

E-mail: seccoep@uepg.br

Continuação do Parecer: 631.282

aumentar a longevidade das restaurações. Dentre as técnicas discutidas se inclui a aplicação de uma camada adicional de resina hidrofóbica sobre o adesivo simplificado. (King et al.63 2005) Esta camada extra de resina tem como objetivo aumentar a espessura e uniformidade da interface adesiva, bem como reduzir o fluxo de fluido através dela. (king et al.63 2005, Andrade e Silva et al.64 2009) Excelente resultados in vitro e clínicos tem sido mostrados após a colocação de um revestimento de resina hidrofóbica sobre um adesivos AC de 1- passo (Reis et al.65 2008, Reis et al.66 2009), no entanto esta técnica não tem sido testado com os novos adesivos universais ou multi-mode.

Objetivo da Pesquisa:

O proposito deste estudo será avaliar o efeito nas propriedades adesivas da união a dentina de diferentes sistemas adesivos Universais aplicados segundo as estratégias adesivas convencional e autocondicionante, e modificadas com a aplicação de uma camada hidrófoba adicional.

-Objetivos específicos

1. Avaliar a resistência de união a microtração, nanoinfiltração, grau de conversão in situ de adesivos Universais aplicados em dentina de acordo as estratégias adesivas convencional e autocondicionante no tempo imediato.
2. Avaliar a resistência de união a microtração, nanoinfiltração de adesivos Universais aplicados em dentina de acordo as estratégias adesivas convencional e autocondicionante nos tempos imediato e 6 meses.
3. Avaliar a resistência de união à microtração, nanoinfiltração e grau de conversão in situ de adesivos Universais aplicados com uma camada hidrofóbica em dentina para as estratégias adesivas convencional e autocondicionante no tempo imediato.

Avaliação dos Riscos e Benefícios:

Riscos:

Não haverá riscos, pois serão utilizados dentes obtidos a partir do banco de dentes, para os pesquisadores também não haverá riscos, pois serão utilizados todos os EPIs necessários.

Benefícios:

Melhorar as propriedades de sistemas adesivos universais com o uso de protocolos alternativos à dentina(camada hidrofobica), se rejeitada a hipótese nula desse estudo, o que será de grande beneficio para a prática clinica.

Endereço: Av. Gen. Carlos Cavalcanti, nº 4748 bl M sala 12

Bairro:

CEP: 84.030-900

UF: PR

Município: PONTA GROSSA

Telefone: (42-)3220-3108

Fax: (42-)3220-3102

E-mail: seccoep@uepg.br

Continuação do Parecer: 631.282

Comentários e Considerações sobre a Pesquisa:

Cento e quarenta molares humanos livres de caries serão necessários para o desenvolvimento total da pesquisa. Serão utilizados 7 tipos de materiais adesivos distribuídos nos três experimentos desenvolvidos. Trata-se, portanto, de um trabalho experimental dividido em três partes:

1. Avaliação das propriedades imediatas de união de adesivos universais a dentina (40 dentes)
2. Avaliação das longevidade das propriedades de união de adesivos universais a dentina (40 dentes)
3. Influência de uma camada de resina hidrofóbica na performance adesiva de adesivos universais a dentina (60 dentes)

A metodologia é compatível aos objetivos

Considerações sobre os Termos de apresentação obrigatória:

Como se trata de estudo em dentes humanos, não há a necessidade de TCLE.

O aceite do Banco de Dentes da UEPG foi anexado

Os trabalhos in vitro da pós-graduação em Odontologia são autorizados diretamente pela Coordenação da pós-graduação (conforme justificativa anexada).

Folha de rosto preenchida adequadamente.

Recomendações:

Foi solicitado ao pesquisador alterar o grupo temático da pesquisa, tendo em vista que não há necessidade de parecer da CONEP, este projeto não é de área temática especial: Novos procedimentos terapêuticos invasivos.

Para o relator, o grupo aparece em branco.

Conclusões ou Pendências e Lista de Inadequações:

Sem pendências do ponto de vista ético.

Necessita confirmação do ajuste do grupo temático.

Situação do Parecer:

Aprovado

Necessita Apreciação da CONEP:

Não

Considerações Finais a critério do CEP:

Endereço: Av. Gen. Carlos Cavalcanti, nº 4748 bl M sala 12

Bairro:

CEP: 84.030-900

UF: PR

Município: PONTA GROSSA

Telefone: (42-)3220-3108

Fax: (42-)3220-3102

E-mail: seccoep@uepg.br

Continuação do Parecer: 631.282

PONTA GROSSA, 30 de Abril de 2014

Assinador por:
ULISSES COELHO
(Coordenador)

Endereço: Av. Gen. Carlos Cavalcanti, nº 4748 bl M sala 12

Bairro:

CEP: 84.030-900

UF: PR

Município: PONTA GROSSA

Telefone: (42-)3220-3108

Fax: (42-)3220-3102

E-mail: seccoep@uepg.br